1	Review article: Terrestrial dissolved organic carbon in northern permafrost
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22 Abstract

23 As the permafrost region warms and permafrost soils thaw, vast stores of soil organic 24 carbon (C) become vulnerable to enhanced microbial decomposition and lateral transport into 25 aquatic ecosystems as dissolved organic carbon (DOC). The mobilization of permafrost soil C 26 can drastically alter the net northern permafrost C budget. DOC entering aquatic ecosystems 27 becomes biological available for degradation as well as other types of aquatic processing. 28 However, it currently remains unclear which landscape characteristics are most relevant to 29 consider in terms of predicting DOC concentrations entering aquatic systems from permafrost regions. Here, we conducted a systematic review of 111 studies relating to, or including, 30 31 concentrations of DOC in terrestrial permafrost ecosystems in the northern circumpolar region 32 published between 2000 – 2022. We present a new permafrost DOC dataset consisting of 2,276 33 DOC concentrations, collected from the top 3 m in permafrost soils across the northern 34 circumpolar region. Concentrations of DOC ranged from $0.1 - 500 \text{ mg L}^{-1}$ (median = 41 mg L⁻¹) 35 across all permafrost zones, ecoregions, soil types, and thermal horizons. Across permafrost 36 zones the highest median DOC concentrations were in the sporadic permafrost zone (101 mg L^{-1}) while lower concentrations were found in the discontinuous (60 mg L^{-1}) and continuous (59 mg 37 38 L^{-1}) permafrost zones. However, median DOC concentrations varied in these zones across 39 ecosystem type, with the highest median DOC concentrations in each ecosystem type of 66 mg 40 L^{-1} and 63 mg L^{-1} found in coastal tundra and permafrost bog ecosystems, respectively. Coastal tundra (130 mg L^{-1}), permafrost bogs (78 mg L^{-1}), and permafrost wetlands (57 mg L^{-1}) had the 41 42 highest median DOC concentrations in the permafrost lens, representing a potentially long-term 43 store of DOC. Other than in Yedoma ecosystems, DOC concentrations were found to increase 44 following permafrost thaw and were highly constrained by total dissolved nitrogen 45 concentrations. This systematic review highlights how DOC concentrations differ between 46 organic- or mineral-rich deposits across the circumpolar permafrost region and identifies coastal 47 tundra regions as areas of potentially important DOC mobilization. The quantity of permafrost-48 derived DOC exported laterally to aquatic ecosystems is an important step for predicting its 49 vulnerability to decomposition.

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1. Introduction

52 Persistent freezing temperatures since the late Pleistocene and Holocene has led to the 53 accumulation and preservation of 1,460 – 1,600 Pg of organic carbon (C) in northern 54 circumpolar permafrost soils (Hugelius et al., 2014; Schuur et al., 2018). However, in recent 55 decades, there has been an amplified level of warming at high latitudes, occurring at four-times 56 the speed of the global average (Rantanen et al., 2021). This is leading to widespread and rapid 57 permafrost thawing which is predicted to continue under various future climate scenarios 58 (Olefeldt et al., 2016). Under the high C emissions representative concentration pathway 59 (RCP8.5), 90% loss of near-surface permafrost is projected to occur by 2300, with the majority 60 of loss occurring by 2100 (McGuire et al., 2018). Increasing temperatures and widespread thaw 61 exposes permafrost C to heterotrophic decomposition, potentially leading to enhanced emissions 62 of greenhouse gases to the atmosphere in the form of carbon dioxide (CO₂; Schuur et al., 2021) 63 and methane (CH₄; Turetsky et al., 2020). Additionally, previously frozen soil organic carbon 64 may be mobilized into the aquatic network as dissolved organic carbon (DOC), the quantity and 65 quality of which will likely depend on local and regional hydrology, and landscape 66 characteristics (Tank et al., 2012; Vonk et al., 2015). At high latitudes (>50°N), lakes and rivers 67 of various sizes cover 5.6% and 0.47% of the total area, respectively (Olefeldt et al., 2021), and 68 the landscape C balance at these high latitudes is highly dependent on aquatic C processing 69 (Vonk & Gustafsson, 2013). The increased leaching of recently thawed DOC from permafrost soils will increase the currently estimated 25 - 36 Tg DOC year⁻¹ exported into the freshwater 70 71 system, and subsequently into the Arctic Ocean (Holmes et al., 2012; Raymond et al., 2007). It 72 may also lead to enhanced greenhouse gas emissions from freshwater ecosystems (Dean et al., 73 2020). However, uncertainty remains as to which terrestrial ecosystems contain the highest 74 concentrations of DOC, laterally transport the greatest quantities of DOC, and represent the store 75 of DOC most vulnerable to mineralization.

Globally, DOC concentrations have been shown to vary across biomes, and spatial and
temporal scales (Guo et al., 2020; Langeveld et al., 2020). It has been suggested that at such
macro scales hydrology, climate, vegetation type, and soil type are important drivers of DOC
concentrations (Langeveld et al., 2020). Hydrology and climate are important factors shaping
ecosystem structure and function in permafrost regions (Andresen et al., 2020; Wang et al.,

81 2019), which in turn influences the spatial distribution of vegetation and soil types. Vegetation 82 type has been shown to be the most important driver of DOC concentrations in Arctic lakes 83 (Stolpmann et al., 2021). Carbon uptake by vegetation, via gross primary production, and SOC 84 stocks in the permafrost region have both been shown to vary across vegetation and soil types 85 (Ma et al., 2023; Hugelius et al., 2014). This variability across vegetation and soil types has 86 important implications for DOC production, which is associated with plant inputs (Moore & 87 Dalva, 2001) and the decomposition and solubilization of SOC due to soil microbial activity 88 (Guggenberger & Zech, 1993). In permafrost soils, the majority of this production is likely to 89 occur near the soil surface as the microbial production of DOC via input of plant-derived labile 90 substrates has been shown to decrease with depth (Hultman et al., 2015; Monteux et al., 2018; 91 Wild et al., 2016) and 65 - 70 % of the SOC store is found in the top 3 m (Hugelius et al., 2014). 92 The spatial distribution discrepancies observed in DOC concentrations from global assessment 93 efforts (Guo et al., 2020; Langeveld et al., 2020) may be reduced for the circumpolar permafrost 94 region by improving understanding of DOC concentrations in the top 3 m across ecosystem 95 types.

96 Previous studies have highlighted that the mineralization and lateral transport of DOC, i.e., 97 mobilization, represents a source of terrestrial permafrost C that can potentially play an 98 important role in both terrestrial and aquatic biogeochemical cycles (Hugelius et al., 2020; 99 Parmentier et al., 2017; Schuur et al., 2022). However, none have quantified DOC mobilization 100 across the permafrost region. Inclusion of DOC mobilization in attempts to determine the 101 permafrost climate feedback (Schaefer et al., 2014), may reduce current uncertainty in the 102 magnitude and location of permafrost C losses (Miner et al., 2022), particularly as permafrost 103 thaws. Warming of near surface permafrost causes widespread thawing (Camill, 2005; Jorgenson 104 et al., 2006), which can lead to drastic changes in hydrology, vegetation, and soil carbon 105 dynamics (Liljedahl et al., 2016; Pries et al., 2012; Varner et al., 2022), thus impacting both 106 DOC production and mobilization. Several studies have demonstrated that DOC has the potential 107 to be rapidly degraded and mineralized following thermokarst formation (Burd et al., 2020; 108 Payandi-Rolland et al., 2020; Wickland et al., 2018), particularly in higher latitude ecosystems 109 (Ernakovich et al., 2017; Vonk et al., 2013). However, few have compared this lability across 110 ecosystems (Abbot et al., 2014; Fouche et al., 2020; Textor et al., 2019) and less have done so

111 across the permafrost region (Vonk et al., 2015). Determining the ecosystems with the greatest 112 store of DOC that is readily mineralized upon thermokarst formation represents a potentially 113 important step in reducing uncertainty in the permafrost climate feedback.

114 Here, we conduct a systematic review of the literature and compiled 111 studies published 115 between 2000 – 2022 on DOC concentrations in the top 3 m of soil in terrestrial ecosystems 116 found in the northern circumpolar permafrost region. Our aim was to build a database to assess 117 the concentration and mobilization of DOC across terrestrial permafrost ecosystems. We used 118 this database to address the following hypotheses; (i) the highest DOC concentrations would be 119 found in organic rich wetland ecosystems; (ii) disturbance would lead to increased export and 120 biodegradability of DOC; and (iii) the most biodegradable DOC would be found in Yedoma and 121 tundra ecosystems. A quantitative assessment of studies pertaining to DOC concentrations in 122 permafrost soils can identify evidence-based recommendations for future topics, standardisation 123 of methods, and areas of research to improve our understanding on terrestrial and aquatic 124 biogeochemical cycling in northern permafrost regions. Our database contains ancillary data 125 describing the geographical and ecological conditions associated with each DOC concentration, 126 allowing us to reveal patterns in DOC concentrations and lability measures for 562 sampling 127 sites across multiple ecosystem types and under varying disturbance regimes. This study 128 represents the first systematic review of DOC concentrations within terrestrial permafrost 129 ecosystems found in the circumpolar north. As such, it provides unique and valuable insights into 130 identifying ecosystems associated with the highest DOC concentrations, and thus ecosystems 131 with the greatest potential for DOC mobilization.

132 **2. Methods**

133 This systematic review used a methodological framework proposed by Arksey & 134 O'Malley (2005) and follows five steps: 1) develop research questions and a search query; 2) 135 identify relevant studies; 3) study selection; 4) data extraction; and 5) data analysis, summary, 136 and reporting. The literature search was guided by four research questions: 1) what are the 137 concentrations of DOC found in terrestrial ecosystems across the northern circumpolar 138 permafrost region?; 2) what are the rates of export and/or degradation (mobilization) of DOC 139 within these ecosystems?; 3) What are the major controls on DOC concentrations and rates of 140 mobilization?; and 4) how are concentrations and mobilization rates impacted by thermokarst

141 formation? Mobilization rates represent DOC loss and include specific discharge of DOC (g

142 DOC m^{-2}), export rate of DOC per day (g C m^{-2} day⁻¹) and per year (g C m^{-2} year⁻¹), and

143 biodegradable DOC (BDOC; %).

144 2.1 Literature Search

145 Based on *a priori* tests, we used the following search query string to find papers using 146 information found in their title, abstract, and keywords: ("dissolved organic carbon") AND 147 (permafrost OR thermokarst OR "thaw slump") AND (soil OR peat) AND (export OR degrad* 148 OR decomposition OR mineralization). We used Web of Science, Science Direct, Scopus, 149 PubMed, and Google Scholar to generate a database of tier 1, peer-reviewed articles published 150 between 2000 - 2022. The search function on Science Direct does not support the use of 151 wildcards such as "*", so "degrad*" was changed to "degradation". We removed duplicate 152 references found across multiple databases using Mendeley[©] referencing software (v1.17.1,

153 Mendeley Ltd. 2016).

154 2.2 Systematic Screening of Peer-Reviewed Publications

155 The selection of relevant studies was comprised of inclusion criteria and relevance screening in three steps. In the first step we placed limits on initial study searches in the 156 157 electronic databases mentioned above. Studies were included in the review if they were primary 158 research, published in English, and published between 2000 - 2022 (Table 1). Only quantitative 159 studies conducted in terrestrial ecosystems within the northern circumpolar permafrost region, as 160 defined by Brown et al., (1997), and reporting DOC concentration and mobilization rates were 161 included. Studies not meeting these criteria were eliminated and the remaining studies proceeded 162 to the second screening step.

Table 1. Summary of criteria used to identify suitable studies in the preliminary screening stage

	Inclusion criteria	Exclusion criteria
Timeline	Study published between 2000 – 2022	Study published prior to 2000

Study type	Primary research article published in peer-reviewed journal using quantitative methods	Thesis/dissertations and secondary research studies (reviews, commentaries, editorials)
Language	Published in English	Studies published in other languages
Region Conducted within the northern circumpolar permafrost region		Conducted outside of the northern circumpolar permafrost region
Outcome	Studies on DOC concentration, export or degradation in permafrost environments	Studies not on DOC concentration, export or degradation in permafrost environments

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- 164 In the second step, the primary relevance of articles was screened, based on article titles,
- abstracts, and keywords, and the eligibility criteria provided in Table 2. Studies deemed

166 irrelevant were eliminated and the remaining studies proceeded to the third and final screening

167 step, or secondary screening stage, which was based on was based on more specific eligibility

168 criteria (Table 2) applied to the full text.

Table 2. Primary and secondary relevance screening tools. Primary screening tool used in the article title, abstract, and keyword screening stage. Secondary screening tool used in full-text screening stage

Screening stage	Screening questions	Response details
Primary	Does the study involve quantitative data collected from a	Yes – reports on quantitative data collected from a permafrost environment
	permafrost environment?	No – does not report on the above
Primary and Secondary	Is the study region within the northern circumpolar permafrost region?	Yes – reports on quantitative data (including field observations and lab data) collected from the circumpolar permafrost environment.
		No – study region is not in the northern circumpolar permafrost regions; other examples could be mountainous permafrost or Tibetan plateau
Primary and Secondary	Is the article in English and NOT a review, book chapter, commentary, correspondence,	Yes – study is in English and is a primary research article that includes quantitative studies (field and lab based), including model-based research as it relies on observational data.*

	letter, editorial, case report, or reflection?	No – study is not in English and/or is a review, book, editorial, working paper, commentary, conference proceeding, supplementary text, or qualitative study which does not address outcomes relevant to this review
Primary and Secondary	Does the study involve the concentration, export or degradation of terrestrially derived DOC?	Yes – reports on terrestrial DOC concentration, export, or degradation, including concentrations and characterization No – does not report on terrestrial DOC concentration, export, or degradation
Secondary	Is the article in English, longer than 500 words, and published between 2000 - 2022?	Yes – study is published between 2000 – 2022 No – study is published prior to 2000

169 *For model-based studies, the original field/lab data used to parametrise or develop the model

170 was used. If this data was taken from previously published work, then those studies were used

and the model-based study removed.

172 *2.3 Database compilation*

173 A database with reported DOC concentrations and mobilization rates i.e., rates of either 174 DOC export or degradation, was compiled using data from all studies that were deemed relevant 175 following the study selection phase. The database was compiled to compare DOC concentrations 176 and mobilization rates between different sites. We define a site as an area where either soil, 177 water, or ice samples were taken from that has similar vegetation composition, water table 178 position, permafrost regime, and was either disturbed or pristine. Site descriptions were derived 179 from the text of each study. Where possible, individual daily measurements of DOC 180 concentrations and mobilization rates were taken. When replicates of the same daily 181 measurement were provided, we used the mean of those replicates, which was relevant for 10 182 studies within the database, representing 72 DOC concentrations. All data was extracted from 183 data tables, text, supplementary material, or extracted from data figures using WebPlotDigitizer 184 (https://automeris.io/WebPlotDigitizer).

All studies reported measuring DOC concentrations collected from either open-water, pore
 water, ice, or soil using a median filter pore size of 0.45 μm with first and third quartiles pore
 size of 0.45 and 0.7 μm. Measurements from all 12 months of the year were included in the

188 database with the majority occurring during the growing season (May – August), a small portion 189 during the non-growing season, and the remaining sampling times were either not reported or are 190 averages over multiple sampling occasions. We included data from studies that were both field 191 and lab based. However, any data where a treatment was applied was excluded, except for 192 temperature treatments during incubation experiments when assessing the biodegradability of 193 DOC. When lab-based studies included an incubation, only Day 0 DOC concentrations were 194 used when comparing DOC concentrations across studies. We chose to remove any DOC 195 concentrations from samples taken below 3 m depth, which represented 3% of all DOC 196 measurements. These measurements were removed for better comparability with the current best 197 estimation of soil organic carbon stocks within the northern circumpolar permafrost zone 198 (Hugelius et al., 2014). We also removed any DOC concentrations greater than 500 mg L^{-1} , which represented 2% of all DOC concentrations. Samples that were above 500 mg L⁻¹ and were 199 200 sampled below 3 m represented 1% of all DOC concentrations.

201 Site averaged daily DOC concentrations (mg L^{-1}) and mobilization rates were estimated from 202 the average concentration and mobilization rates measured within a single day or sampling 203 occasion. Repeated measurements at a site, either over the growing season or multiyear 204 measurements, were treated as an individual estimate of DOC concentrations and mobilization 205 rates. Other continuous variables that were similarly estimated include soil moisture, water table 206 position, organic layer depth, active layer depth, bulk density of soil, soil carbon content (%), 207 soil nitrogen content (%), soil carbon:nitrogen (C:N), pH, electrical conductivity (µS cm⁻¹), specific UV absorbance at 254 nm (SUVA; L mg C⁻¹ m⁻¹), total dissolved nitrogen (mg L⁻¹), 208 nitrate (mg L⁻¹), ammonium (mg L⁻¹), chloride (mg L⁻¹), calcium (mg L⁻¹), and magnesium (mg 209 210 L^{-1}). The aromatic content of organic matter is positively correlated with SUVA (Weishaar et al., 211 2003), with high SUVA values being used as an indication of high aromatic content (Hansen et 212 al., 2016). Ratios of soil C:N have been shown to be a good proxy for decomposition (Biester et 213 al., 2014), where high C:N values indicate higher decomposition has previously occurred. Mean 214 annual temperatures and precipitation, sampling depth, filter size, the number of days over which 215 sampling took place, how many years following disturbance measurements were taken were also 216 recorded. Several continuous variables other than those mentioned above were also recorded in 217 the database, but not used for analysis if they represented < 20% of the database. We chose 20%

as the cut-off point for use in comparison of the relationship between DOC concentrations andmobilization with other site continuous variables.

220 Categorical variables included in the database (Table S1) were site location within the 221 permafrost zone (continuous, discontinuous, sporadic; Brown et al., 1997) and ecoregion (arctic 222 tundra, sub-arctic tundra, sub-arctic boreal, and continental boreal; Olson et al., 2001). We 223 included site surface permafrost conditions (present or absent), the thermal horizon layer 224 sampled (active layer, permafrost lens, permafrost free, water, and thaw stream), and if present 225 what type of disturbance occurred at the site (fire, active layer thickening, thermokarst terrestrial, 226 or thermokarst aquatic). Active layer represents the seasonally unfrozen soil layer above the 227 permafrost layer. Permafrost lens represents the permanently frozen (below 0 °C) layer. 228 Permafrost lens DOC concentrations are determined from soil and pore water within the 229 permafrost layer and extracted via frozen cores, whereas active layer samples are taken from soil 230 cores or porewater that are unfrozen at the time of sampling. Thaw Stream represents flowing 231 surface waters following permafrost thaw. Permafrost Free represents areas that are not underlain 232 by permafrost. We also included the soil class found at the site (Histel, Histosol, Orthel, and 233 Turbel; USDA, 1999) and whether the DOC was from the organic or mineral soil. Histosols are 234 organic rich, non-permafrost soils. Histels, Orthels, and Turbels are permafrost-affected soils 235 (Gelisol order). Histels are organic rich, Orthels are non cryoturbated affected mineral soils, and 236 Turbels are cryoturbated permafrost soils. Organic rich Histel and Histosol soils have been 237 previously shown to contain greater SOC stocks in the top 3 m of soil than the mineral rich 238 Orthel and Histel soils (Hugelius et al., 2014). To assess the influence of sampling approach and 239 method of analysis, we included method of DOC extraction (centrifugation of soil sample, 240 leaching and dry leaching of soil, dialysis, grab sample, ice core extraction, potassium sulphate 241 extraction, lysimeter, piezometer, pump, rhizons) and DOC measurement method (combustion, 242 persulphate, photometric, or solid-phase extraction).

Sites were classified according to ecosystem type, and these included coastal tundra, forest, peatland, permafrost bog, permafrost wetland, retrogressive thaw slump, upland tundra, and Yedoma. Ecosystem classification is based on the general site description in the article, the provided ecosystem classification within the article, and site data including vegetation composition, permafrost conditions, and ecoregion. Coastal tundra sites includes typical

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248 polygonal tundra features found along the coastline in the permafrost region (Lantuit et al., 249 2012). Forests include any forested ecosystem, such as a black spruce forest (Kane et al., 2006) 250 or larch forest (Kawahigashi et al., 2011) where the soil is not a wetland soil. Peatlands are sites 251 classified as either fens (Olefeldt and Roulet 2012) or bogs (Olsrund and Christensen 2011) that 252 are within the permafrost domain but are not underlain by permafrost. Permafrost bogs are sites 253 that are bogs and are either underlain by permafrost (O'Donnel et al., 2016) or are thermokarst 254 bogs (Burd et al., 2020) that were previously underlain by permafrost prior to thawing. 255 Permafrost wetlands sites include saturated soils that are underlain by permafrost, or were 256 previously underlain by permafrost prior to permafrost thaw. They contain sampling locations 257 typical of moist acidic tundra (Trusiak et al., 2018), tundra meadows (Tanski et al., 2017), and 258 high-latitude fens (Nielsen et al., 2017). Retrogressive thaw slumps are areas where substantial 259 ground ice degradation leads to thermokarst and the resulting feature contains a retreating 260 headwall (Abbott et al., 2015). Upland tundra sites are high-latitude, non-wetland, mineral soils 261 that include tundra heath (Stutter and Billett 2003) and meadows (Hirst et al., 2022). Yedoma 262 sites include pristine forest, upland tundra, and coastal tundra, as well as retrogressive thaw slumps and other thermokarst features found within the Yedoma permafrost domain (Strauss et 263 264 al., 2021). The ecosystem classification retrogressive thaw slump only includes these 265 thermokarst features found outside the Yedoma permafrost domain. Each ecosystem type was 266 further classified based on the type of permafrost thaw or thermokarst formation that occurred 267 there. These thaw or thermokarst types included thermokarst bog, thermokarst wetland, active 268 layer thickening, retrogressive thaw slump, exposure, thermo-erosion gully, and active layer 269 detachment.

270 2.4 Database analysis

All statistical analyses were carried out in R (Version 3.4.4, R Core Team, 2015). We aimed to assess how DOC concentrations differed across study regions and ecosystems. To do this we used Kruskal-Wallis analysis to test for differences in median DOC concentrations among the various study regions and areas that included permafrost zones, ecoregions, soil class, thermal horizon, and ecosystems. Post-hoc comparisons of median DOC concentrations among the categories were performed using pairwise Wilcox test. Within and between each ecosystem type we assessed the differences in DOC concentrations found in different thermal horizons (i.e., active layer and permafrost lens). To do this, data was first transformed using a Box Cox transformation and the optimal λ using the *MASS* package (Ripley et al., 2019). We then performed analysis of covariance (ANCOVA) to test for differences in DOC concentrations in different thermal horizons between ecosystem types, while controlling for seasonal effects by including the month in which sampling occurred as the covariate.

283 Following the assessment of differences in DOC concentrations across these study regions 284 and ecosystems we aimed to assess the influence of extraction and analysis method on DOC 285 concentrations. The aim of this was to determine if extraction and analysis method was having a 286 greater effect on DOC concentrations than study region or ecosystem. To do so we first used 287 ANOVAs and Bonferroni post-hoc tests on linear mixed effects models, that include either 288 extraction method, filter size, or analysis method as a fixed effect and ecosystem type as a 289 random factor, to evaluate significant differences in DOC concentrations between DOC 290 extraction and measurement methods. We then performed Kruskal-Wallis analysis to test for 291 differences in median DOC concentrations among the extraction method, filter size, and analysis 292 method in each permafrost zone, ecoregion, soil class, thermal horizon, and ecosystem. Post-hoc 293 comparisons of median DOC concentrations among these categories were performed using 294 pairwise Wilcox test.

295 We used partial least squares regression (PLS) when assessing the relationship of DOC 296 concentrations with continuous and categorical variables. We performed this analysis to 297 determine how the drivers of DOC concentrations across ecosystems may explain the variability 298 in DOC concentrations. Predictor variables were categorized based on their Variable Importance 299 in Projections (VIP) method in the *plsVarSel* package (Mehmood et al., 2012), whereby variables 300 with a score > 0.6 - 1 are deemed to be significant (Chong and Jun 2005). We ran several PLS 301 including predictor variables with a VIP of > 0.6, 0.7, 0.8, 0.9, and 1. The most parsimonious 302 PLS model contained predictor variables with a VIP > 1 and was selected based on the 303 proportion of variability in the predictors explained by the model, significant PLS components, 304 Q^2 , and background correlation (Andersen and Bro 2010). PLS was performed using the pls 305 package (Mevik & Wehrens, 2007) and we chose to use PLS as it is tolerant of co-correlation of 306 predictor variable, deviations from normality, and missing values, all of which were found within 307 the database. In the PLS ecosystem classes were subdivided into pristine or disturbed (i.e.,

impacted by permafrost thaw). Pristine sites were further subdivided by the thermal horizon in which the DOC concentrations were measured (active layer and permafrost lens). Sites were split into disturbed and pristine to assess whether disturbances has an impact on DOC concentrations. Pristine sites were divided by their thermal horizon to assess whether DOC concentrations were more positively related to the active layer exposed to both microbial decomposition and fresh annual carbon inputs from surface vegetation, or the permafrost lens.

314 To evaluate the change in ecosystem DOC concentrations following thermokarst formation, 315 based on all studies from the systematic review, we calculated the response ratio using the 316 SingleCaseES package (Pustejovsky et al., 2021). We define thermokarst as the process by which 317 ice-rich permafrost deposits undergo complete thaw, resulting in surface subsidence and the 318 formation of a new, thermokarst feature that is ecological different regarding water table 319 position, redox conditions, and vegetation type, from the preceding pristine ecosystem. Very few 320 studies in our database report DOC concentrations for both pristine and thermokarst affected 321 ecosystem (< 20 %). To include as much data as possible we chose an effect size metric that is 322 unlikely to be influenced by studies with large sample number and variance. The response ratio 323 is:

324 Pristine to Thermokarst Effect Response ratio =
$$\ln(\frac{X_P}{X_T})$$
 Eqn. 1

where X_P = mean DOC concertation of pristine ecosystems and X_T = mean DOC concertation of thermokarst effected ecosystems (Lajeunesse, 2011). This represents the log proportional difference in mean DOC concentrations between thermokarst and pristine ecosystems, where a positive response ratio indicates a decrease in DOC concentrations following thermokarst.

The distribution of the data was inspected visually and with the Shapiro–Wilk test. We tested homogeneity of variances using the *car* package and Levene's test (Fox and Weisberg, 2011). We report DOC concentrations as the median value with uncertainty as \pm the interquartile range, except for response ratios which we report as \pm 95% confidence intervals. We here define the statistical significance level at 5%.

334 3. Results

335 3.1 Database generation

336 Our initial search using Web of Knowledge, Science Direct, Scopus, PubMed, and 337 Google Scholar returned a total of 577 unique papers published between 2000 – 2022 that assess 338 the concentrations and rates of mobilization of DOC in terrestrial ecosystems within the northern 339 circumpolar permafrost region. Of these initial 577 studies, 111 remained after the systematic screening process (Table 1 & 2). From these 111 studies we generated our database. The final 340 database of 111 studies contained a total of 3,340 DOC concentrations (mg L⁻¹), with 2,845 DOC 341 concentrations between $0 - 500 \text{ mg L}^{-1}$, found within the top 3 m of permafrost soils from field 342 343 and lab-based studies (using only Day 0 lab-based DOC concentrations). These concentrations 344 were taken from 562 different sampling locations, representing 8 different ecosystem types 345 (Figure 1; Table S2) across the northern circumpolar permafrost region. All studies except, for 346 one (Olefeldt et al., 2012), reported DOC concentrations.

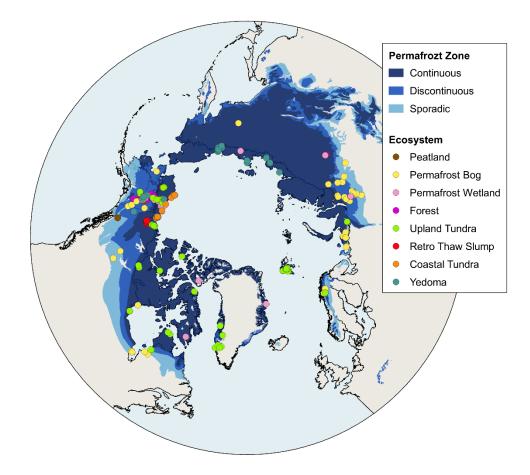


Figure 1. Map of sampling locations where DOC measurements (n = 562) from the top 3 m for each ecosystem type. In many cases, the same sampling location was used in multiple studies leading to some overlap, therefore the number of sampling sites included in the data set (562) are not all clearly identifiable from this map. Similarly, several points overlay others even when the ecosystems differ. For a full list of site coordinates please see the database (repository link). Retro Thaw Slump = Retrogressive Thaw Slump. Blue shading represents permafrost zonation (Brown et al., 1997).

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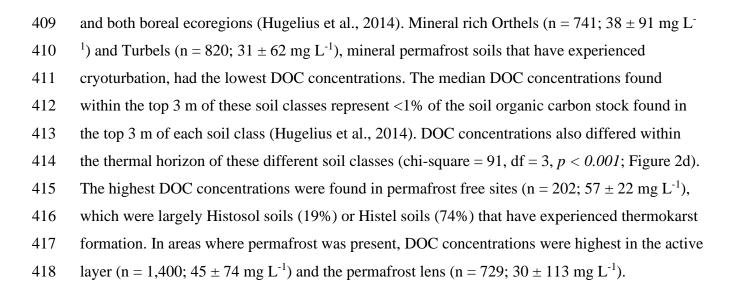
356 The final database contained a considerably lower number of DOC mobilization 357 measurements. The database includes 16 measurements of specific discharge of DOC (g DOC m⁻ ²) from 3 studies, 9 export rate of DOC per day (g C m⁻² day⁻¹) and per year (g C m⁻² year⁻¹) 358 359 measurements were each found in 2 studies. The number of specific discharge, export of DOC 360 per day, and export of DOC per year measurements combined were <1% of the number of DOC 361 concentration measurements. As such they were not considered for analysis of DOC 362 mobilization. A total of 146 BDOC (%) measurements, 4% of the total number of DOC concentration measurements, were found in 14 studies. These measurements of BDOC were 363 from Yedoma (30:5, number of measurements: studies), Upland Tundra (55:5), Forest (18:3), 364 365 Permafrost Wetland (12:2), and Permafrost Bog (31:5) ecosystems. Given the low number of 366 other forms of DOC mobilization and relatively comparable spread of BDOC measurements 367 across ecosystem types, we chose to include BDOC measurements in our analysis despite a low 368 total number of measurements compared to DOC concentrations, and we consider this lower 369 sample size during our interpretation of results.

370 Filter size used in studies ranged from $0.15 - 0.7 \mu m$. The majority of DOC 371 concentrations reported were determined using a filter size of 0.45 µm (58%), 0.7 µm was the 372 second most common filter size (21%), followed by $0.22 \mu m$ (14%). We identified eleven 373 different DOC extraction methods in total from both soils and water that are broadly grouped 374 into the following six extraction types; leaching, suction, grab, centrifuged, dialysis, and 375 potassium sulphate (K₂SO₄) extraction. Leaching includes the leaching and dry leaching of soil; 376 suction includes lysimeter, piezometer, pump, and rhizons; grab includes grab samples and ice 377 core extraction; and centrifuged, dialysis, and (K₂SO₄) extraction remain on their own. Suction 378 (42%), leaching (37%), and grab (14%) were the three most common extraction methods across 379 all samples. Leaching and suction extraction methods were used for 66% and 24%, respectively, for all soil samples. For water samples, suction (65%) and grab (31%) were the most common extraction methods. The most common measurement method to determine DOC concentrations was by the combustion method (89%), followed by the persulphate (9%) and photometric (1%) methods.

384 *3.2 DOC concentrations and study regions*

385 Upon inspection of DOC concentrations in the database, we determined that the data was 386 non-normally distributed. The DOC concentrations were skewed toward the lower end of our 0-500 mg L⁻¹ range; thus, we report median, upper, and lower quartiles below. Across all studies, 387 388 within the top 3 m of soil, the median DOC concentration was 41 ± 74 mg L⁻¹. DOC concentrations were found to differ among the three permafrost zones (chi-square = 32, df = 2, p389 390 < 0.001; Figure 2a). The highest median DOC concentrations were found within the sporadic permafrost zone (n = 83; 62 \pm 144 mg L⁻¹). The lowest median of 33 \pm 77 mg L⁻¹ was found in 391 392 the continuous permafrost zone (n = 1,648), with the greatest density of samples having lower 393 DOC concentrations than observed in the violin plots of both the discontinuous and sporadic 394 (Figure 2a). This change in DOC concertation's along the latitudinal gradient of the permafrost 395 zonation was also seen in the latitudinal gradient associated with ecoregion, where Arctic Tundra 396 and Sub-Arctic Tundra are found at higher latitudes than both boreal ecoregions (chi-square = 397 78, df = 3, p < 0.001; Figure 2b). The highest DOC concentrations were found in the continental 398 boreal (n = 389; 56 \pm 56 mg L⁻¹) and Sub-Arctic Boreal (n = 442; 58 \pm 97 mg L⁻¹) ecoregions, 399 and lowest in the Arctic Tundra (n = 1,209; $25 \pm 75 \text{ mg L}^{-1}$) and Sub-Arctic Tundra (n = 493; 43) \pm 61 mg L⁻¹) ecoregions. Inspection of the distribution of DOC concentrations across the 400 401 ecoregions highlights that the Arctic Tundra ecoregion had the highest density of samples at the 402 lowest DOC concentration (Figure 2b).

These latitudinal differences are also reflected in the observed differences (chi-square = 20, df = 3, p < 0.001) in DOC concentrations found within different soil classes. The highest DOC concentrations are found within organic rich Histosol (n = 37; 61 ± 39 mg L⁻¹) and Histel soils (n = 935; 53 ± 72 mg L⁻¹; Figure 2c), with the distribution of the data from these soils types having a higher density at greater DOC concentrations (Figure 2c). Histel and Histosol soils are the main type of permafrost soil found within the sporadic and discontinuous permafrost zone



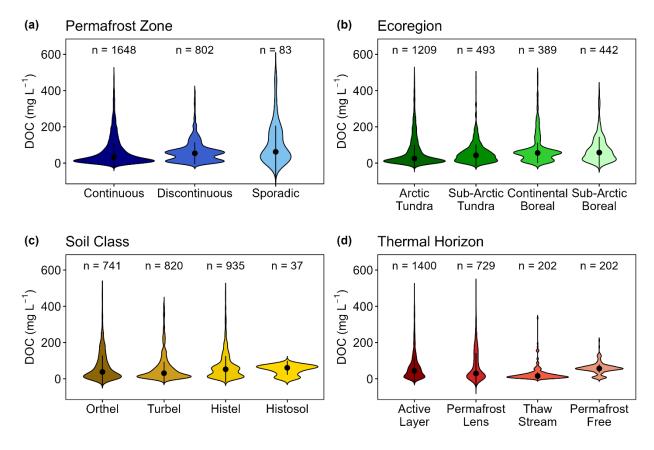


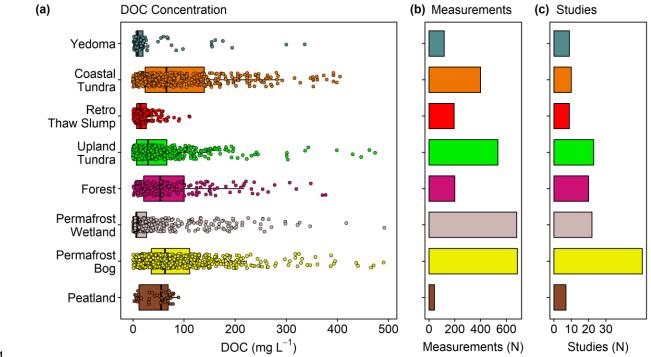
Figure 2. Violin plots of DOC concentrations (mg L⁻¹⁻) found in the top 3 m across (a) permafrost
zones, (b) ecoregions, (c) soil classes, and (d) thermal horizons. (a) Dark to light blue shading
represents the permafrost zones Continuous, Discontinuous, and Sporadic, according to Brown
et al., (1997). (b) Dark to light green shading represents the ecoregions Arctic Tundra, SubArctic Tundra, Continental Boreal, and Sub-Arctic Boreal, according to Olson et al., (2001). (c)
Dark to light yellow shading represents the soil classes Histosol, Histel, Orthel, and Turbel,

according to the USDA Soil Taxonomy (USDA, 1999). (d) Dark to light red shading represents 426 427 the thermal horizons Active Layer, Permafrost Lens, Thaw Stream, and Permafrost Free. Black 428 dots on each violin plot represents the median. Black vertical lines represent the interguartile 429 range with the upper and lower limits representing the 75th and 25th percentiles, respectively. 430 Either side of the black vertical line represents a kernel density estimation. This shape shows 431 the distribution of the data, with wider areas representing a higher probability that samples 432 within the database will have that DOC concentrations. The number of samples (n) found in 433 each sub-category is found above each corresponding violin plot.

- 434
- 435

3.3 Trends in DOC concentrations across ecosystems

436 Similar to other categorical variables (i.e. permafrost zone, ecoregion, soil class, and 437 thermal horizon data), DOC concentrations within each of the eight ecosystem types were found 438 to be non-normally distributed, with median values skewed toward the lower end of the 0-500mg L⁻¹ range of concentrations (Figure S1). Permafrost bogs, upland tundra, and permafrost 439 440 wetlands were the most represented in the database with regards to DOC concentrations (Table 441 S2). The majority of permafrost bog measurements came from studies with field sites within 442 Canada (Figure 1; Table S2), as was the case for upland tundra and retrogressive thaw slump 443 DOC concentration data. The majority of permafrost wetland sample locations were found in 444 Russia, whereas the majority of the 414 coastal tundra sampling locations were in the USA. The 445 least represented ecosystem classes included the peatland ecosystem class, which is not strictly a 446 permafrost ecosystem as the other are, and the Yedoma ecosystem class (145 DOC 447 concentrations from 9 studies, Table S2). DOC concentrations differed significantly across the 448 eight ecosystem types (chi-square = 700, df = 7, p < 0.001; Figure 3). The highest DOC 449 concentrations were found in coastal tundra ($66 \pm 116 \text{ mg L}^{-1}$) and permafrost bogs ($63 \pm 75 \text{ mg}$) 450 L^{-1}) ecosystems. The lowest DOC concentrations were found in permafrost wetlands (7 ± 20 mg 451 L^{-1}) and Yedoma ecosystems (9 ± 18 mg L^{-1}), both of which had only slightly lower median DOC concentrations than retrogressive thaw slumps $(15 \pm 21 \text{ mg L}^{-1})$. 452



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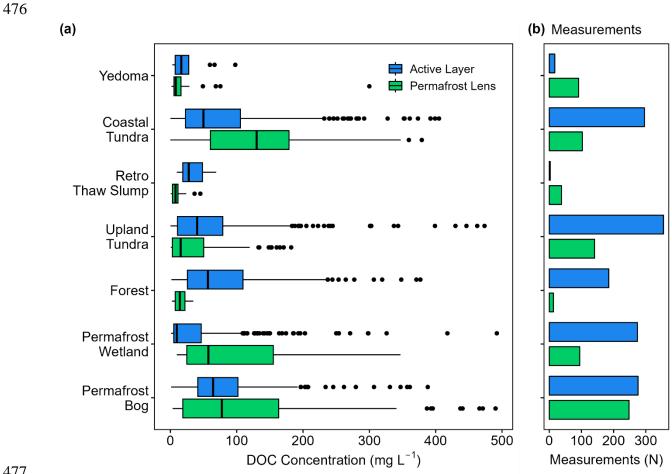


455 Figure 3. Boxplot and jitter plot of (a) DOC concentrations (mg L⁻¹), (b) the number of DOC 456 measurements, and (c) number of studies including DOC measurements were taken from the 457 top 3 m for each ecosystem type. Retro Thaw Slump = Retrogressive Thaw Slump. Boxes 458 represents the interquartile range (25 - 75%), with median shown as black horizontal line. 459 Whiskers extend to 1.5 times the interguartile range (distance between first and third quartile) in 460 each direction. Jitter points represent the concentration of each individual DOC measurement. 461 with random variation applied to each points location vertically in the plot, to avoid overplotting. 462 Yedoma = dark teal. Coastal Tundra = orange. Retro Thaw Slump = red. Upland Tundra = 463 green. Forest = purple. Permafrost Wetland = light pink. Permafrost bog = yellow. Peatland = 464 brown.

465

466 When grouping all DOC concentrations by ecosystem types and differentiating between 467 the active layer and permafrost lens thermal horizons, we found that DOC concentrations 468 differed between the active layer and permafrost lens for all ecosystems (ANCOVA: $F_{(1, 1277)} =$ 469 49.8, p < 0.001), except for permafrost bogs (chi-square = 0.37, df = 1, p = 0.5) and Yedoma 470 (chi-square = 3.5, df = 1, p = 0.06) ecosystems (Figure 4). Within the permafrost lens thermal 471 horizon, the highest DOC concentrations were found in coastal tundra (n = 103; 130 \pm 119 mg L⁻ 472 ¹) and permafrost bogs (n = 248; 78 \pm 144 mg L⁻¹) sites, and lowest found in Yedoma sites (n = 473 91; $8 \pm 10 \text{ mg L}^{-1}$). The highest active layer DOC concentrations were in permafrost bogs (n =

276; $64 \pm 61 \text{ mg } \text{L}^{-1}$) and forest (n = 185; $57 \pm 84 \text{ mg } \text{L}^{-1}$) sites, and lowest found in permafrost 474 wetland sites (n = 274; $10 \pm 42 \text{ mg L}^{-1}$). 475





478 Figure 4. Boxplot of (a) DOC concentrations (mg L⁻¹) and (b) the number of DOC

479 measurements in the Active Layer and Permafrost Lens thermal horizons of each ecosystem 480 type. Only DOC concentrations from ecosystems with these thermal horizons present is used.

481 thus no permafrost-free sites are included. Retro Thaw Slump = Retrogressive Thaw Slump.

482 Boxes represents the interguartile range (25 - 75%), with median shown as black horizontal

- 483 line. Whiskers extend to 1.5 times the interguartile range (distance between first and third
- 484 quartile) in each direction. Blue boxplots represent DOC concentrations in the active layer.
- 485 Green boxplots represent DOC concentrations in the permafrost lens.

486

487 3.4 Effect of extraction and analysis methods on DOC concentrations

- We found that DOC concentrations differed between filter sizes (ANOVA: $F_{(4, 2339)} =$ 488
- 489 22.9, p < 0.001). The highest DOC median concentrations reported were filtered using 0.45 μ m

 $(53 \pm 78 \text{ mg L}^{-1})$ and 0.22 µm $(42 \pm 54 \text{ mg L}^{-1})$ and lowest using 0.7 µm $(17 \pm 78 \text{ mg L}^{-1})$. The 490 491 majority of DOC concentrations were determined using 0.45, 0.7, and 0.22 µm filter sizes. The 492 trends observed in in DOC concentrations across study regions and ecosystems were also found 493 when exploring these trends for the three main filter sizes used (Table S3, S3). Using 0.45 and 494 0.7 µm filter sizes, which represents 79% of all reported DOC concentrations, we find that DOC 495 concentrations are generally higher in the discontinuous and sporadic permafrost zone, the two 496 boreal ecoregions, Histel soils, and the active layer thermal horizons (Table S3). Similarly, the 497 highest DOC concentrations using these two most common filter sizes were highest in 498 permafrost bog and coastal tundra ecosystems (Table S4). Given these similarities when 499 considering and not considering filter size, and the large variation in DOC concentrations within 500 each filter size, we consider the effect of filter size on the trends observed in DOC concentrations 501 across study regions and ecosystems reported above (Figure 2, 3) to be minor.

502 DOC concentrations were found to be significantly different between samples subject to 503 the six broader groups of extraction method used (ANOVA: $F_{(5, 2518)} = 30.8$, p < 0.001), and 504 between water based and soil (solid) based extraction methods (ANOVA: $F_{(1, 2524)} = 182.1$, p < 100505 0.001). The trends observed in DOC concentrations across study regions (Figure 2) and 506 ecosystems (Figure 3) were also found when exploring study region and ecosystem trends for the 507 three main DOC extraction methods used (Table S5, S6). We found that 93% of DOC 508 concentrations were determined using the suction (42%), leach (37%), and grab (14%) extraction 509 methods. Using these three most common approaches the highest DOC concentrations across 510 study regions (Table S5) and ecosystems (Table S6) were found in the discontinuous and 511 sporadic permafrost zone, the two boreal ecoregions, Histel soils, the active layer thermal 512 horizons, and in permafrost bog and coastal tundra ecosystems.

The different methods of measuring DOC concentrations also produced significantly different DOC concentrations (ANOVA: $F_{(3, 2515)} = 36.2$, p < 0.001). The three most common accounted for 99% of all DOC concentrations and were combustion, persulphate, and photometric. Of these three combustion was the most common and used for 89% of DOC measurements. The persulphate and photometric methods were not used in all study regions (Table S7) and ecosystems (Table S8), thus comparison of all three methods is not complete. Trends in DOC measured using the combustion and persulphate method (Table S7, S) were similar to those found across study regions (Figure 2) and ecosystems (Figure 3). This is
unsurprising given that both of these methods account for 98% of all DOC concentrations.

522 We consider the effect of filter size, extraction method, and method of DOC 523 measurement to be minor in determining trends in DOC concentrations across study regions and 524 ecosystems. We find that trends in DOC concentrations across study regions and ecosystems are 525 similar when you both consider and do not consider the methods used to determine those 526 concentrations. Also, the variability observed in DOC concentrations for each study region and 527 ecosystem remains high even when considering filter size, extraction method, and measurement 528 method. Thus, each method or approach similarly impacts DOC concentrations from each study 529 region and ecosystem, and cannot explain the DOC concentration variability observed within 530 each. However, these different approaches did have an impact on DOC concentrations. In this 531 study we did not focus on systematically testing the effect of filter sizes, extraction methods, or 532 DOC measurement methods. Our goal was to assess the concentration and mobilization of DOC 533 in terrestrial permafrost ecosystems across circumpolar regions and ecosystems. The assessment 534 of methods is outside the scope of our study. Rather, we compare DOC concentrations collected 535 from samples using a variety of these methods and suggest that future studies use this 536 information to decide on methods to be consistent with compiled measurements, thus far.

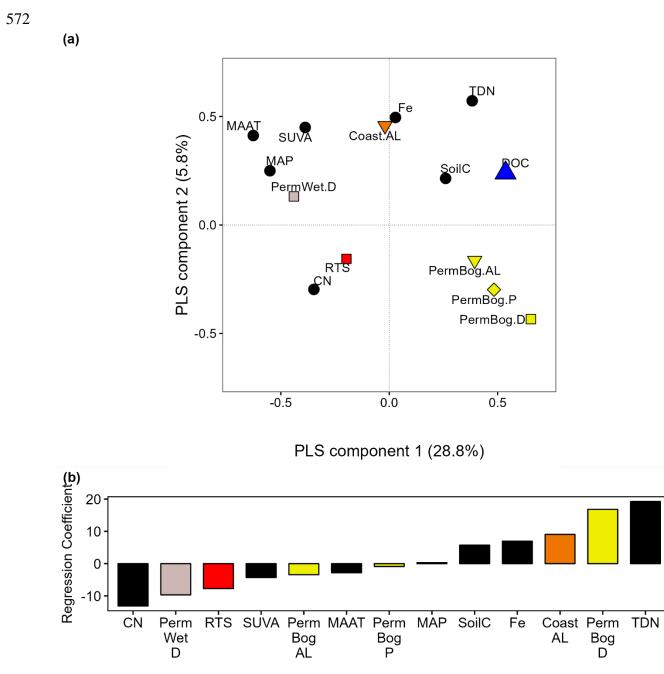
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3.5 Drivers of DOC concentrations

538 No continuous variables recorded in the dataset were available for all DOC concentration 539 database entries, with no sites containing data for all continuous variables. This limited our 540 ability to explore relationships between continuous environmental and ecological data and DOC 541 concentrations across the permafrost region. To address drivers of DOC concentrations across 542 the circumpolar permafrost region we used partial least squares regression (PLS) as it is tolerant 543 to missing values. Multiple PLS regressions were run using various combinations of continuous 544 and categorical data with similar model performance throughout. We chose the PLS to determine the drivers of DOC concentrations using environmental continuous variables and ecosystem type 545 546 as this contained the lowest background correlation. The most parsimonious PLS regression 547 extracted 9 significant components, captured 79% variation of the predictor variables, and 548 explained 37% of the variance in DOC concentrations in the dataset. The majority of the

549 variance in DOC (35%) is explained along the first two axes of the model. The model was robust 550 and not overfitted as model predictability was moderate ($Q^2 = 0.35$) and background correlation 551 was low (0.006).

552 The PLS plot (Figure 5a) shows the correlation between DOC concentrations and 553 selected environmental and ecological variables for the first two axes of the model. The two 554 variables with the greatest positive and negative relationship with DOC concentrations were total 555 dissolved nitrogen content (mg L⁻¹) and C:N ratios, respectively (Figure 5b). The positive 556 relationship of DOC with total dissolved nitrogen and soil carbon content (SoilC), and negative 557 relationship with the specific UV absorbance at 254 nm (SUVA), may be a result of ecosystem 558 properties. The strong negative relationship with C:N ratios indicates that DOC concentrations 559 decrease with increased decomposition. Other than higher soil carbon content (SoilC) in 560 permafrost bogs, there was no clear or obvious observable trends in SoilC, TDN, C:N ratios, and 561 SUVA across ecosystem types (Figure S3). The PLS demonstrates that ecosystem type strongly 562 affects DOC concentrations, with DOC positively related with the highest ecosystems where the 563 highest DOC concentrations are observed, permafrost bogs and coastal tundra, and negatively 564 related to the lower DOC ecosystems, permafrost wetland and retrogressive thaw slumps (Figure 565 5). This negative relationship may be due to the higher latitudes these ecosystems are generally 566 found at, which is supported by the negative relationship with DOC and the climate indicators 567 mean annual temperature (MAAT) and mean annual precipitation (MAP). Additionally, it may 568 be due to the high number of thermokarst affected sites found within these ecosystem classes, 569 particularly retrogressive thaw slumps. There is a clear negative relationship between DOC 570 concentrations and disturbed permafrost wetlands, retrogressive thaw slumps, and permafrost 571 bogs.



573

574 Figure 5. Partial least squares regression (PLS) (a) loadings plot explaining 37% of the 575 variability observed in DOC concentrations. (b) Bar plot of PLS regression coefficients showing 576 the relative importance of each variable in predicting DOC concentrations. Regression 577 coefficients on y-axis are normalized so their absolute sum is 100, with positive and negative 578 values indicating the direction of the relationship. In the loadings plot squares depict ecosystem 579 classes and the blue triangle represents DOC concentrations. Black circles in the (a) loadings 580 plot and black bars in the (b) bar plot represent continuous environmental data that had at lest 581 20% coverage of DOC data. Continuous data variables are represented by the colour black. CN 582 = carbon:nitrogen ratio. SUVA = the specific UV absorbance at 254 nm (L mg C⁻¹ m⁻¹). MAP =

583 mean annal precipitation (mm). MAAT = mean annual temperature. SoilC = carbon content of 584 soil (g C kg⁻¹). TDN = total dissolved nitrogen (mg L⁻¹). Fe = dissolved iron (mg L⁻¹). PermWet.D 585 = disturbed permafrost wetland ecosystem class and is light pink (as in Figure 3) to represent 586 this ecosystem class. RTS = retrogressive thaw slump ecosystem class and is red (as in Figure 587 3) to represent this ecosystem class. Coast.AL = active layer of coastal tundra ecosystem class 588 and is orange. PermBog.AL = active layer of permafrost bog ecosystem class and is yellow. 589 PermBog.P = permafrost lens of permafrost bog ecosystem class and is yellow\. PermBog.D = 590 disturbed permafrost bog ecosystem class and is yellow.

591

3.6 Response and mobilization of DOC and BDOC to thermokarst formation

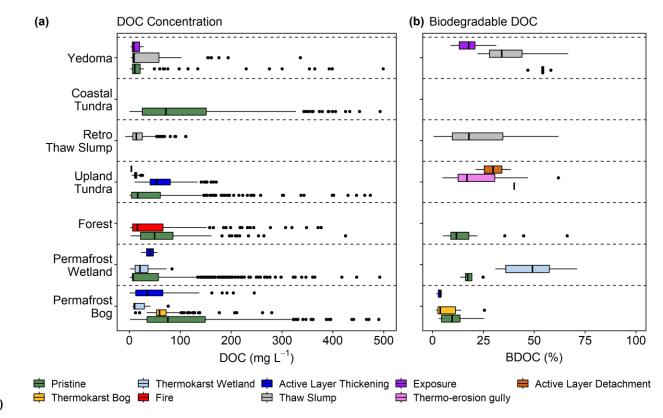
592 The highest DOC concentrations were found in pristine permafrost bog (n = 442; 75 \pm 593 112 mg L⁻¹) and coastal tundra ecosystems (n = 427; 72 \pm 126 mg L⁻¹; Figure 6a). No 594 thermokarst affected coastal tundra ecosystems were recorded within the dataset. Whereas, in 595 permafrost bogs DOC concentrations were found to differ across different thermokarst 596 disturbances (ANOVA: $F_{(3,720)} = 23.04$, p < 0.001), with the lowest found in thermokarst 597 wetlands (n = 16; $10 \pm 21 \text{ mg L}^{-1}$). DOC concentrations were also found to differ between thermokarst affected and pristine sites in upland tundra ecosystems (ANOVA: $F_{(3, 539)} = 5.91$, p 598 599 < 0.001). The highest DOC concentrations in upland tundra ecosystems were found in sites that had experienced active layer thickening (n = 142; 53 \pm 39 mg L⁻¹), whereas the lowest were 600 found in sites that had experienced active layer detachment (n = 6; $4 \pm 2 \text{ mg L}^{-1}$). Pristine sites 601 had the highest DOC concentrations in both Yedoma (n = 114; $11 \pm 15 \text{ mg L}^{-1}$) and forest (n =602 189; 49 \pm 64 mg L⁻¹) ecosystems. However, in permafrost wetland ecosystems pristine sites had 603 the lowest DOC concentrations (n = 766; $7 \pm 51 \text{ mg L}^{-1}$) with sites that were affected by both 604 thermokarst wetland formation (n = 17; 21 \pm 26 mg L⁻¹) and active layer thickening (n = 12; 41 \pm 605 13 mg L^{-1}) having higher DOC concentrations. 606

607 Our database contained limited data regarding BDOC (n = 146), thus BDOC results 608 across ecosystems should be interpreted with caution. Due to limited data we have combined 609 BDOC over all incubation lengths when assessing BDOC between pristine and thermokarst sites 610 (Figure 6). BDOC was found to differ between thermokarst disturbances within ecosystem types 611 in only Yedoma (ANOVA: $F_{(2,27)} = 23.09$, p < 0.001) and permafrost wetland (ANOVA: $F_{(1,10)}$ 612 = 15.87, p < 0.001) ecosystems. The highest BDOC was found in both of these ecosystem types 613 also, with 54% (n = 5) in pristine Yedoma sites and 49% (n = 8) in thermokarst wetland affected 614 permafrost wetland sites (Figure 6b), with the latter exhibiting the highest BDOC across all

615 permafrost affected sites followed by thaw slumps (18%, n = 11) in Yedoma ecosystems and 616 active layer thickening (40%, n = 1) in upland tundra sites. The lowest median BDOC of 4% 617 were seen in thermokarst bogs (n = 5) and active layer thickening (n = 3) affected sites, with 618 pristine sites experiencing BDOC of 9% (n = 15). However, not all ecosystem types in the 619 database had BDOC data for both pristine and disturbance sites. For example, only pristine sites 620 data was available for forests, whereas there was no pristine site data available for upland tundra 621 sites. No BDOC data was available for coastal tundra sites.

622 All ecosystem types that had BDOC data, reported BDOC observed following 40-90623 incubation days, and this also corresponded to the highest BDOC values for each ecosystem type 624 (Figure S4). When comparing the greatest BDOC observed within this incubation length 625 window, we found that values varied across ecosystem type (ANOVA: $F_{(5, 131)} = 14.6, p < 1000$ 626 0.001). The highest loss rates were observed in Yedoma and permafrost wetland ecosystems, 627 whereas the lowest we observed in organic rich forest and permafrost bog ecosystems (Figure 628 S4). Forest (ANOVA: $F_{(1, 16)} = 2.31$, p = 0.15) and permafrost bog (ANOVA: $F_{(3, 24)} = 2.49$, p = 0.15) 629 0.09) BDOC did not differ over incubation length, whereas Yedoma (ANOVA: $F_{(4, 25)} = 24.92, p$ 630 < 0.001) and permafrost wetland (ANOVA: F_(1,10) = 15.87, p < 0.01) did differ over time, with 631 their max occurring during this 40 - 90-day incubation length. This suggests that when incubated 632 for the same number of days, we would expect greater BDOC in Yedoma and permafrost 633 wetland ecosystems. Note, for this analysis BDOC values from all thermokarst and non-634 thermokarst affected sites within an ecosystem type were included. Given the limited BDOC data 635 available we have compared BDOC across ecosystems in two ways. The first is using data from 636 all measurement days to assess BDOC across pristine and disturbed ecosystems (Figure 6b). The 637 second is assessing max BDOC within each ecosystem type, which includes pristine and 638 disturbed sites (Figure S4). Using both approaches we find that the highest BDOC is observed in

639 high-latitude Yedoma and permafrost wetland sites.



640

641 Figure 6. DOC concentrations (mg L⁻¹) and biodegradable DOC (BDOC; %) from the top 3 m following disturbance including data from both field based and incubation studies. (a) DOC 642 643 concentrations from each ecosystem type following disturbance where data was available. (b) 644 Biodegradable DOC (BDOC) from each ecosystem type following disturbance where data was 645 available. BDOC loss was determined following 3 - 304 days of incubation. Data from different 646 incubation lengths was combined due to low sample size. Retro Thaw Slump = Retrogressive 647 Thaw Slump. Boxes represents the interquartile range (25 - 75%), with median shown as black 648 horizontal line. Whiskers extend to 1.5 times the interguartile range (distance between first and 649 third quartile) in each direction, with outlier data plotted individually as black dots. Note colours 650 associated with boxplots in this figure are only relevant for this figure.

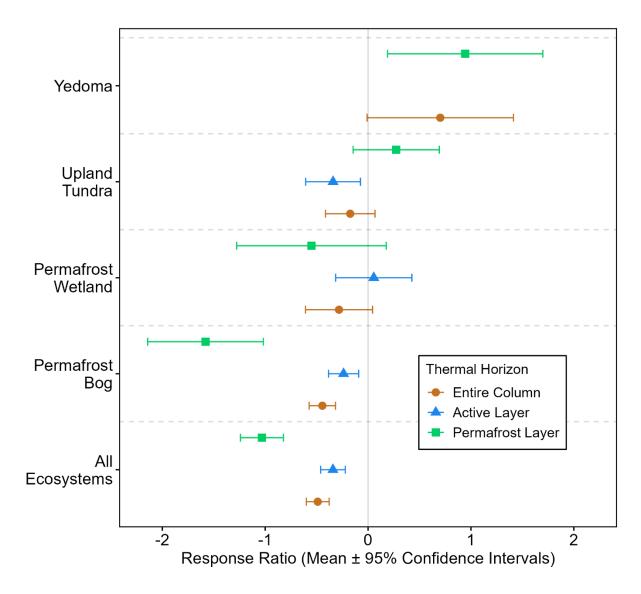
Response ratios comparing the change in DOC concentrations between pristine and thermokarst affected sites were calculated from our dataset from 108 studies using Eq. 1 (Figure 7). Only 17 studies provided data for both pristine and thermokarst affected ecosystems, with 87 papers providing DOC concentrations from pristine and 34 from thermokarst affected sites. When considering all ecosystems together we found that response ratios were negative, suggesting that DOC concentrations were higher in thermokarst affected sites compared to

- 657 pristine sites (Figure 7). These negative response ratios were most evident in permafrost bogs,
- 658 where they found throughout the entire column and individual thermal horizons. The greatest

659 increase in DOC concentrations following thermokarst was seen when comparing DOC

- 660 concentrations in the permafrost lens of permafrost bogs, and to a lesser extent permafrost
- 661 wetlands (Figure 7). Only in Yedoma ecosystems did we see positive response ratios throughout
- the entire profile, suggesting a decrease in DOC concentrations following thermokarst formation
- 663 in Yedoma sites. This was also seen for DOC concentrations within the permafrost lens of
- 664 upland tundra sites, which include DOC concentrations from retrogressive thaw slumps and
- thermo-erosion gullies in their thermokarst affected sites. The large confidence intervals for
- some response ratios suggests high variability in the response of DOC concentrations to
- 667 thermokarst formation.

668



669

670 Figure 7. Response ratios of DOC concentrations from the top 3 m following thermokarst 671 formation (calculated using Eq. 1). Response ratio means allow for relative comparison of 672 changes in DOC following thermokarst formation between different ecosystem types. Negative 673 values indicate lower DOC concentrations found in pristine ecosystems, whereas positive value 674 indicates a decrease in DOC concentrations following thermokarst. Studies reporting DOC 675 concentrations from Exposures, Retrogressive Thaw Slumps, and Thermo-Erosion Gullies from 676 sites within the continuous permafrost zone were combined into the Upland Tundra ecosystem 677 category. This did not include DOC concentrations from studies within the Yedoma permafrost 678 domain (Strauss et al., 2021). Blue line represent DOC concentrations in the active layer, as per 679 Figure 4. Green lines represent DOC concentrations in the permafrost lens, as per Figure 4.

Brown lines represent DOC concentrations from the entire column (i.e., both active layer andpermafrost lens).

682 **4. Discussion**

683 In this systematic review, we evaluated patterns of DOC concentrations in the top 3 m of 684 soil in terrestrial ecosystems across the northern circumpolar permafrost region based on results 685 from 111 studies and 2,845 DOC measurements. We focused on comparing concentrations of 686 DOC in soils across various geographical regions, ecological conditions, and disturbance types. 687 Our synthesis shows that median DOC concentrations across ecosystems range from 9-61 mg 688 L^{-1} , which represents similar albeit slightly higher DOC concentrations when compared to the 689 median DOC concentrations found in top soils of other land cover groups below 50°N (25 mg L⁻ 690 ¹; Langeveld et al., 2020), globally distributed lakes (6 mg L⁻¹; Sobek et al., 2007), and lakes across the permafrost region (11 mg L⁻¹; Stolpmann et al., 2021). In general, we show that 691 692 organic soils have higher DOC concentrations than mineral soils, and that DOC concentrations 693 are positively related to total dissolved nitrogen concentrations and negatively to C:N ratios, 694 which corroborate previous findings of factors correlating with DOC concentrations (Aitkenhead 695 & McDowell, 2000; Lajtha et al., 2005). Overall, we found that properties associated with 696 ecosystem type are the main constraint on DOC concentrations. Furthermore, disturbance 697 through permafrost thaw has little impact on measured DOC concentrations, however this may 698 be due to the loss of biologically reactive DOC or the loss of an initially larger pulse of DOC 699 having been previously mobilised prior to the timing of sampling.

700 4.1 Environmental factors influencing DOC

701 Our database confirmed our first hypothesis that the highest DOC concentrations would be 702 found in organic rich soils. Previous synthesis efforts estimating global distributions of terrestrial 703 DOC concentrations have presented similar findings (Guo et al., 2020; Langeveld et al., 2020). 704 Both of these previous studies also show that some of the highest terrestrial DOC concentrations 705 are found within the northern circumpolar permafrost region, highlighting that these high DOC 706 concentrations found in organic rich permafrost soils are of global significance. Concentrations 707 of DOC in the top 3 m of soils closely mirrored stocks of SOC across the circumpolar permafrost 708 region (Hugelius et al., 2014). Organic rich Histosol and Histel soils contain the greatest SOC

709 per km², followed by Turbels and Orthels (Hugelius et al., 2014). The leaching of organic C from 710 soils act as a major source of DOC (Kalbitz et al., 2000; Marschner & Bredow, 2002), thus it is 711 not surprising that we find the highest DOC concentrations in the soil types with the greatest 712 quantities of SOC (Figure 2a). While the highest DOC concentrations are found within organic 713 rich soils, the amount of C found as DOC represent a small amount of the total SOC pool. Using 714 the current best estimates of Histel SOC stocks (Hugelius et al., 2020), the DOC pool represents 715 <1% of the total C stock in permafrost-affected peatlands as has been shown for both permafrost 716 and global soils (Guo et al., 2020; Prokushkin et al., 2008).

717 *4.2 Variation in DOC across ecosystems*

718 The accumulation of high DOC concentrations we show in permafrost bogs and permafrost 719 wetlands (Figure 3), is a result of the prevalence of cold and anoxic conditions throughout the 720 Holocene (Blodau, 2002). This leads to a reduction in microbial decomposition, and the 721 accumulation of both a large SOC (Hugelius et al., 2020) and DOC pool. Our results suggest that 722 the pristine permafrost bog and permafrost wetland DOC pool is relatively stable following 723 permafrost thaw (Figure 6, 7a). The lower DOC pool found in the active layer of permafrost 724 wetland (Figure 4a)may represent a potentially labile DOC pool (Figure 7a), but this is likely due 725 to fresh, plant derived inputs rather than the exposure and mineralization of previously frozen 726 organic matter (Figure 7a). Peatland vegetation, in particular Sphagnum mosses, produces litter 727 that has anti-microbial properties and is decay resistant (Hamard et al., 2019; Limpens, Bohlin, 728 & Nilsson, 2017), limiting the amount of SOC that is degraded and assimilated into the DOC 729 pool (Tfaily et al., 2013). This is further enhanced by the build-up of decomposition end products 730 and the thermodynamic constraint on decay observed in anoxic soils (Beer et al., 2008). 731 Permafrost has been continuously present in peatlands across the northern circumpolar 732 permafrost region for the past 6,000 years, with the greatest rates of permafrost formation 733 occurring within the past 3,000 years (Treat & Jones, 2018). Thus, a large proportion of the 734 organic matter found peatlands and wetlands in this region were present prior to permafrost 735 aggradation (i.e., permafrost formation), which indicates that permafrost formed epigenetically in 736 these areas. Permafrost aggradation impacts soil biogeochemical properties, leading to 737 potentially less decomposed organic matter with higher C/N ratios than non-permafrost 738 equivalent soils, particularly in permafrost wetlands (Treat et al., 2016). This can lead to the

31

build-up of high DOC concentrations that are vulnerable to potential mobilization following

740 thermokarst. Decomposition in epigenetic permafrost bogs following thermokarst has been

shown to be relatively slow (Heffernan et al., 2020; Manies et al., 2021), which further supports

our finding (Figure 6) that the large DOC pool found in these systems in relatively stable

following permafrost thaw.

744 Coastal tundra ecosystems had similarly high DOC concentrations to those found in 745 permafrost bogs (Figure 3a). Coastal tundra ecosystems represented the highest concentrations of 746 DOC in mineral permafrost soils, with the highest concentrations found in the permafrost lens 747 (Figure 4a). This is contrary to findings that deeper coastal permafrost consists of low organic 748 matter Pleistocene marine sediments (Bristol et al., 2021) and the proximity of the active layer to 749 vegetation inputs, although this productivity and inputs are vulnerable to projected climatic 750 warming and regional "browning" and "greening" (Lara et al., 2018). Recent work has shown 751 that DOC in the active layer within the coastal permafrost is more biodegradable that OC in the 752 permafrost lens (Speetjens et al., 2022) and a substantial proportion of organic carbon derived 753 from thawing coastal permafrost is vulnerable to mineralization upon thawing, particularly when 754 exposed to sea water (Tanski et al., 2021). Export of terrestrial coastal permafrost DOC directly 755 into the Arctic Ocean can significantly influence marine biogeochemical cycles and food webs 756 within the Arctic ocean (Bruhn et al., 2021). Arctic coasts are eroding at rates of up to 25 m yr⁻¹ 757 (Fritz, Vonk, & Lantuit, 2017) and exporting large quantities of terrestrial organic matter export 758 directly to the ocean that is rapidly mineralized (Tanski et al., 2019). Enhanced DOC export from 759 these coastal tundra ecosystems may disrupt aquatic food webs through altering nutrient and 760 light supply, as has been shown for Swedish coastal systems (Peacock et al., 2022). These 761 coastal tundra sites represent a large DOC pool that is highly vulnerable to enhanced 762 mobilization and deserve further attention.

We found that DOC concentrations increased along a clear latitudinal gradient, from north to south, in the remaining ecosystems characterised by mineral soils with an upper organic layer, i.e., forests, upland tundra, and Yedoma. In forest ecosystems, the upper organic layer, and the impact of soil temperature, moisture, and pH on SOC found there, strongly influences the production, concentration, and composition of DOC (Neff & Hooper, 2002; Wickland et al., 2007). Furthermore, the sorption of DOC to charcoal (Guggenberger et al., 2008), and high 769 lignin and phenolic input from vegetation (O'Donnell et al., 2016) produce a difficult to degrade 770 DOC pool, leading to the accumulation of the large DOC pool in the active layer (Figure 4a) this 771 ecosystem type. This trend with depth has also been observed in the vertical distribution of DOC 772 across global soils, with 50% of the DOC pool found in the top 0 - 30 cm (Guo et al., 2020). 773 While not included in the most parsimonious PLS model (Figure 5), Yedoma and upland tundra 774 ecosystems were found to negatively correlate with DOC concentrations (Figure S5). The 775 greatest proportions of OC and nutrients used for DOC production in these ecosystems are found 776 in shallow organic layers (Semenchuk et al., 2015; Wild et al., 2013). Beneath the upper organic 777 horizons in these mineral soils processes such as sorption of DOC to minerals and the formation 778 of Fe-DOC or Al-DOC complexes may remove DOC from the dissolved pool (Kawahigashi et 779 al., 2006) and mechanically protect it from mobilization (Gentsch et al., 2015). The majority of 780 vegetation and its leachates found in the permafrost region produce relatively stable DOC 781 consisting of lignin-derived compounds, highly aromatic polyphenolic compounds, and low 782 molecular weight organic acids (Chen et al., 2018; Drake et al., 2015; Ewing et al., 2015; Selvam 783 et al., 2017). While differences in the stability of different DOC source end-members have been 784 shown (MacDonald et al., 2021), differences in redox conditions are likely a major driver in 785 differences in the accumulation and mineralization of DOC across permafrost ecosystem types 786 (Mohammed et al., 2022).

787

4.3 Vulnerability of DOC to enhanced mobilization following thermokarst

788 We define DOC mobilization as DOC lost from an ecosystem either via export or 789 degradation. Our second hypothesis that permafrost thaw would lead to enhanced mobilization of 790 DOC cannot be fully supported by the findings from this database. Using our chosen systematic 791 approach and focusing on data from terrestrial ecosystems, our database was limited to 3 studies 792 which represented <1% of the DOC concentration data. Several previous studies have detailed 793 the export of DOC in Arctic inland waters, see Table 2 in Ma et al., (2019). These studies were 794 excluded using our systematic approach (Table 1 and 2) as they do not directly measure DOC 795 export from a terrestrial ecosystem, rather they determine the quantity of terrestrial derived DOC 796 found in inland waters. This is a key distinction, as by not quantifying the export rates for 797 terrestrial ecosystems the net ecosystem carbon balance and vulnerability to enhanced export 798 may not be assessed. We acknowledge the limitation in our approach regarding the inclusion of

799 DOC export data. Thus, this database cannot be used to determine how permafrost thaw will 800 influence DOC export from terrestrial ecosystems within the northern circumpolar permafrost 801 region. However, we identify this lack of export data from terrestrial permafrost ecosystems as a 802 key knowledge gap in our current understanding of the permafrost carbon pool. Currently, Arctic 803 rivers are estimated to export 25 - 36 Tg DOC year⁻¹ (Amon et al., 2012; Holmes et al., 2012), 804 with this being dominated by modern carbon sources (Estop-Aragonés et al., 2020), most likely 805 derived from the top 1 m of terrestrial ecosystems. Using current best estimates of the areal 806 extent and soil organic carbon stores in the top 1 m of Histosols, Histels, Orthels and Turbels 807 (Hugelius et al., 2014), and if we assume that the DOC pool represents ~1% of the SOC pool, we 808 estimate that <1% of the current DOC pool found in the top 1 m of Histosols, Histels, Orthels 809 and Turbels is exported annually to Arctic rivers. Quantifying the proportion of these DOC pools 810 annually lost, and particularly the proportions lost in headwater streams while being exported to 811 Arctic rivers, is vital to assess the importance of the mobilization of the terrestrial permafrost 812 DOC pool.

813 Our calculated response ratios (Figure 7) for all ecosystems, indicating the difference in DOC 814 concentrations between pristine and permafrost thaw affected sites, partly supports of our second 815 hypothesis that disturbance would lead to increased export and biodegradability of DOC. The 816 increase in DOC following thaw observed in permafrost bogs is likely due to increased inputs 817 due to increased runoff and shifts in vegetation following permafrost thaw (Burd, Estop-818 Aragonés, Tank, & Olefeldt, 2020), enhanced release of DOC (Loiko et al., 2017), a relatively 819 stable soil organic carbon pool at depth due to several millennia of microbial processing (Manies 820 et al., 2021), the prevalence of anoxic conditions, and the potential hydrological isolation of 821 thermokarst bogs (Quinton, Hayashi, & Pietroniro, 2003). While not included in our analysis, 822 DOC found near the surface of the permafrost lens in forest ecosystems has been shown to be 823 more biodegradable than DOC found in the active layer (Wickland et al., 2018), and may 824 represent a decrease in DOC following thermokarst not captured here. Our findings of limited 825 mobilization of permafrost bog DOC upon thawing are supported by the findings that the ¹⁴C 826 signature of DOC in Arctic rivers is dominated by modern sources (Estop-Aragonés et al., 2020). 827 However, individual studies have determined that thawing may release a large pool of permafrost 828 peatland DOC into aquatic networks (Lim et al., 2021). We do see a reduction in DOC

829 concentrations in thermokarst affected sites at the higher latitude Yedoma, upland tundra, and 830 permafrost wetland ecosystems. This reduction in DOC concentrations in these ecosystems may 831 be due to the greater biodegradability and lability of the DOC found there (Figure 6b), 832 supporting our third hypothesis that the most biodegradable DOC would be found in higher 833 latitude ecosystems. Permafrost DOC in higher latitude ecosystems, particularly Yedoma 834 ecosystems, is characterised by syngenetic permafrost aggradation which have not undergone 835 centuries to millennia of soil formation and microbial processes, have been shown contain a 836 greater proportion of low oxygen, aliphatic compounds and labile substrates (Ewing et al., 837 2015b; MacDonald et al., 2021). This leads to a greater biolability and rapid mineralization of 838 DOC (Vonk et al., 2015), potentially causing the reduction in DOC concentrations observed 839 following thaw. If this hypothesis is to be found true across all high latitude ecosystems with 840 further data, it further highlights the vulnerability of the large DOC pool found in coastal tundra 841 ecosystems.

842 In this study, we focus on the dissolved fraction of the OC pool, however the particulate 843 fraction should also be considered when discussing the mobilization of terrestrial OC in 844 permafrost landscapes. In boreal freshwater networks, particulate organic carbon (POC) 845 represents a small but highly labile fraction of terrestrially derived OC exported to the fluvial 846 network (Attermeyer et al., 2018). The degradation of permafrost derived POC is much slower 847 than that of POC in the boreal freshwater network and POC derived from younger sources along 848 the riverbank (Shakil, Tank, Kokelj, Vonk, & Zolkos, 2020). The DOC pool in Arctic 849 freshwaters in dominated by modern terrestrial sources (Estop-Aragonés et al., 2020), whereas 850 the POC pool has been shown to be dominated by older sources in both permafrost peatland 851 dominated areas (Wild et al., 2019), following the formation of retrogressive thaw slumps 852 (Keskitalo et al., 2021), and in thermokarst affected periglacial streams (Bröder et al., 2022). 853 This older POC has been shown to accumulate following export due to low lability and 854 degradation and mineral association, which suggests that upon thermokarst formation, previously 855 frozen OC exported in the particulate phase is not readily consumed by microbes and that 856 permafrost derived DOC is the more labile fraction of exported terrestrial OC.

857 *4.4 Future considerations for study design*

35

858 Determining the fate of mobilized terrestrial DOC in both permafrost thaw affected, and 859 pristine sites should be prioritized in future studies to constrain current estimates of the 860 permafrost C climate feedback. There are large spatial gaps in the database, particularly in areas 861 with large stock of permafrost C such as the Hudson Bay Lowlands and Mackenzie River Basin, 862 both in Canada and two of the three largest deposits of permafrost peatland C in the circumpolar 863 permafrost region (Olefeldt et al., 2021). Similarly, coastal tundra sites, which along with 864 permafrost bog represent the ecosystems with the highest DOC concentrations, were sampled 865 only along the northern shoreline of Alaska and the Yukon (USA and Canada, respectively; 866 Table S2). From our analysis of this database, we determine that DOC mobilization is poorly 867 understood for terrestrial permafrost ecosystems. To address this, the two main needs of future 868 studies are 1) more direct estimates of DOC fluxes and export from terrestrial ecosystems into 869 aquatic ecosystems, and 2) more DOC degradation (BDOC) and mineralization studies. Our 870 results suggest that the high concentrations of DOC in permafrost bogs remains relatively stable 871 upon thermokarst formation, although individual studies do indicate that thawing peat may 872 provide a reactive source of DOC (Panneer Selvam et al., 2017). The database did not include 873 any studies that reported on the mineralization of DOC from coastal tundra sites, thus we are 874 unable to comment on the stability of the high DOC concentrations found in this ecosystem type. 875 Further sampling and assessing the mineralization of DOC is required to characterize the 876 potential pool of vulnerable DOC in areas with high DOC concentrations. Overall, our database 877 and systematic approach only included 5 studies (Olefeldt & Roulet, 2012, 2014; Olefeldt et al., 878 2012; Prokushkin et al., 2006; Prokushkin et al., 2005) that explicitly reported rates of DOC 879 discharge, export, or fluxes from terrestrial ecosystems into the fluvial network. Given the 880 importance of terrestrial DOC as a source for CO₂ production within the aquatic network 881 (Weyhenmeyer et al., 2012), and the findings that previously frozen DOC is being exported to 882 the freshwater network (Estop-Aragones et al., 2020), improved estimates of the quantity of 883 terrestrial DOC being exported is essential to determine the potential aquatic greenhouse gas 884 fluxes derived from the mineralization of terrigenous organic matter. To improve current 885 estimates of the permafrost C feedback further studies are needed to determine how much DOC 886 is laterally exported from terrestrial ecosystems, and the mineralization potential of this DOC 887 along the terrestrial-freshwater-aquatic continuum.

888 Lastly, we suggest that future studies should consider a standardization of methods and 889 approached used to determine DOC concentrations for better comparison across studies. In 890 constructing this database we identified three different filter sizes, eleven different extraction 891 procedures, and four different measurement methods. The most common filter size used was 892 0.45 µm and this has previously been described as the cut off to separate DOC from colloid 893 materials (Thurman 1985; Bolan et al., 1999). In extracting DOC concentrations from soils the 894 mostly commonly used approach (70% of all soil samples) was via soil leaching with no 895 chemical treatment of the soils, although some added filtered water to promote leaching. From 896 the seven approaches identified to extract water samples from terrestrial sites in determining 897 DOC, 48% of samples were collected using a variety of suction devices and 46% done via grab 898 samples. Of the four DOC measurements methods the most common approach was by 899 combustion, with 90% of all DOC concentrations measured using this approach. As such, in 900 order to continue measuring DOC concentrations in terrestrial permafrost ecosystems using the 901 most consistent approach we suggest using 0.45 μ m filters, extracting pore water via some type 902 of sucking device or soils via leaching, and using a combustion based method to determine DOC 903 concentrations

904 Data availability

All data is freely and publicly available at https://doi.org/10.17043/heffernan-2024-doc-1

906 Author contributions

LH, DK, and LT designed and planned the systematic review approach; LH built the database.
LH and DK analyzed the data; LH wrote the manuscript draft; DK and LT edited and reviewed

909 the manuscript.

910 Competing interests

911 The authors declare that they have no conflict of interest.

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919 **References (in text)**

- Abbott, B. W., Larouche, J. R., Jones, J. J. B., Bowden, W. B., & Balser, A. W. (2014). From
 Thawing and Collapsing Permafrost. *Journal of Geophysical Research: Biogeosciences*,
 119, 2049–2063. https://doi.org/10.1002/2014JG002678.Received
- Aitkenhead, J. A., & McDowell, W. H. (2000). Soil C:N ratio as a predictor of annual riverine
 DOC flux at local and global scales. *Global Biogeochemical Cycles*, 14(1).
 https://doi.org/10.1029/1999GB900083
- Amon, R. M. W., Rinehart, A. J., Duan, S., Louchouarn, P., Prokushkin, A., Guggenberger, G.,
 ... Zhulidov, A. V. (2012). Dissolved organic matter sources in large Arctic rivers. *Geochimica et Cosmochimica Acta*, 94, 217–237.
- 929 https://doi.org/https://doi.org/10.1016/j.gca.2012.07.015
- Andersen, C.M. and Bro, R. (2010), Variable selection in regression—a tutorial. J.
 Chemometrics, 24: 728-737. https://doi.org/10.1002/cem.1360
- Andresen, C. G., Lawrence, D. M., Wilson, C. J., McGuire, A. D., Koven, C., Schaefer, K.,
 Jafarov, E., Peng, S., Chen, X., Gouttevin, I., Burke, E., Chadburn, S., Ji, D., Chen, G.,
- Hayes, D., and Zhang, W.: Soil moisture and hydrology projections of the permafrost region
- 935
 - a model intercomparison, The Cryosphere, 14, 445–459, https://doi.org/10.5194/tc-14

 936
 445-2020, 2020.
- Arksey, H., & O'Malley, L. (2005). Scoping studies: Towards a methodological framework.
 International Journal of Social Research Methodology: Theory and Practice, 8(1).
- 939 https://doi.org/10.1080/1364557032000119616
- Attermeyer, K., Catalán, N., Einarsdottir, K., Freixa, A., Groeneveld, M., Hawkes, J. A., ...
 Tranvik, L. J. (2018). Organic Carbon Processing During Transport Through Boreal Inland
 Waters: Particles as Important Sites. *Journal of Geophysical Research: Biogeosciences*, *123*(8). https://doi.org/10.1029/2018JG004500
- 944 Beckebanze, L., Runkle, B. R. K., Walz, J., Wille, C., Holl, D., Helbig, M., ... Kutzbach, L.

- 945 (2022). Lateral carbon export has low impact on the net ecosystem carbon balance of a
- polygonal tundra catchment. *BIOGEOSCIENCES*, *19*(16), 3863–3876.
- 947 https://doi.org/10.5194/bg-19-3863-2022
- Beer, J., Lee, K., Whiticar, M., & Blodau, C. (2008). Geochemical controls on anaerobic organic
 matter decomposition in a northern peatland. *Limnology and Oceanography*, 53(4), 1393–
 1407. https://doi.org/10.4319/lo.2008.53.4.1393
- Biester, H., Knorr, K. H., Schellekens, J., Basler, A., & Hermanns, Y. M. (2014). Comparison of
 different methods to determine the degree of peat decomposition in peat bogs. *Biogeosciences*. https://doi.org/10.5194/bg-11-2691-2014
- Blodau, C. (2002). Carbon cycling in peatlands A review of processes and controls.
 Environmental Reviews, 10(2), 111–134. https://doi.org/10.1139/a02-004
- Bolan, N.S., Baskaran, S., Thiagarajan, S. (1999). Methods of Measurement of Dissolved
 Organic Carbon of Plant Origin in Soils, Manures, Sludges and Stream Water. In: Linskens,
 H.F., Jackson, J.F. (eds) Analysis of Plant Waste Materials. Modern Methods of Plant
 Analysis, vol 20. Springer, Berlin, Heidelberg. https://doi.org/10.1007/978-3-662-038871_1
- Bristol, E. M., Connolly, C. T., Lorenson, T. D., Richmond, B. M., Ilgen, A. G., Choens, R. C.,
 McClelland, J. W. (2021). Geochemistry of Coastal Permafrost and Erosion-Driven
 Organic Matter Fluxes to the Beaufort Sea Near Drew Point, Alaska. *Frontiers in Earth Science*, 8. https://doi.org/10.3389/feart.2020.598933
- Bröder, L., Hirst, C., Opfergelt, S., Thomas, M., Vonk, J. E., Haghipour, N., ... Fouché, J.
 (2022). Contrasting Export of Particulate Organic Carbon From Greenlandic Glacial and
 Nonglacial Streams. *Geophysical Research Letters*, 49(21).
 https://doi.org/10.1029/2022GL101210
- Brown, J., Ferrians Jr., O. J., Heginbottom, J. A., & Melnikov, E. S. (1997). Circum-Arctic map
 of permafrost and ground ice conditions. USGS Numbered Series, 1.
 https://doi.org/10.1016/j.jallcom.2010.03.054
- Bruhn, A. D., Stedmon, C. A., Comte, J., Matsuoka, A., Speetjens, N. J., Tanski, G., ... Sjöstedt,
 J. (2021). Terrestrial Dissolved Organic Matter Mobilized From Eroding Permafrost
 Controls Microbial Community Composition and Growth in Arctic Coastal Zones.
- 975 Frontiers in Earth Science, 9. https://doi.org/10.3389/feart.2021.640580
- Burd, K., Estop-Aragonés, C., Tank, S. E., & Olefeldt, D. (2020). Lability of dissolved organic
 carbon from boreal peatlands: interactions between permafrost thaw, wildfire, and season. *Canadian Journal of Soil Science*, *13*(February), 1–13. https://doi.org/10.1139/cjss-20190154
- 980 Camill, P. (2005). Permafrost thaw accelerates in boreal peatlands during late-20th century

- 981 climate warming. *Climatic Change*, 68(1–2), 135–152. https://doi.org/10.1007/s10584-005982 4785-y
- 983 Chanton, J. P., Glaser, P. H., Chasar, L. S., Burdige, D. J., Hines, M. E., Siegel, D. I., ... Cooper,
 984 W. T. (2008). Radiocarbon evidence for the importance of surface vegetation on
 985 fermentation and methanogenesis in contrasting types of boreal peatlands. *Global*986 *Biogeochemical Cycles*, 22(4), 1–11. https://doi.org/10.1029/2008GB003274
- Chen, H., Yang, Z., Chu, R. K., Tolic, N., Liang, L., Graham, D. E., ... Gu, B. (2018). Molecular
 Insights into Arctic Soil Organic Matter Degradation under Warming. *ENVIRONMENTAL SCIENCE & TECHNOLOGY*, 52(8), 4555–4564. https://doi.org/10.1021/acs.est.7b05469
- 990 Chong, I. G., & Jun, C. H. (2005). Performance of some variable selection methods when
- multicollinearity is present. *Chemometrics and Intelligent Laboratory Systems*, 78(1).
 https://doi.org/10.1016/j.chemolab.2004.12.011\
- Connon, R. F., Quinton, W. L., Craig, J. R., & Hayashi, M. (2014). Changing hydrologic
 connectivity due to permafrost thaw in the lower Liard River valley, NWT, Canada.
 Hydrological Processes, 28(14). https://doi.org/10.1002/hyp.10206
- Dean, J. F., Meisel, O. H., Rosco, M. M., Marchesini, L. B., Garnett, M. H., Lenderink, H., ...
 Dolman, A. J. (2020). East Siberian Arctic inland waters emit mostly contemporary carbon. *NATURE COMMUNICATIONS*, 11(1). https://doi.org/10.1038/s41467-020-15511-6
- Drake, T. W., Wickland, K. P., Spencer, R. G. M., McKnight, D. M., & Striegl, R. G. (2015).
 Ancient low-molecular-weight organic acids in permafrost fuel rapid carbon dioxide
 production upon thaw. *PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA*, *112*(45), 13946–13951.
 https://doi.org/10.1073/pnas.1511705112
- Ernakovich, J. G., Lynch, L. M., Brewer, P. E., Calderon, F. J., & Wallenstein, M. D. (2017).
 Redox and temperature-sensitive changes in microbial communities and soil chemistry
 dictate greenhouse gas loss from thawed permafrost. *BIOGEOCHEMISTRY*, *134*(1–2),
 183–200. https://doi.org/10.1007/s10533-017-0354-5
- Estop-Aragones, C., Olefeldt, D., Abbott, B. W., Chanton, J. P., Czimczik, C. I., Dean, J. F., ...
 Anthony, K. W. (2020). Assessing the Potential for Mobilization of Old Soil Carbon After
 Permafrost Thaw: A Synthesis of C-14 Measurements From the Northern Permafrost
 Region. *GLOBAL BIOGEOCHEMICAL CYCLES*, *34*(9).
- 1012 https://doi.org/10.1029/2020GB006672
- Estop-Aragonés, Cristian, Czimczik, C. I., Heffernan, L., Gibson, C., Walker, J. C., Xu, X., &
 Olefeldt, D. (2018). Respiration of aged soil carbon during fall in permafrost peatlands
 enhanced by active layer deepening following wildfire but limited following thermokarst. *Environmental Research Letters*, *13*(8). https://doi.org/10.1088/1748-9326/aad5f0
- 1017 Ewing, S. A., Paces, J. B., O'Donnell, J. A., Jorgenson, M. T., Kanevskiy, M. Z., Aiken, G. R.,

- 1018 ... Striegl, R. (2015a). Uranium isotopes and dissolved organic carbon in loess permafrost:
- 1019Modeling the age of ancient ice. GEOCHIMICA ET COSMOCHIMICA ACTA, 152, 143–1020165. https://doi.org/10.1016/j.gca.2014.11.008

Ewing, S. A., Paces, J. B., O'Donnell, J. A., Jorgenson, M. T., Kanevskiy, M. Z., Aiken, G. R.,
Striegl, R. (2015b). Uranium isotopes and dissolved organic carbon in loess permafrost:
Modeling the age of ancient ice. *Geochimica et Cosmochimica Acta*, *152*, 143–165.

- 1024 https://doi.org/10.1016/j.gca.2014.11.008
- Fouché, J., Christiansen, C. T., Lafrenière, M. J., Grogan, P., & Lamoureux, S. F. (2020).
 Canadian permafrost stores large pools of ammonium and optically distinct
 dissolved organic matter. *Nature Communications*, *11*(1), 4500.
 https://doi.org/10.1038/s41467-020-18331-w
- Fritz, M., Vonk, J. E., & Lantuit, H. (2017). Collapsing Arctic coastlines. *Nature Climate Change*. https://doi.org/10.1038/nclimate3188

Gentsch, N., Mikutta, R., Shibistova, O., Wild, B., Schnecker, J., Richter, A., ... Guggenberger,
G. (2015). Properties and bioavailability of particulate and mineral-associated organic
matter in Arctic permafrost soils, Lower Kolyma Region, Russia. *European Journal of Soil Science*, 66(4). https://doi.org/10.1111/ejss.12269

Guggenberger, G., & Zech, W. (1993). Dissolved organic carbon control in acid forest soils of
 the Fichtelgebirge (Germany) as revealed by distribution patterns and structural
 composition analyses. *Geoderma*, 59(1–4). https://doi.org/10.1016/0016-7061(93)90065-S

Guggenberger, Georg, Rodionov, A., Shibistova, O., Grabe, M., Kasansky, O. A., Fuchs, H., ...
Flessa, H. (2008). Storage and mobility of black carbon in permafrost soils of the forest
tundra ecotone in Northern Siberia. *Global Change Biology*, *14*(6), 1367–1381.
https://doi.org/10.1111/j.1365-2486.2008.01568.x

- Guo, Z., Wang, Y., Wan, Z., Zuo, Y., He, L., Li, D., ... Xu, X. (2020). Soil dissolved organic
 carbon in terrestrial ecosystems: Global budget, spatial distribution and controls. *Global Ecology and Biogeography*, 29(12). https://doi.org/10.1111/geb.13186
- Hamard, S., Robroek, B. J. M., Allard, P. M., Signarbieux, C., Zhou, S., Saesong, T., ... Jassey,
 V. E. J. (2019). Effects of Sphagnum Leachate on Competitive Sphagnum Microbiome
 Depend on Species and Time. *Frontiers in Microbiology*, *10*.
 https://doi.org/10.3389/fmicb.2019.02042
- Hansen, A. M., Kraus, T. E. C., Pellerin, B. A., Fleck, J. A., Downing, B. D., & Bergamaschi, B. A. (2016). Optical properties of dissolved organic matter (DOM): Effects of biological and photolytic degradation. *Limnology and Oceanography*, 61(3), 1015–1032. https://doi.org/10.1002/lno.10270
- 1053 Heffernan, L., Cavaco, M. A., Bhatia, M. P., Estop-Aragonés, C., Knorr, K.-H., & Olefeldt, D.

- 1054 (2022). High peatland methane emissions following permafrost thaw: enhanced acetoclastic
- 1055 methanogenesis during early successional stages. *Biogeosciences*, 19(12).
- 1056 https://doi.org/10.5194/bg-19-3051-2022
- Heffernan, L., Estop-Aragonés, C., Knorr, K.-H., Talbot, J., & Olefeldt, D. (2020). Long-term
 impacts of permafrost thaw on carbon storage in peatlands: deep losses offset by surficial
 accumulation. *Journal of Geophysical Research: Biogeosciences*, 2011(2865),
 e2019JG005501. https://doi.org/10.1029/2019JG005501
- Heslop, J. K., Chandra, S., Sobzcak, W. V, Davydov, S. P., Davydova, A. I., Spektor, V. V, &
 Anthony, K. M. W. (2017). Variable respiration rates of incubated permafrost soil extracts
 from the Kolyma River lowlands, north-east Siberia. *POLAR RESEARCH*, *36*.
 https://doi.org/10.1080/17518369.2017.1305157
- Holmes, R. M., McClelland, J. W., Peterson, B. J., Tank, S. E., Bulygina, E., Eglinton, T. I., ...
 Zimov, S. A. (2012). Seasonal and Annual Fluxes of Nutrients and Organic Matter from
 Large Rivers to the Arctic Ocean and Surrounding Seas. *ESTUARIES AND COASTS*, *35*(2),
 369–382. https://doi.org/10.1007/s12237-011-9386-6
- Hugelius, G., Strauss, J., Zubrzycki, S., Harden, J. W., Schuur, E. A. G., Ping, C. L., ... Kuhry,
 P. (2014). Estimated stocks of circumpolar permafrost carbon with quantified uncertainty
 ranges and identified data gaps. *Biogeosciences*, *11*(23), 6573–6593.
 https://doi.org/10.5194/bg-11-6573-2014
- Hugelius, Gustaf, Loisel, J., Chadburn, S., Jackson, R. B., Jones, M., MacDonald, G., ... Yu, Z.
 (2020). Large stocks of peatland carbon and nitrogen are vulnerable to permafrost thaw. *Proceedings of the National Academy of Sciences*, *117*(34), 20438–20446.
 https://doi.org/10.1073/pnas.1916387117
- Hultman, J., Waldrop, M. P., Mackelprang, R., David, M. M., McFarland, J., Blazewicz, S. J., ...
 Jansson, J. K. (2015). Multi-omics of permafrost, active layer and thermokarst bog soil
 microbiomes. *Nature*, *521*(7551). https://doi.org/10.1038/nature14238
- Jorgenson, M. T., Shur, Y. L., & Pullman, E. R. (2006). Abrupt increase in permafrost
 degradation in Arctic Alaska. *Geophysical Research Letters*, 33(2).
 https://doi.org/10.1029/2005GL024960
- Kalbitz K, Sloinger S, Park JH, Michalzik B, Matzner E (2000) Controls on the dynamics of
 dissolved organic matter in soils: a review. Soil Science, 165, 277–304.
- Kawahigashi, M., Kaiser, K., Rodionov, A., & Guggenberger, G. (2006). Sorption of dissolved
 organic matter by mineral soils of the Siberian forest tundra. *GLOBAL CHANGE BIOLOGY*, *12*(10), 1868–1877. https://doi.org/10.1111/j.1365-2486.2006.01203.x
- Keskitalo, K. H., Bröder, L., Shakil, S., Zolkos, S., Tank, S. E., van Dongen, B. E., ... Vonk, J.
 E. (2021). Downstream Evolution of Particulate Organic Matter Composition From

- Permafrost Thaw Slumps. *Frontiers in Earth Science*, 9.
 https://doi.org/10.3389/feart.2021.642675
- Kicklighter, D. W., Hayes, D. J., McClelland, J. W., Peterson, B. J., McGuire, A. D., & Melillo,
 J. M. (2013). Insights and issues with simulating terrestrial DOC loading of Arctic river
 networks. *ECOLOGICAL APPLICATIONS*, 23(8), 1817–1836. https://doi.org/10.1890/111050.1
- Kokelj, S. V., & Jorgenson, M. T. (2013). Advances in thermokarst research. *Permafrost and Periglacial Processes*, 24(2), 108–119. https://doi.org/10.1002/ppp.1779
- Lajeunesse, M. J. (2011). On the meta-analysis of response ratios for studies with correlated and
 multi-group designs. *Ecology*, 92(11). https://doi.org/10.1890/11-0423.1
- Lajtha, K., Crow, S. E., Yano, Y., Kaushal, S. S., Sulzman, E., Sollins, P., & Spears, J. D. H.
 (2005). Detrital controls on soil solution N and dissolved organic matter in soils: A field
 experiment. *Biogeochemistry*, 76(2). https://doi.org/10.1007/s10533-005-5071-9
- Lamit, L. J., Romanowicz, K. J., Potvin, L. R., Lennon, J. T., Tringe, S. G., Chimner, R. A., ...
 Lilleskov, E. A. (2021). Peatland microbial community responses to plant functional group
 and drought are depth-dependent. *Molecular Ecology*, *30*(20).
 https://doi.org/10.1111/mec.16125
- 1107 Langeveld, J., Bouwman, A. F., van Hoek, W. J., Vilmin, L., Beusen, A. H. W., Mogollón, J. M.,
- 1108 & Middelburg, J. J. (2020). Estimating dissolved carbon concentrations in global soils: a
- 1109 global database and model. *SN Applied Sciences*, 2(10), 1–21.
- 1110 https://doi.org/10.1007/s42452-020-03290-0
- Lantuit, H., Overduin, P. P., Couture, N., Wetterich, S., Aré, F., Atkinson, D., Brown, J.,
 Cherkashov, G., Drozdov, D., Forbes, D. L., & Graves-Gaylord, A. (2012). The Arctic
 coastal dynamics database: A new classification scheme and statistics on Arctic permafrost
 coastlines. *Estuaries and Coasts*, 35(2), 383–400. https://doi.org/10.1007/s12237-010-93626
- Lara, M. J., Nitze, I., Grosse, G., Martin, P., & David McGuire, A. (2018). Reduced arctic tundra productivity linked with landform and climate change interactions. *Scientific Reports*, 8(1).
 https://doi.org/10.1038/s41598-018-20692-8
- Liljedahl, A. K., Boike, J., Daanen, R. P., Fedorov, A. N., Frost, G. V., Grosse, G., ... Zona, D.
 (2016). Pan-Arctic ice-wedge degradation in warming permafrost and its influence on
 tundra hydrology. *Nature Geoscience*, 9(4). https://doi.org/10.1038/ngeo2674
- Limpens, J., Bohlin, E., & Nilsson, M. B. (2017). Phylogenetic or environmental control on the
 elemental and organo-chemical composition of Sphagnum mosses? *Plant and Soil*.
 https://doi.org/10.1007/s11104-017-3239-4

1125 Loiko, S. V, Pokrovsky, O. S., Raudina, T. V, Lim, A., Kolesnichenko, L. G., Shirokova, L. S., 1126 ... Kirpotin, S. N. (2017). Abrupt permafrost collapse enhances organic carbon, CO2, 1127 nutrient and metal release into surface waters. Chemical Geology, 471, 153-165. 1128 https://doi.org/https://doi.org/10.1016/j.chemgeo.2017.10.002 1129 Ma, Q., Jin, H., Yu, C., & Bense, V. F. (2019). Dissolved organic carbon in permafrost regions: 1130 A review. Science China Earth Sciences. https://doi.org/10.1007/s11430-018-9309-6 1131 MacDonald, E. N., Tank, S. E., Kokelj, S. V., Froese, D. G., & Hutchins, R. H. S. (2021). 1132 Permafrost-derived dissolved organic matter composition varies across permafrost end-1133 members in the western Canadian Arctic. Environmental Research Letters, 16(2). 1134 https://doi.org/10.1088/1748-9326/abd971 1135 Manies, K. L., Jones, M. C., Waldrop, M. P., Leewis, M. C., Fuller, C., Cornman, R. S., & 1136 Hoefke, K. (2021). Influence of Permafrost Type and Site History on Losses of Permafrost Carbon After Thaw. Journal of Geophysical Research: Biogeosciences, 126(11). 1137 1138 https://doi.org/10.1029/2021JG006396 1139 Marschner B, Bredow A (2002) Temperature effects on release and ecologically relevant 1140 properties of dissolved organic carbon in sterilised and biologically active soil samples. Soil 1141 Biology and Biochemistry, 34, 459–466. 1142 McGuire, A. D., Lawrence, D. M., Koven, C., Clein, J. S., Burke, E., Chen, G., ... Zhuang, Q. (2018). Dependence of the evolution of carbon dynamics in the northern permafrost region 1143 1144 on the trajectory of climate change. Proceedings of the National Academy of Sciences of the 1145 United States of America, 115(15). https://doi.org/10.1073/pnas.1719903115 1146 Mehmood, T., Liland, K. H., Snipen, L., & Sæbø, S. (2012). A review of variable selection methods in Partial Least Squares Regression. Chemometrics and Intelligent Laboratory 1147 1148 Systems. https://doi.org/10.1016/j.chemolab.2012.07.010 1149 Mevik, B. H., & Wehrens, R. (2007). The pls package: Principal component and partial least 1150 squares regression in R. Journal of Statistical Software, 18(2). 1151 https://doi.org/10.18637/jss.v018.i02 1152 Miner, K. R., Turetsky, M. R., Malina, E., Bartsch, A., Tamminen, J., McGuire, A. D., ... Miller, 1153 C. E. (2022). Permafrost carbon emissions in a changing Arctic. Nature Reviews Earth and 1154 Environment. https://doi.org/10.1038/s43017-021-00230-3 1155 Mohammed, A. A., Guimond, J. A., Bense, V. F., Jamieson, R. C., McKenzie, J. M., & Kurylyk, 1156 B. L. (2022). Mobilization of subsurface carbon pools driven by permafrost thaw and reactivation of groundwater flow: a virtual experiment. Environmental Research Letters, 1157 1158 17(12), 124036. https://doi.org/10.1088/1748-9326/ACA701 1159 Monteux, S., Weedon, J. T., Blume-Werry, G., Gavazov, K., Jassey, V. E. J., Johansson, M., ... 1160 Dorrepaal, E. (2018). Long-term in situ permafrost thaw effects on bacterial communities

- 1161
 and potential aerobic respiration. ISME Journal, 12(9), 2129–2141.

 1162
 https://doi.org/10.1038/s41396-018-0176-z
- Moore, T. R., & Dalva, M. (2001). Some controls on the release of dissolved organic carbon by
 plant tissues and soils. *Soil Science*, *166*(1), 38–47. https://doi.org/10.1097/00010694200101000-00007
- Neff, J. C., & Hooper, D. U. (2002). Vegetation and climate controls on potential CO2, DOC and
 DON production in northern latitude soils. *Global Change Biology*, 8(9), 872–884.
 https://doi.org/10.1046/j.1365-2486.2002.00517.x
- O'Donnell, J. A., Aiken, G. R., Butler, K. D., Guillemette, F., Podgorski, D. C., & Spencer, R.
 G. M. (2016). DOM composition and transformation in boreal forest soils: The effects of
 temperature and organic-horizon decomposition state. *Journal of Geophysical Research: Biogeosciences*, *121*(10), 2727–2744. https://doi.org/10.1002/2016JG003431.Received
- Olefeldt, D., Heffernan, L., Jones, M. C., Sannel, A. B. K., Treat, C. C., & Turetsky, M. R.
 (2021). Permafrost Thaw in Northern Peatlands: Rapid Changes in Ecosystem and Landscape Functions (pp. 27–67). https://doi.org/10.1007/978-3-030-71330-0_3
- Olefeldt, D., & Roulet, N. T. (2012). Effects of permafrost and hydrology on the composition
 and transport of dissolved organic carbon in a subarctic peatland complex. *Journal of Geophysical Research: Biogeosciences*, *117*(1). https://doi.org/10.1029/2011JG001819
- Olefeldt, D., & Roulet, N. T. (2014). Permafrost conditions in peatlands regulate magnitude,
 timing, and chemical composition of catchment dissolved organic carbon export. *GLOBAL CHANGE BIOLOGY*, 20(10), 3122–3136. https://doi.org/10.1111/gcb.12607
- Olefeldt, D., Roulet, N. T., Bergeron, O., Crill, P., Bäckstrand, K., & Christensen, T. R. (2012).
 Net carbon accumulation of a high-latitude permafrost palsa mire similar to permafrost-free peatlands. *Geophysical Research Letters*, *39*(3). https://doi.org/10.1029/2011GL050355
- Olefeldt, D., Goswami, S., Grosse, G., Hayes, D., Hugelius, G., Kuhry, P., ... Turetsky, M. R.
 (2016). Circumpolar distribution and carbon storage of thermokarst landscapes. Nature
 Communications, 7, 13043. https://doi.org/10.1038/ncomms13043
- Olson, D. M., Dinerstein, E., Wikramanayake, E. D., Burgess, N. D., Powell, G. V. N.,
 Underwood, E. C., ... others. (2001). Terrestrial Ecoregions of the World: A New Map of
 Life on Earth: A new global map of terrestrial ecoregions provides an innovative tool for
 conserving biodiversity. *BioScience*, *51*(11).
- Panneer Selvam, B., Lapierre, J.-F., Guillemette, F., Voigt, C., Lamprecht, R. E., Biasi, C., ...
 Berggren, M. (2017). Degradation potentials of dissolved organic carbon (DOC) from
 thawed permafrost peat. *SCIENTIFIC REPORTS*, *7*, 45811.
- 1195 https://doi.org/10.1038/srep45811

- Parmentier, FJ.W., Christensen, T.R., Rysgaard, S. et al. A synthesis of the arctic terrestrial and
 marine carbon cycles under pressure from a dwindling cryosphere. Ambio 46 (Suppl 1), 53–
 69 (2017). https://doi.org/10.1007/s13280-016-0872-8
- Payandi-Rolland, D., Shirokova, L. S., Tesfa, M., Bénézeth, P., Lim, A. G., Kuzmina, D., ...
 Pokrovsky, O. S. (2020). Dissolved organic matter biodegradation along a hydrological
 continuum in permafrost peatlands. *Science of The Total Environment*, 749, 141463.
 https://doi.org/10.1016/J.SCITOTENV.2020.141463
- Peacock, M., Futter, M. N., Jutterström, S., Kothawala, D. N., Moldan, F., Stadmark, J., &
 Evans, C. D. (2022). Three Decades of Changing Nutrient Stoichiometry from Source to
 Sea on the Swedish West Coast. *Ecosystems*, 25(8). https://doi.org/10.1007/s10021-02200798-x
- Pries, C. E. H., Schuur, E. A. G., & Crummer, K. G. (2012). Holocene Carbon Stocks and
 Carbon Accumulation Rates Altered in Soils Undergoing Permafrost Thaw. *Ecosystems*,
 15(1). https://doi.org/10.1007/s10021-011-9500-4
- Prokushkin, A. S., Gavrilenko, I. V., Abaimov, A. P., Prokushkin, S. G., & Samusenko, A. V.
 (2006). Dissolved organic carbon in upland forested watersheds underlain by continuous
 permafrost in Central Siberia. *Mitigation and Adaptation Strategies for Global Change*, *11*(1), 223–240. https://doi.org/10.1007/s11027-006-1022-6
- Prokushkin, A S, Kajimoto, T., Prokushkin, S. G., McDowell, W. H., Abaimov, A. P., &
 Matsuura, Y. (2005). Climatic factors influencing fluxes of dissolved organic carbon from
 the forest floor in a continuous-permafrost Siberian watershed. *CANADIAN JOURNAL OF FOREST RESEARCH*, 35(9), 2130–2140. https://doi.org/10.1139/X05-150
- Prokushkin, Anatoly S., Kawahigashi, M., & Tokareva, I. V. (2008). Global Warming and
 Dissolved Organic Carbon Release from Permafrost Soils. In *Permafrost Soils* (pp. 237–
 250). https://doi.org/10.1007/978-3-540-69371-0_16
- Quinton, W. L., Hayashi, M., & Chasmer, L. E. (2011). Permafrost-thaw-induced land-cover
 change in the Canadian subarctic: Implications for water resources. *Hydrological Processes*,
 25(1), 152–158. https://doi.org/10.1002/hyp.7894
- 1224 Quinton, W. L., Hayashi, M., & Pietroniro, A. (2003). Connectivity and storage functions of
 1225 channel fens and flat bogs in northern basins. *Hydrological Processes*.
 1226 https://doi.org/10.1002/hyp.1369
- Rantanen, M., Karpechko, A., Lipponen, A., Nordling, K., Hyvärinen, O., Ruosteenoja, K., ...
 Laaksonen, A. (2021). The Arctic has warmed four times faster than the globe since 1980. *Nature Portfolio*, (2022), 0–29. https://doi.org/https://doi.org/10.1038/s43247-022-00498-3
- Raymond, P. A., McClelland, J. W., Holmes, R. M., Zhulidov, A. V., Mull, K., Peterson, B. J.,
 ... Gurtovaya, T. Y. (2007). Flux and age of dissolved organic carbon exported to the Arctic

- 1232 Ocean: A carbon isotopic study of the five largest arctic rivers. *Global Biogeochemical* 1233 *Cycles*, *21*(4). https://doi.org/10.1029/2007GB002934
- Ripley, B., Venables, B., Bates, D. M., Hornik, K., Gebhardt, A., & Firth, D. (2019). Package
 'MASS' (Version 7.3-51.4). *Cran-R Project*.
- Schaefer, K., Lantuit, H., Romanovsky, V. E., Schuur, E. A. G., & Witt, R. (2014). The impact
 of the permafrost carbon feedback on global climate. *Environmental Research Letters*.
 https://doi.org/10.1088/1748-9326/9/8/085003
- Schuur, E. A. G., Abbott, B. W., Commane, R., Ernakovich, J., Euskirchen, E., Hugelius, G.,
 Grosse, G., Jones, M., Koven, C., Leshyk, V., Lawrence, D., Loranty, M. M., Mauritz, M.,
 Olefeldt, D., Natali, S., Rodenhizer, H., Salmon, V., Schädel, C., Strauss, J., ... Turetsky,
 M. (2022). Permafrost and climate change: Carbon cycle feedbacks from the warming
 arctic. Annual Review of Environment and Resources, 47, 343–371.
- Schuur, E. A. G., Bracho, R., Celis, G., Belshe, E. F., Ebert, C., Ledman, J., ... Webb, E. E.
 (2021). Tundra Underlain By Thawing Permafrost Persistently Emits Carbon to the
 Atmosphere Over 15 Years of Measurements. *Journal of Geophysical Research: Biogeosciences*, *126*(6), 1–23. https://doi.org/10.1029/2020jg006044
- Schuur, T., McGuire, A. D., Romanovsky, V., Schädel, C., & Mack, M. (2018). Chapter 11:
 Arctic and Boreal Carbon. Second State of the Carbon Cycle Report. Second State of the
 Carbon Cycle Report (SOCCR2): A Sustained Assessment Report, 428–468. Retrieved from
 https://carbon2018.globalchange.gov/chapter/11/
- Selvam, B. P., Lapierre, J.-F., Guillemette, F., Voigt, C., Lamprecht, R. E., Biasi, C., ...
 Berggren, M. (2017). Degradation potentials of dissolved organic carbon (DOC) from
 thawed permafrost peat. *SCIENTIFIC REPORTS*, 7. https://doi.org/10.1038/srep45811
- Semenchuk, P. R., Elberling, B., Amtorp, C., Winkler, J., Rumpf, S., Michelsen, A., & Cooper,
 E. J. (2015). Deeper snow alters soil nutrient availability and leaf nutrient status in high
 Arctic tundra. *Biogeochemistry*, *124*(1–3), 81–94. https://doi.org/10.1007/s10533-0150082-7
- Shakil, S., Tank, S. E., Kokelj, S. V., Vonk, J. E., & Zolkos, S. (2020). Particulate dominance of
 organic carbon mobilization from thaw slumps on the Peel Plateau, NT: Quantification and
 implications for stream systems and permafrost carbon release. *Environmental Research Letters*, 15(11). https://doi.org/10.1088/1748-9326/abac36
- Sobek, S., Tranvik, L. J., Prairie, Y. T., Kortelainen, P., & Cole, J. J. (2007). Patterns and
 regulation of dissolved organic carbon: An analysis of 7,500 widely distributed lakes. *Limnology and Oceanography*, 52(3). https://doi.org/10.4319/lo.2007.52.3.1208
- Speetjens, N. J., Tanski, G., Martin, V., Wagner, J., Richter, A., Hugelius, G., ... Vonk, J. E.
 (2022). Dissolved organic matter characterization in soils and streams in a small coastal

- low-arctic catchment. *Biogeosciences*, *19*(July), 3073–3097. Retrieved from
 https://doi.org/10.5194/bg-19-3073-2022
- Stolpmann, L., Coch, C., Morgenstern, A., Boike, J., Fritz, M., Herzschuh, U., ... Grosse, G.
 (2021). First pan-Arctic assessment of dissolved organic carbon in lakes of the permafrost region. *BIOGEOSCIENCES*, *18*(12), 3917–3936. https://doi.org/10.5194/bg-18-3917-2021
- Strauss, J., Laboor, S., Schirrmeister, L., Fedorov, A. N., Fortier, D., Froese, D., ... Grosse, G.
 (2021). Circum-Arctic Map of the Yedoma Permafrost Domain. *Frontiers in Earth Science*,
 9. https://doi.org/10.3389/feart.2021.758360
- Striegl, R. G., Aiken, G. R., Dornblaser, M. M., Raymond, P. A., & Wickland, K. P. (2005). A
 decrease in discharge-normalized DOC export by the Yukon River during summer through
 autumn. *GEOPHYSICAL RESEARCH LETTERS*, *32*(21).
 https://doi.org/10.1029/2005GL024413
- Tank, S. E., Frey, K. E., Striegl, R. G., Raymond, P. A., Holmes, R. M., McClelland, J. W., &
 Peterson, B. J. (2012). Landscape-level controls on dissolved carbon flux from diverse
 catchments of the circumboreal. *GLOBAL BIOGEOCHEMICAL CYCLES*, 26.
 https://doi.org/10.1029/2012GB004299
- Tanski, G., Wagner, D., Knoblauch, C., Fritz, M., Sachs, T., & Lantuit, H. (2019). Rapid CO2
 Release From Eroding Permafrost in Seawater. *Geophysical Research Letters*, 46(20).
 https://doi.org/10.1029/2019GL084303
- Tanski, George, Bröder, L., Wagner, D., Knoblauch, C., Lantuit, H., Beer, C., ... Vonk, J. E.
 (2021). Permafrost Carbon and CO2 Pathways Differ at Contrasting Coastal Erosion Sites
 in the Canadian Arctic. *Frontiers in Earth Science*, *9*.
 https://doi.org/10.3389/feart.2021.630493
- Textor, S. R., Wickland, K. P., Podgorski, D. C., Johnston, S. E., & Spencer, R. G. M. (2019).
 Dissolved Organic Carbon Turnover in Permafrost-Influenced Watersheds of Interior
 Alaska: Molecular Insights and the Priming Effect. *FRONTIERS IN EARTH SCIENCE*, 7.
 https://doi.org/10.3389/feart.2019.00275
- Tfaily, M. M., Hamdan, R., Corbett, J. E., Chanton, J. P., Glaser, P. H., & Cooper, W. T. (2013).
 Investigating dissolved organic matter decomposition in northern peatlands using
 complimentary analytical techniques. *Geochimica et Cosmochimica Acta*.
 https://doi.org/10.1016/j.gca.2013.03.002
- 1299 Thurman, E. M. (1985). Organic geochemistry of natural waters (Vol. 2). Springer Science &1300 Business Media.
- Treat, C. C., Jones, M. C., Camill, P., Gallego-Sala, A., Garneau, M., Harden, J. W., ...
 Väliranta, M. (2016). Effects of permafrost aggradation on peat properties as determined
 from a pan-Arctic synthesis of plant macrofossils. *Journal of Geophysical Research:*

- 1304 Biogeosciences, 121(1), 78–94. https://doi.org/10.1002/2015JG003061
- Treat, Claire C., & Jones, M. C. (2018). Near-surface permafrost aggradation in Northern
 Hemisphere peatlands shows regional and global trends during the past 6000 years.
 Holocene. https://doi.org/10.1177/0959683617752858
- Turetsky, M. R., Wieder, R. K., Vitt, D. H., Evans, R. J., & Scott, K. D. (2007). The
 disappearance of relict permafrost in boreal north America: Effects on peatland carbon
- 1310 storage and fluxes. *Global Change Biology*, *13*(9), 1922–1934.
- 1311 https://doi.org/10.1111/j.1365-2486.2007.01381.x
- Turetsky, Merritt R., Abbott, B. W., Jones, M. C., Anthony, K. W., Olefeldt, D., Schuur, E. A.
 G., ... McGuire, A. D. (2020). Carbon release through abrupt permafrost thaw. *Nature Geoscience*. https://doi.org/10.1038/s41561-019-0526-0
- 1315 USDA. (1999). Soil Taxonomy: A Basic System of Soil Classification for Making and
 1316 Interpreting Soil Surveys, 2nd Edition. Landscape and Land Capacity.
- Varner, R. K., Crill, P. M., Frolking, S., McCalley, C. K., Burke, S. A., Chanton, J. P., ... Palace,
 M. W. (2022). Permafrost thaw driven changes in hydrology and vegetation cover increase
 trace gas emissions and climate forcing in Stordalen Mire from 1970 to 2014. *Philosophical Transactions of the Royal Society A: Mathematical, Physical and Engineering Sciences*,
 380(2215). https://doi.org/10.1098/rsta.2021.0022
- 1322 Vonk, J E, Tank, S. E., Mann, P. J., Spencer, R. G. M., Treat, C. C., Striegl, R. G., ... Wickland,
 1323 K. P. (2015). Biodegradability of dissolved organic carbon in permafrost soils and aquatic
 1324 systems: a meta-analysis. *BIOGEOSCIENCES*, *12*(23), 6915–6930.
 1325 https://doi.org/10.5194/bg-12-6915-2015
- 1326 Vonk, Jorien E., & Gustafsson, Ö. (2013). Permafrost-carbon complexities. *Nature Geoscience*.
 1327 https://doi.org/10.1038/ngeo1937
- 1328 Vonk, J. E., Mann, P. J., Davydov, S., Davydova, A., Spencer, R. G. M., Schade, J., ... Holmes,
 1329 R. M. (2013). High biolability of ancient permafrost carbon upon thaw. *GEOPHYSICAL*1330 *RESEARCH LETTERS*, 40(11), 2689–2693. https://doi.org/10.1002/grl.50348
- Wang JA, Sulla-Menashe D, Woodcock CE, Sonnentag O, Keeling RF, Friedl MA. Extensive
 land cover change across Arctic–Boreal Northwestern North America from disturbance and
 climate forcing. Glob Change Biol. 2020; 26: 807–822. https://doi.org/10.1111/gcb.14804
- Weishaar, J. L., Aiken, G. R., Bergamaschi, B. A., Fram, M. S., Fujii, R., & Mopper, K. (2003).
 Evaluation of specific ultraviolet absorbance as an indicator of the chemical composition
 and reactivity of dissolved organic carbon. Environmental Science and Technology, 37(20),
 4702–4708. https://doi.org/10.1021/es030360x
- 1338 Weyhenmeyer, G. A., Fröberg, M., Karltun, E., Khalili, M., Kothawala, D., Temnerud, J., &

1339 Tranvik, L. J. (2012). Selective decay of terrestrial organic carbon during transport from

- 1340 land to sea. *Global Change Biology*, 18(1). https://doi.org/10.1111/j.1365-
- 1341 2486.2011.02544.x
- Wickland, K.P., Neff, J. C., & Aiken, G. R. (2007). Dissolved organic carbon in Alaskan boreal
 forest: Sources, chemical characteristics, and biodegradability. *Ecosystems*, 10(8), 1323–
 1340. https://doi.org/10.1007/s10021-007-9101-4
- Wickland, Kimberly P, Waldrop, M. P., Aiken, G. R., Koch, J. C., Jorgenson, Mt., & Striegl, R.
 G. (2018). Dissolved organic carbon and nitrogen release from boreal Holocene permafrost and seasonally frozen soils of Alaska. *ENVIRONMENTAL RESEARCH LETTERS*, *13*(6).
 https://doi.org/10.1088/1748-9326/aac4ad
- Wild, B., Andersson, A., Broder, L., Vonk, J., Hugelius, G., McClelland, J. W., ... Gustafsson,
 O. (2019). Rivers across the Siberian Arctic unearth the patterns of carbon release from
 thawing permafrost. *PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES OF THE UNITED STATES OF AMERICA*, *116*(21), 10280–10285.
- 1353 https://doi.org/10.1073/pnas.1811797116
- Wild, B., Gentsch, N., Capek, P., Diáková, K., Alves, R. J. E., Bárta, J., ... Richter, A. (2016).
 Plant-derived compounds stimulate the decomposition of organic matter in arctic permafrost soils. *Scientific Reports*, 6. https://doi.org/10.1038/srep25607
- Wild, B., Schnecker, J., Bárta, J., Čapek, P., Guggenberger, G., Hofhansl, F., ... Richter, A.
 (2013). Nitrogen dynamics in Turbic Cryosols from Siberia and Greenland. *Soil Biology and Biochemistry*, 67, 85–93. https://doi.org/https://doi.org/10.1016/j.soilbio.2013.08.004
- 1360 Woo, M. (1986). Permafrost hydrology in north america. *Atmosphere Ocean*, 24(3).
 1361 https://doi.org/10.1080/07055900.1986.9649248
- 1362
- 1363
- 1364
- 10.55
- 1365
- 1366
- 1367
- 1368
- 1369

1	3	7	0
T	J	1	υ

1371

1372 Studies used to generate database

- Abbott, B. W., Jones, J. B., Godsey, S. E., Larouche, J. R., & Bowden, W. B. (2015). Patterns
 and persistence of hydrologic carbon and nutrient export from collapsing upland permafrost. *Biogeosciences*, *12*(12), 3725–3740. https://doi.org/10.5194/bg-12-3725-2015
- Abbott, B. W., Larouche, J. R., Jones, J. J. B., Bowden, W. B., & Balser, A. W. (2014). From
 Thawing and Collapsing Permafrost. *Journal of Geophysical Research: Biogeosciences*, *119*, 2049–2063. https://doi.org/10.1002/2014JG002678.Received
- Beckebanze, L., Runkle, B. R. K., Walz, J., Wille, C., Holl, D., Helbig, M., ... Kutzbach, L.
 (2022). Lateral carbon export has low impact on the net ecosystem carbon balance of a
 polygonal tundra catchment. *BIOGEOSCIENCES*, *19*(16), 3863–3876.
 https://doi.org/10.5194/bg-19-3863-2022
- Boddy, E., Roberts, P., Hill, P. W., Farrar, J., & Jones, D. L. (2008). Turnover of low molecular
 weight dissolved organic C (DOC) and microbial C exhibit different temperature
 sensitivities in Arctic tundra soils. *SOIL BIOLOGY & BIOCHEMISTRY*, 40(7), 1557–1566.
 https://doi.org/10.1016/j.soilbio.2008.01.030
- Bristol, E. M., Connolly, C. T., Lorenson, T. D., Richmond, B. M., Ilgen, A. G., Choens, R. C.,
 McClelland, J. W. (2021). Geochemistry of Coastal Permafrost and Erosion-Driven
 Organic Matter Fluxes to the Beaufort Sea Near Drew Point, Alaska. *Frontiers in Earth Science*, 8. https://doi.org/10.3389/feart.2020.598933
- Bruhn, A. D., Stedmon, C. A., Comte, J., Matsuoka, A., Speetjens, N. J., Tanski, G., ... Sjöstedt,
 J. (2021). Terrestrial Dissolved Organic Matter Mobilized From Eroding Permafrost
 Controls Microbial Community Composition and Growth in Arctic Coastal Zones. *Frontiers in Earth Science*, 9. https://doi.org/10.3389/feart.2021.640580
- Buckeridge, K. M., & Grogan, P. (2008). Deepened snow alters soil microbial nutrient
 limitations in arctic birch hummock tundra. *Applied Soil Ecology*, *39*(2), 210–222.
 https://doi.org/https://doi.org/10.1016/j.apsoil.2007.12.010
- Burd, K., Estop-Aragonés, C., Tank, S. E., & Olefeldt, D. (2020). Lability of dissolved organic
 carbon from boreal peatlands: interactions between permafrost thaw, wildfire, and season. *Canadian Journal of Soil Science*, *13*(February), 1–13. https://doi.org/10.1139/cjss-20190154

<sup>Burd, K., Tank, S. E., Dion, N., Quinton, W. L., Spence, C., Tanentzap, A. J., & Olefeldt, D.
(2018). Seasonal shifts in export of DOC and nutrients from burned and unburned peatland-</sup>

- rich catchments, Northwest Territories, Canada. *Hydrology and Earth System Sciences*,
 4455–4472. https://doi.org/10.5194/hess-22-4455-2018
- Carey, S. K. (2003). Dissolved organic carbon fluxes in a discontinuous permafrost subarctic
 alpine catchment. *PERMAFROST AND PERIGLACIAL PROCESSES*, *14*(2), 161–171.
 https://doi.org/10.1002/ppp.444
- Chiasson-Poirier, G., Franssen, J., Lafreniere, M. J., Fortier, D., & Lamoureux, S. F. (2020).
 Seasona evolution of active layer thaw depth and hillslope-stream connectivity in a
 permafrost watershed. *WATER RESOURCES RESEARCH*, 56(1).
- 1412 https://doi.org/10.1029/2019WR025828
- 1413 Connolly, C. T., Cardenas, M. B., Burkart, G. A., Spencer, R. G. M., & McClelland, J. W.
 1414 (2020). Groundwater as a major source of dissolved organic matter to Arctic coastal waters.
 1415 NATURE COMMUNICATIONS, 11(1). https://doi.org/10.1038/s41467-020-15250-8
- Cory, R. M., Crump, B. C., Dobkowski, J. A., & Kling, G. W. (2013). Surface exposure to
 sunlight stimulates CO 2 release from permafrost soil carbon in the Arctic. *Proceedings of the National Academy of Sciences*, *110*(9), 3429–3434.
- 1419 https://doi.org/10.1073/pnas.1214104110
- 1420 Deshpande, B. N., Crevecoeur, S., Matveev, A., & Vincent, W. F. (2016). Bacterial production
 1421 in subarctic peatland lakes enriched by thawing permafrost. *BIOGEOSCIENCES*, *13*(15),
 1422 4411–4427. https://doi.org/10.5194/bg-13-4411-2016
- Douglas, T. A., Fortier, D., Shur, Y. L., Kanevskiy, M. Z., Guo, L., Cai, Y., & Bray, M. T.
 (2011). Biogeochemical and Geocryological Characteristics of Wedge and ThermokarstCave Ice in the CRREL Permafrost Tunnel, Alaska. *PERMAFROST AND PERIGLACIAL PROCESSES*, 22(2), 120–128. https://doi.org/10.1002/ppp.709
- 1427 Drake, T. W., Wickland, K. P., Spencer, R. G. M., McKnight, D. M., & Striegl, R. G. (2015).
 1428 Ancient low-molecular-weight organic acids in permafrost fuel rapid carbon dioxide
 1429 production upon thaw. *PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES*1430 *OF THE UNITED STATES OF AMERICA*, *112*(45), 13946–13951.
 1431 https://doi.org/10.1073/pnas.1511705112
- 1432 Dutta, K., Schuur, E. A. G., Neff, J. C., & Zimov, S. A. (2006). Potential carbon release from
 1433 permafrost soils of Northeastern Siberia. *GLOBAL CHANGE BIOLOGY*, *12*(12), 2336–
 1434 2351. https://doi.org/10.1111/j.1365-2486.2006.01259.x
- Edwards, K. A., & Jefferies, R. L. (2013). Inter-annual and seasonal dynamics of soil microbial
 biomass and nutrients in wet and dry low-Arctic sedge meadows. *Soil Biology and Biochemistry*, 57, 83–90. https://doi.org/https://doi.org/10.1016/j.soilbio.2012.07.018
- Edwards, K. A., McCulloch, J., Kershaw], G. [Peter, & Jefferies, R. L. (2006). Soil microbial
 and nutrient dynamics in a wet Arctic sedge meadow in late winter and early spring. *Soil*

1440 *Biology and Biochemistry*, *38*(9), 2843–2851.

1441 https://doi.org/https://doi.org/10.1016/j.soilbio.2006.04.042

- Ernakovich, J. G., Lynch, L. M., Brewer, P. E., Calderon, F. J., & Wallenstein, M. D. (2017).
 Redox and temperature-sensitive changes in microbial communities and soil chemistry
 dictate greenhouse gas loss from thawed permafrost. *BIOGEOCHEMISTRY*, *134*(1–2),
 183–200. https://doi.org/10.1007/s10533-017-0354-5
- 1445 105 200. https://doi.org/10.100//310555/01/ 0554/5
- Ewing, S. A., Paces, J. B., O'Donnell, J. A., Jorgenson, M. T., Kanevskiy, M. Z., Aiken, G. R.,
 Striegl, R. (2015). Uranium isotopes and dissolved organic carbon in loess permafrost:
 Modeling the age of ancient ice. *GEOCHIMICA ET COSMOCHIMICA ACTA*, *152*, 143–
 https://doi.org/10.1016/j.gca.2014.11.008

Fenger-Nielsen, R., Hollesen, J., Matthiesen, H., Andersen, E. A. S., Westergaard-Nielsen, A.,
Harmsen, H., ... Elberling, B. (2019). Footprints from the past: The influence of past human
activities on vegetation and soil across five archaeological sites in Greenland. *Science of the Total Environment*, 654, 895–905. https://doi.org/10.1016/j.scitotenv.2018.11.018

- Fouché, J., Christiansen, C. T., Lafrenière, M. J., Grogan, P., & Lamoureux, S. F. (2020).
 Canadian permafrost stores large pools of ammonium and optically distinct
 dissolved organic matter. *Nature Communications*, *11*(1), 4500.
 https://doi.org/10.1029/c41467.020.18221.ml
- 1457 https://doi.org/10.1038/s41467-020-18331-w
- Fouche, J., Bouchez, C., Keller, C., Allard, M., & Ambrosi, J.-P. (2021). Seasonal cryogenic
 processes control supra-permafrost pore water chemistry in two contrasting Cryosols. *GEODERMA*, 401. https://doi.org/10.1016/j.geoderma.2021.115302
- Fouché, J., Keller, C., Allard, M., Ambrosi, J. P., Fouche, J., Keller, C., ... Ambrosi, J. P. (2014).
 Increased CO2 fluxes under warming tests and soil solution chemistry in Histic and Turbic
 Cryosols, Salluit, Nunavik, Canada. *Soil Biology and Biochemistry*, 68, 185–199.
 https://doi.org/https://doi.org/10.1016/j.soilbio.2013.10.007
- Fritz, M., Opel, T., Tanski, G., Herzschuh, U., Meyer, H., Eulenburg, A., & Lantuit, H. (2015).
 Dissolved organic carbon (DOC) in Arctic ground ice. *CRYOSPHERE*, 9(2), 737–752.
 https://doi.org/10.5194/tc-9-737-2015
- Gagné, K. R., Ewers, S. C., Murphy, C. J., Daanen, R., Walter Anthony, K., & Guerard, J. J.
 (2020). Composition and photo-reactivity of organic matter from permafrost soils and surface waters in interior Alaska. *Environmental Science: Processes and Impacts*, 22(7), 1525–1539. https://doi.org/10.1039/d0em00097c
- Gao, L., Zhou, Z., Reyes V, A., & Guo, L. (2018). Yields and Characterization of Dissolved
 Organic Matter From Different Aged Soils in Northern Alaska. *JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES*, *123*(7), 2035–2052.
 https://doi.org/10.1020/2018JC004408
- 1475 https://doi.org/10.1029/2018JG004408

1476 Herndon, E. M., Mann, B. F., Chowdhury, T. R., Yang, Z., Wullschleger, S. D., Graham, D., ... 1477 Gu, B. (2015). Pathways of anaerobic organic matter decomposition in tundra soils from 1478 Barrow, Alaska. JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES, 1479 120(11), 2345–2359. https://doi.org/10.1002/2015JG003147 1480 Herndon, E. M., Yang, Z., Bargar, J., Janot, N., Regier, T. Z., Graham, D. E., ... Liang, L. 1481 (2015). Geochemical drivers of organic matter decomposition in arctic tundra soils. BIOGEOCHEMISTRY, 126(3), 397-414. https://doi.org/10.1007/s10533-015-0165-5 1482 1483 Herndon, E., AlBashaireh, A., Singer, D., Chowdhury], T. [Roy, Gu, B., & Graham, D. (2017). 1484 Influence of iron redox cycling on organo-mineral associations in Arctic tundra soil. 1485 Geochimica et Cosmochimica Acta, 207, 210–231. 1486 https://doi.org/https://doi.org/10.1016/j.gca.2017.02.034 1487 Heslop, J. K., Chandra, S., Sobzcak, W. V, Davydov, S. P., Davydova, A. I., Spektor, V. V, & 1488 Anthony, K. M. W. (2017). Variable respiration rates of incubated permafrost soil extracts 1489 from the Kolyma River lowlands, north-east Siberia. POLAR RESEARCH, 36. 1490 https://doi.org/10.1080/17518369.2017.1305157 1491 Hirst, C., Mauclet, E., Monhonval, A., Tihon, E., Ledman, J., Schuur, E. A. G., & Opfergelt, S. 1492 (2022). Seasonal Changes in Hydrology and Permafrost Degradation Control Mineral 1493 Element-Bound DOC Transport From Permafrost Soils to Streams. GLOBAL 1494 BIOGEOCHEMICAL CYCLES, 36(2). https://doi.org/10.1029/2021GB007105 1495 Hodgkins, S. B., Tfaily, M. M., Podgorski, D. C., McCalley, C. K., Saleska, S. R., Crill, P. M., 1496 ... Cooper, W. T. (2016). Elemental composition and optical properties reveal changes in 1497 dissolved organic matter along a permafrost thaw chronosequence in a subarctic peatland. 1498 Geochimica et Cosmochimica Acta, 187, 123-140. 1499 https://doi.org/10.1016/j.gca.2016.05.015 1500 Jilkova, V., Devetter, M., Bryndova, M., Hajek, T., Kotas, P., Lulakova, P., ... Macek, P. (2021). 1501 Carbon Sequestration Related to Soil Physical and Chemical Properties in the High Arctic. 1502 GLOBAL BIOGEOCHEMICAL CYCLES, 35(9). https://doi.org/10.1029/2020GB006877 1503 Kane, E. S., Chivers, M. R., Turetsky, M. R., Treat, C. C., Petersen, D. G., Waldrop, M., ... 1504 McGuire, A. D. (2013). Response of anaerobic carbon cycling to water table manipulation 1505 in an Alaskan rich fen. Soil Biology and Biochemistry, 58, 50-60. 1506 https://doi.org/https://doi.org/10.1016/j.soilbio.2012.10.032 1507 Kane, E. S., Valentine, D. W., Michaelson, G. J., Fox, J. D., & Ping, C.-L. (2006). Controls over 1508 pathways of carbon efflux from soils along climate and black spruce productivity gradients 1509 in interior Alaska. Soil Biology and Biochemistry, 38(6), 1438–1450. 1510 https://doi.org/https://doi.org/10.1016/j.soilbio.2005.11.004 1511 Kane, E. S., Turetsky, M. R., Harden, J. W., McGuire, A. D., & Waddington, J. M. (2010). 1512 Seasonal ice and hydrologic controls on dissolved organic carbon and nitrogen

- 1513 concentrations in a boreal-rich fen. JOURNAL OF GEOPHYSICAL RESEARCH 1514 BIOGEOSCIENCES, 115. https://doi.org/10.1029/2010JG001366
- Kawahigashi, M., Prokushkin, A., & Sumida, H. (2011). Effect of fire on solute release from
 organic horizons under larch forest in Central Siberian permafrost terrain. *Geoderma*, *166*(1), 171–180. https://doi.org/https://doi.org/10.1016/j.geoderma.2011.07.027
- Koch, J. C., Runkel, R. L., Striegl, R., & McKnight, D. M. (2013). Hydrologic controls on the
 transport and cycling of carbon and nitrogen in a boreal catchment underlain by continuous
 permafrost. *JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES*, *118*(2),
 698–712. https://doi.org/10.1002/jgrg.20058
- Lim, A. G., Loiko, S. V, Kuzmina, D. M., Krickov, I. V, Shirokova, L. S., Kulizhsky, S. P., ...
 Pokrovsky, O. S. (2021). Dispersed ground ice of permafrost peatlands: Potential
 unaccounted carbon, nutrient and metal sources. *Chemosphere*, 266, 128953.
 https://doi.org/10.1016/j.chemosphere.2020.128953
- Lindborg, T., Rydberg, J., Tröjbom, M., Berglund, S., Johansson, E., Löfgren, A., ... Laudon, H.
 (2016). Biogeochemical data from terrestrial and aquatic ecosystems in a periglacial
 catchment, West Greenland. *Earth System Science Data*, 8(2), 439–459.
 https://doi.org/10.5194/essd-8-439-2016
- Littlefair, C. A., & Tank, S. E. (2018). Biodegradability of Thermokarst Carbon in a TillAssociated, Glacial Margin Landscape: The Case of the Peel Plateau, NWT, Canada. *JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES*, *123*(10), 3293–3307.
 https://doi.org/10.1029/2018JG004461
- Liu, N., Michelsen, A., & Rinnan, R. (2020). Vegetation and soil responses to added carbon and
 nutrients remain six years after discontinuation of long-term treatments. *Science of the Total Environment*, 722, 137885. https://doi.org/10.1016/j.scitotenv.2020.137885
- Loiko, S. V, Pokrovsky, O. S., Raudina, T. V, Lim, A., Kolesnichenko, L. G., Shirokova, L. S.,
 ... Kirpotin, S. N. (2017). Abrupt permafrost collapse enhances organic carbon, CO2,
 nutrient and metal release into surface waters. *Chemical Geology*, 471, 153–165.
 https://doi.org/https://doi.org/10.1016/j.chemgeo.2017.10.002
- MacDonald, E. N., Tank, S. E., Kokelj, S. V., Froese, D. G., & Hutchins, R. H. S. (2021).
 Permafrost-derived dissolved organic matter composition varies across permafrost endmembers in the western Canadian Arctic. *Environmental Research Letters*, *16*(2).
 https://doi.org/10.1088/1748-9326/abd971
- Mangal, V., DeGasparro, S., Beresford, D. V, & Guéguen, C. (2020). Linking molecular and
 optical properties of dissolved organic matter across a soil-water interface on Akimiski
 Island (Nunavut, Canada). *Science of The Total Environment*, 704, 135415.
 https://doi.org/https://doi.org/10.1016/j.scitotenv.2019.135415

1549	Masyagina, O. V, Tokareva, I. V, & Prokushkin, A. S. (2016). Post fire organic matter
1550	biodegradation in permafrost soils: Case study after experimental heating of mineral
1551	horizons. <i>Science of The Total Environment</i> , 573, 1255–1264.
1552	https://doi.org/https://doi.org/10.1016/j.scitotenv.2016.04.195
1553	McFarlane, K. J., Throckmorton, H. M., Heikoop, J. M., Newman, B. D., Hedgpeth, A. L.,
1554	Repasch, M. N., Wilson, C. J. (2022). Age and chemistry of dissolved organic carbon
1555	reveal enhanced leaching of ancient labile carbon at the permafrost thaw zone.
1556	<i>BIOGEOSCIENCES</i> , 19(4), 1211–1223. https://doi.org/10.5194/bg-19-1211-2022
1557	Mörsdorf, M. A., Baggesen, N. S., Yoccoz, N. G., Michelsen, A., Elberling, B., Ambus, P. L., &
1558	Cooper, E. J. (2019). Deepened winter snow significantly influences the availability and
1559	forms of nitrogen taken up by plants in High Arctic tundra. <i>Soil Biology and Biochemistry</i> ,
1560	135, 222–234. https://doi.org/https://doi.org/10.1016/j.soilbio.2019.05.009
1561 1562 1563	Neff, J. C., & Hooper, D. U. (2002). Vegetation and climate controls on potential CO2, DOC and DON production in northern latitude soils. <i>Global Change Biology</i> , 8(9), 872–884. https://doi.org/10.1046/j.1365-2486.2002.00517.x
1564	Nielsen, C. S., Michelsen, A., Strobel, B. W., Wulff, K., Banyasz, I., & Elberling, B. (2017).
1565	Correlations between substrate availability, dissolved CH4, and CH4 emissions in an arctic
1566	wetland subject to warming and plant removal. <i>JOURNAL OF GEOPHYSICAL</i>
1567	<i>RESEARCH-BIOGEOSCIENCES</i> , 122(3), 645–660. https://doi.org/10.1002/2016JG003511
1568 1569 1570 1571 1572	O'Donnell, J. A., Aiken, G. R., Butler, K. D., Guillemette, F., Podgorski, D. C., & Spencer, R. G. M. (2016). DOM composition and transformation in boreal forest soils: The effects of temperature and organic-horizon decomposition state. <i>JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES</i> , 121(10), 2727–2744. https://doi.org/10.1002/2016JG003431
1573	O'Donnell, J. A., Turetsky, M. R., Harden, J. W., Manies, K. L., Pruett, L. E., Shetler, G., &
1574	Neff, J. C. (2009). Interactive Effects of Fire, Soil Climate, and Moss on CO2 Fluxes in
1575	Black Spruce Ecosystems of Interior Alaska. <i>ECOSYSTEMS</i> , 12(1), 57–72.
1576	https://doi.org/10.1007/s10021-008-9206-4
1577	Oiffer, L., & Siciliano, S. D. (2009). Methyl mercury production and loss in Arctic soil. Science
1578	of the Total Environment, 407(5), 1691–1700.
1579	https://doi.org/10.1016/j.scitotenv.2008.10.025
1580 1581 1582	Olefeldt, D., & Roulet, N. T. (2012). Effects of permafrost and hydrology on the composition and transport of dissolved organic carbon in a subarctic peatland complex. <i>Journal of Geophysical Research: Biogeosciences</i> , <i>117</i> (1). https://doi.org/10.1029/2011JG001819
1583 1584 1585	Olefeldt, D., & Roulet, N. T. (2014). Permafrost conditions in peatlands regulate magnitude, timing, and chemical composition of catchment dissolved organic carbon export. <i>Global Change Biology</i> , <i>20</i> (10), 3122–3136. https://doi.org/10.1111/gcb.12607

- Olefeldt, D., Roulet, N. T., Bergeron, O., Crill, P., Bäckstrand, K., & Christensen, T. R. (2012).
 Net carbon accumulation of a high-latitude permafrost palsa mire similar to permafrost-free peatlands. *Geophysical Research Letters*. https://doi.org/10.1029/2011GL050355
- Olsrud, M., & Christensen, T. R. (2011). Carbon partitioning in a wet and a semiwet subarctic
 mire ecosystem based on in situ 14C pulse-labelling. *Soil Biology and Biochemistry*, 43(2),
 231–239. https://doi.org/10.1016/j.soilbio.2010.09.034
- Pastor, A., Poblador, S., Skovsholt, L. J., & Riis, T. (2020). Microbial carbon and nitrogen
 processes in high-Arctic riparian soils. *PERMAFROST AND PERIGLACIAL PROCESSES*, *31*(1), 223–236. https://doi.org/10.1002/ppp.2039
- Patzner, M. S., Mueller, C. W., Malusova, M., Baur, M., Nikeleit, V., Scholten, T., ... Bryce, C.
 (2020). Iron mineral dissolution releases iron and associated organic carbon during
 permafrost thaw. *Nature Communications*, 11(1), 1–11. https://doi.org/10.1038/s41467-02020102-6
- Patzner, M. S., Logan, M., McKenna, A. M., Young, R. B., Zhou, Z., Joss, H., ... Bryce, C.
 (2022). Microbial iron cycling during palsa hillslope collapse promotes greenhouse gas
 emissions before complete permafrost thaw. *Communications Earth & Environment*, 3(1),
 76. https://doi.org/10.1038/s43247-022-00407-8
- Payandi-Rolland, D., Shirokova, L. S., Tesfa, M., Bénézeth, P., Lim, A. G., Kuzmina, D., ...
 Pokrovsky, O. S. (2020). Dissolved organic matter biodegradation along a hydrological
 continuum in permafrost peatlands. *Science of The Total Environment*, 749, 141463.
 https://doi.org/10.1016/J.SCITOTENV.2020.141463
- Payandi-Rolland, D., Shirokova, L. S., Labonne, F., Bénézeth, P., & Pokrovsky, O. S. (2021).
 Impact of freeze-thaw cycles on organic carbon and metals in waters of
 permafrost peatlands. *Chemosphere*, 279, 130510.
- 1610 https://doi.org/10.1016/j.chemosphere.2021.130510
- Payandi-Rolland, D., Shirokova, L. S., Nakhle, P., Tesfa, M., Abdou, A., Causserand, C., ...
 Pokrovsky, O. S. (2020). Aerobic release and biodegradation of dissolved organic matter
 from frozen peat: Effects of temperature and heterotrophic bacteria. *CHEMICAL*
- 1614 *GEOLOGY*, *536*. https://doi.org/10.1016/j.chemgeo.2019.119448
- Petersen, D. G., Blazewicz, S. J., Firestone, M., Herman, D. J., Turetsky, M., & Waldrop, M.
 (2012). Abundance of microbial genes associated with nitrogen cycling as indices of
 biogeochemical process rates across a vegetation gradient in Alaska. *Environmental Microbiology*, 14(4), 993–1008. https://doi.org/10.1111/j.1462-2920.2011.02679.x
- Pokrovsky, O. S., Reynolds, B. C., Prokushkin, A. S., Schott, J., & Viers, J. (2013). Silicon
 isotope variations in Central Siberian rivers during basalt weathering in permafrost-
- 1621 dominated larch forests. *Chemical Geology*, 355, 103–116.
- 1622 https://doi.org/https://doi.org/10.1016/j.chemgeo.2013.07.016

1625 5659-5680. https://doi.org/10.1016/j.gca.2005.07.018 1626 Pokrovsky, O. S., Manasypov, R. M., Loiko, S. V, & Shirokova, L. S. (2016). Organic and 1627 organo-mineral colloids in discontinuous permafrost zone. Geochimica et Cosmochimica 1628 Acta, 188, 1–20. https://doi.org/https://doi.org/10.1016/j.gca.2016.05.035 1629 Poulin, B. A., Ryan, J. N., Tate, M. T., Krabbenhoft, D. P., Hines, M. E., Barkay, T., ... Aiken, 1630 G. R. (2019). Geochemical Factors Controlling Dissolved Elemental Mercury and 1631 Methylmercury Formation in Alaskan Wetlands of Varying Trophic Status. Environmental 1632 Science and Technology, 53(11), 6203-6213. https://doi.org/10.1021/acs.est.8b06041 Prokushkin, A. S., Gavrilenko, I. V, Abaimov, A. P., Prokushkin, S. G., & Samusenko, A. V. 1633 1634 (2006). Dissolved organic carbon in upland forested watersheds underlain by continuous permafrost in Central Siberia. *Mitigation and Adaptation Strategies for Global Change*, 1635 1636 11(1), 223-240. https://doi.org/10.1007/s11027-006-1022-6 1637 Prokushkin, A. S., Gleixner, G., McDowell, W. H., Ruehlow, S., & Schulze, E.-D. (2007). 1638 Source- and substrate-specific export of dissolved organic matter from permafrost-1639 dominated forested watershed in central Siberia. GLOBAL BIOGEOCHEMICAL CYCLES, 1640 21(4). https://doi.org/10.1029/2007GB002938 1641 Prokushkin, A. S., Kajimoto, T., Prokushkin, S. G., McDowell, W. H., Abaimov, A. P., & 1642 Matsuura, Y. (2005). Climatic factors influencing fluxes of dissolved organic carbon from 1643 the forest floor in a continuous-permafrost Siberian watershed. CANADIAN JOURNAL OF 1644 FOREST RESEARCH, 35(9), 2130-2140. https://doi.org/10.1139/X05-150 Rasmussen, L. H., Michelsen, A., Ladegaard-Pedersen, P., Nielsen, C. S., & Elberling, B. 1645 1646 (2020). Arctic soil water chemistry in dry and wet tundra subject to snow addition, summer 1647 warming and herbivory simulation. Soil Biology and Biochemistry, 141, 107676. 1648 https://doi.org/https://doi.org/10.1016/j.soilbio.2019.107676 1649 Raudina, T. V, Loiko, S. V, Lim, A., Manasypov, R. M., Shirokova, L. S., Istigechev, G. I., ... 1650 Pokrovsky, O. S. (2018). Permafrost thaw and climate warming may decrease the CO2, 1651 carbon, and metal concentration in peat soil waters of the Western Siberia Lowland. Science 1652 of The Total Environment, 634, 1004–1023. https://doi.org/https://doi.org/10.1016/j.scitotenv.2018.04.059 1653 1654 Raudina, T. V, Loiko, S. V, Lim, A. G., Krickov, I. V, Shirokova, L. S., Istigechev, G. I., ... 1655 Pokrovsky, O. S. (2017). Dissolved organic carbon and major and trace elements in peat porewater of sporadic, discontinuous, and continuous permafrost zones of western Siberia. 1656 BIOGEOSCIENCES, 14(14), 3561-3584. https://doi.org/10.5194/bg-14-3561-2017 1657

Pokrovsky, O. S., Schott, J., Kudryavtzev, D. I., & Dupré, B. (2005). Basalt weathering in

Central Siberia under permafrost conditions. Geochimica et Cosmochimica Acta, 69(24),

1623

1624

- Ro, H.-M., Ji, Y., & Lee, B. (2018). Interactive effect of soil moisture and temperature regimes
 on the dynamics of soil organic carbon decomposition in a subarctic tundra soil.
 GEOSCIENCES JOURNAL, 22(1), 121–130. https://doi.org/10.1007/s12303-017-0052-2
- Roehm, C. L., Giesler, R., & Karlsson, J. (2009). Bioavailability of terrestrial organic carbon to
 lake bacteria: The case of a degrading subarctic permafrost mire complex. *JOURNAL OF GEOPHYSICAL RESEARCH-BIOGEOSCIENCES*, 114.
- 1664 https://doi.org/10.1029/2008JG000863
- Rogers, J. A., Galy, V., Kellerman, A. M., Chanton, J. P., Zimov, N., & Spencer, R. G. M.
 (2021). Limited Presence of Permafrost Dissolved Organic Matter in the Kolyma River,
 Siberia Revealed by Ramped Oxidation. *JOURNAL OF GEOPHYSICAL RESEARCH- BIOGEOSCIENCES*, *126*(7). https://doi.org/10.1029/2020JG005977
- Roth, V.-N., Dittmar, T., Gaupp, R., & Gleixner, G. (2013). Latitude and pH driven trends in the
 molecular composition of DOM across a north south transect along the Yenisei River. *Geochimica et Cosmochimica Acta*, 123, 93–105.
- 1672 https://doi.org/https://doi.org/10.1016/j.gca.2013.09.002
- Schostag, M., Stibal, M., Jacobsen, C. S., Baelum, J., Tas, N., Elberling, B., ... Prieme, A.
 (2015). Distinct summer and winter bacterial communities in the active layer of Svalbard
 permafrost revealed by DNA- and RNA-based analyses. *FRONTIERS IN MICROBIOLOGY*, 6. https://doi.org/10.3389/fmicb.2015.00399
- Shakil, S., Tank, S. E., Kokelj, S. V., Vonk, J. E., & Zolkos, S. (2020). Particulate dominance of
 organic carbon mobilization from thaw slumps on the Peel Plateau, NT: Quantification and
 implications for stream systems and permafrost carbon release. *Environmental Research Letters*, 15(11). https://doi.org/10.1088/1748-9326/abac36
- Shatilla, N. J., & Carey, S. K. (2019). Assessing inter-annual and seasonal patterns of DOC and
 DOM quality across a complex alpine watershed underlain by discontinuous permafrost in
 Yukon, Canada. *Hydrology and Earth System Sciences*, 23(9), 3571–3591.
 https://doi.org/10.5194/hess-23-3571-2019
- Shirokova, L. S., Pokrovsky, O. S., Kirpotin, S. N., Desmukh, C., Pokrovsky, B. G., Audry, S.,
 & Viers, J. (2013). Biogeochemistry of organic carbon, CO2, CH4, and trace elements in
 thermokarst water bodies in discontinuous permafrost zones of Western Siberia. *BIOGEOCHEMISTRY*, *113*(1–3), 573–593. https://doi.org/10.1007/s10533-012-9790-4
- Shirokova, L. S., Bredoire, R., Rols, J.-L. L., & Pokrovsky, O. S. (2017). Moss and Peat
 Leachate Degradability by Heterotrophic Bacteria: The Fate of Organic Carbon and Trace
 Metals. *Geomicrobiology Journal*, *34*(8), 641–655.
 https://doi.org/10.1080/01490451.2015.1111470
- Shirokova, L. S., Chupakov, A. V, Zabelina, S. A., Neverova, N. V, Payandi-Rolland, D.,
 Causserand, C., ... Pokrovsky, O. S. (2019). Humic surface waters of frozen peat bogs

- 1695 (permafrost zone) are highly resistant to bio- and photodegradation. *BIOGEOSCIENCES*,
 1696 *16*(12), 2511–2526. https://doi.org/10.5194/bg-16-2511-2019
- Shirokova, L. S., Labouret, J., Gurge, M., Gerard, E., Ivanova, I. S., Zabelina, S. A., &
 Pokrovsky, O. S. (2017). Impact of Cyanobacterial Associate and Heterotrophic Bacteria on
 Dissolved Organic Carbon and Metal in Moss and Peat Leachate: Application to Permafrost
 Thaw in Aquatic Environments. *AQUATIC GEOCHEMISTRY*, 23(5–6), 331–358.
- 1701 https://doi.org/10.1007/s10498-017-9325-7
- Sistla, S. A., Schaeffer, S., & Schimel, J. P. (2019). Plant community regulates decomposer
 response to freezing more strongly than the rate or extent of the freezing regime.
 ECOSPHERE, 10(2). https://doi.org/10.1002/ecs2.2608
- Speetjens, N. J., Tanski, G., Martin, V., Wagner, J., Richter, A., Hugelius, G., ... Vonk, J. E.
 (2022). Dissolved organic matter characterization in soils and streams in a small coastal
 low-arctic catchment. *Biogeosciences*, *19*(July), 3073–3097. Retrieved from
 https://doi.org/10.5194/bg-19-3073-2022
- Stutter, M. I., & Billett, M. F. (2003). Biogeochemical controls on streamwater and soil solution
 chemistry in a High Arctic environment. *Geoderma*, *113*(1), 127–146.
 https://doi.org/https://doi.org/10.1016/S0016-7061(02)00335-X
- Takano, S., Yamashita, Y., Tei, S., Liang, M., Shingubara, R., Morozumi, T., ... Sugimoto, A.
 (2021). Stable Water Isotope Assessment of Tundra Wetland Hydrology as a Potential
 Source of Arctic Riverine Dissolved Organic Carbon in the Indigirka River Lowland,
 Northeastern Siberia. *Frontiers in Earth Science*, 9.
- 1716 https://doi.org/10.3389/feart.2021.699365
- Tanski, G., Couture, N., Lantuit, H., Eulenburg, A., & Fritz, M. (2016). Eroding permafrost
 coasts release low amounts of dissolved organic carbon (DOC) from ground ice into the
 nearshore zone of the Arctic Ocean. *Global Biogeochemical Cycles*, *30*(7), 1054–1068.
 https://doi.org/10.1002/ 2015GB005337
- Tanski, G., Lantuit, H., Ruttor, S., Knoblauch, C., Radosavljevic, B., Strauss, J., ... Fritz, M.
 (2017). Transformation of terrestrial organic matter along thermokarst-affected permafrost
 coasts in the Arctic. *Science of the Total Environment*, *581–582*, 434–447.
 https://doi.org/10.1016/j.scitotenv.2016.12.152
- Textor, S. R., Wickland, K. P., Podgorski, D. C., Johnston, S. E., & Spencer, R. G. M. (2019).
 Dissolved Organic Carbon Turnover in Permafrost-Influenced Watersheds of Interior
 Alaska: Molecular Insights and the Priming Effect. *FRONTIERS IN EARTH SCIENCE*, 7.
 https://doi.org/10.3389/feart.2019.00275
- Thompson, M. S., Giesler, R., Karlsson, J., & Klaminder, J. (2015). Size and characteristics of
 the DOC pool in near-surface subarctic mire permafrost as a potential source for nearby

- 1731 freshwaters. *Arctic, Antarctic, and Alpine Research*, 47(1), 49–58.
 1732 https://doi.org/10.1657/AAAR0014-010
- Treat, C. C., Wollheim, W. M., Varner, R. K., & Bowden, W. B. (2016). Longer thaw seasons
 increase nitrogen availability for leaching during fall in tundra soils. *ENVIRONMENTAL RESEARCH LETTERS*, *11*(6). https://doi.org/10.1088/1748-9326/11/6/064013
- Trusiak, A., Treibergs, L. A., Kling, G. W., & Cory, R. M. (2018). The role of iron and reactive
 oxygen species in the production of CO2 in arctic soil waters. *GEOCHIMICA ET COSMOCHIMICA ACTA*, 224, 80–95. https://doi.org/10.1016/j.gca.2017.12.022
- Voigt, C., Lamprecht, R. E., Marushchak, M. E., Lind, S. E., Novakovskiy, A., Aurela, M., ...
 Biasi, C. (2017). Warming of subarctic tundra increases emissions of all three important
 greenhouse gases carbon dioxide, methane, and nitrous oxide. *Global Change Biology*,
 23(8), 3121–3138. https://doi.org/10.1111/gcb.13563
- 1743 Voigt, C., Marushchak, M. E., Mastepanov, M., Lamprecht, R. E., Christensen, T. R.,
 1744 Dorodnikov, M., ... Biasi, C. (2019). Ecosystem carbon response of an Arctic peatland to
 1745 simulated permafrost thaw. *Global Change Biology*, 25(5), 1746–1764.
 1746 https://doi.org/10.1111/gcb.14574
- 1747 Voigt, C., Marushchak, M. E., Lamprecht, R. E., Jackowicz-Korczyński, M., Lindgren, A.,
 1748 Mastepanov, M., ... Biasi, C. (2017). Increased nitrous oxide emissions from Arctic
 1749 peatlands after permafrost thaw. *Proceedings of the National Academy of Sciences of the*1750 United States of America, 114(24), 6238–6243. Retrieved from
 1751 https://www.jstor.org/stable/26484198
- 1752 Vonk, J. E., Mann, P. J., Dowdy, K. L., Davydova, A., Davydov, S. P., Zimov, N., ... Holmes,
 1753 R. M. (2013). Dissolved organic carbon loss from Yedoma permafrost amplified by ice
 1754 wedge thaw. *ENVIRONMENTAL RESEARCH LETTERS*, 8(3).
 1755 https://doi.org/10.1088/1748-9326/8/3/035023
- 1756 Vonk, J. E., Mann, P. J., Davydov, S., Davydova, A., Spencer, R. G. M., Schade, J., ... Holmes,
 1757 R. M. (2013). High biolability of ancient permafrost carbon upon thaw. *GEOPHYSICAL*1758 *RESEARCH LETTERS*, 40(11), 2689–2693. https://doi.org/10.1002/grl.50348
- Waldrop, M. P., Harden, J. W., Turetsky, M. R., Petersen, D. G., McGuire, A. D., Briones, M. J.
 I., ... Pruett, L. E. (2012). Bacterial and enchytraeid abundance accelerate soil carbon
 turnover along a lowland vegetation gradient in interior Alaska. *Soil Biology and Biochemistry*, *50*, 188–198. https://doi.org/https://doi.org/10.1016/j.soilbio.2012.02.032
- Waldrop, M. P., & Harden, J. W. (2008). Interactive effects of wildfire and permafrost on
 microbial communities and soil processes in an Alaskan black spruce forest. *GLOBAL CHANGE BIOLOGY*, *14*(11), 2591–2602. https://doi.org/10.1111/j.13652486.2008.01661.x

- Ward, C. P., & Cory, R. M. (2015). Chemical composition of dissolved organic matter draining
 permafrost soils. *Geochimica et Cosmochimica Acta*, 167, 63–79.
 https://doi.org/https://doi.org/10.1016/j.gca.2015.07.001
- Ward, C. P., Nalven, S. G., Crump, B. C., Kling, G. W., & Cory, R. M. (2017). Photochemical alteration of organic carbon draining permafrost soils shifts microbial metabolic pathways and stimulates respiration. *NATURE COMMUNICATIONS*, 8.
 https://doi.org/10.1038/e41467.017.00750.2
- 1773 https://doi.org/10.1038/s41467-017-00759-2
- Whittinghill, K. A., Finlay, J. C., & Hobbie, S. E. (2014). Bioavailability of dissolved organic
 carbon across a hillslope chronosequence in the Kuparuk River region, Alaska. *Soil Biology and Biochemistry*, 79, 25–33. https://doi.org/https://doi.org/10.1016/j.soilbio.2014.08.020
- Wickland, K. P., Neff, J. C., & Aiken, G. R. (2007). Dissolved organic carbon in Alaskan boreal
 forest: Sources, chemical characteristics, and biodegradability. *ECOSYSTEMS*, *10*(8),
 1323–1340. https://doi.org/10.1007/s10021-007-9101-4
- Wickland, K. P., Waldrop, M. P., Aiken, G. R., Koch, J. C., Jorgenson, Mt., & Striegl, R. G.
 (2018). Dissolved organic carbon and nitrogen release from boreal Holocene permafrost and seasonally frozen soils of Alaska. *ENVIRONMENTAL RESEARCH LETTERS*, *13*(6).
 https://doi.org/10.1088/1748-9326/aac4ad
- Yun, J., Jung, J. Y., Kwon, M. J., Seo, J., Nam, S., Lee, Y. K., & Kang, H. (2022). Temporal
 Variations Rather than Long-Term Warming Control Extracellular Enzyme Activities and
 Microbial Community Structures in the High Arctic Soil. *MICROBIAL ECOLOGY*, 84(1),
 168–181. https://doi.org/10.1007/s00248-021-01859-9
- Zolkos, S., & Tank, S. E. (2019). Permafrost geochemistry and retrogressive thaw slump
 morphology (Peel Plateau, Canada), v. 1.0 (2017-2017). https://doi.org/10.5885/45573XD28DD57D553F14BF0

1791