Response to review comments of manuscript " Distinguishing between old and modern permafrost sources with compound-specific δ^2H analysis"
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We thank the three reviewers for their comments, and provide answers to these comments and suggestions below.

Black: review comment
Blue, italic: our response and edits in manuscript

Reviewer Trevor Porter
General comments:
Title should indicate the study region (e.g., Laptev Sea catchment, NE Siberia).
We have added "in the Northeast Siberian land-shelf system".

Abstract is one of the longest I've read in recent years. It includes a lot of useful information, but could (and probably should) be more concisely written. I leave this to authors and editor to decide.
We have shortened the abstract, as indeed it was rather long.

There is now a large body of literature on the isotopic composition of relict ice from ICD's in this very same region (see Opel et al. 2017 Climate of the Past; and references therein). These studies find that precipitation isotope composition recorded in these ICD's was highly variable during Pleistocene cold stages; for example, texture and pore ice 2H values range from roughly -250‰ to -160‰ between ca. 50-30 cal ka BP, while ice wedge values (winter precip) during the same interval range from roughly -260 to -230‰. If the fossil plants were using the same water that is preserved in the pore ice, then there may be a significant amount of variance not yet captured in the n-alkyl dataset from the (n = 9) ICDs sampled in this study. The spatial distributions of distinct ICD units in this region are not equal (see Opel et al., 2017) and, thus, have different potentials for erosion and contribution to the blend of n-alkyls deposited on the shelf. I would like the authors to acknowledge this potentially major source of uncertainty. I would also ask the authors disclose any information they have on the age of the sampled ICDs and, if possible, cross-reference to the regional stratigraphy scheme outlined in Opel et al. (2017).
Thank you for highlighting this point. We had not seen the article by Opel et al. While it is still not a peer-reviewed published paper (status listed as "in discussion"), it clearly provides additional insight to ICD from this region. We have now cited this paper, as well as a few others in the Introduction (and elsewhere) of our revised paper. We agree that the point brought up can be a source of uncertainty, and have acknowledged this fact in section 4.2 quite extensively.
We have also added the 14C information we have available on a few of the samples analyzed (Table 1). The new text reads as follows (line 492-510):
"Finally, we realize that the amount of soil and ICD samples analyzed in this study is limited, and want to point out that the results may change when more data are analyzed in the near future. Additionally, studies have shown that the d^2H signature of ice within ICD permafrost deposits can range from roughly -150‰ to -260‰ depending on the type of ice (wedge ice vs. texture ice) as well as the period of formation (different Pleistocene cold stages) (Opel et al., 2017 and references therein). The source of water (i.e. type of ice) and age of the deposit will therefore influence the n-alkane or n-alkanoic acid d^2H signal. However, regardless of the natural variability associated with the processes mentioned above, both ICD and texture-ice isotopic compositions appear to reflect long-term climate changes (Opel et al., 2017; Schwamborn et al., 2006; Dereviagin et al., 2013; Porter et al., 2016) which, likely, were also captured in the n-alkane or n-alkanoic acid d2H signal. Unfortunately, we do not have ^{14}C-ages available for all ICD samples, so cross-referencing to published stratigraphies in the region is not possible. Coastal sediments, however, will represent a

mixture of material released from different depths, outcrops, and stratigraphies within the catchment or coast. For source-apportionment applications, we reason that a growing body of leaf wax d^2H end-member data from the ICD region can overcome the variability issues highlighted above."

This paper would benefit from another figure that provides photographic examples of the ICDs and topsoil sections.
We do not have high-quality photographic material available from all the sites, unfortunately, but have added a figure as supplementary information with some examples.

Specific comments:
L44, The n-alkane sum and interquartile range given (210±350 ug/gOC) implies negative concentrations are possible, and is not consistent with Figure 2a. This also occurs on L299.
Since we shortened the abstract, we have removed the n-alkane concentrations from the abstract (L44) but have changed the notation in (the previous) L299 as this was perhaps confusing. The interquartile range was 350 (Q1-Q3), so we have chosen to now report this as 210^{494}_{148} to make it more clear that IQR1 is 148 and IQR3 is 494. (lines 344-349 and lines 364-369). We have also adjusted Table 4.

L149, instead of citing the IAEA website, better to cite a peer-reviewed article that supports your statement. Dansgaard (1964, Tellus) is appropriate.
OK, good suggestion, we have done this (line 160).

L158, it might be worth stating the underlying assumption, that colder air temperatures during the Pleistocene generally correlate with 2H-depleted precipitation; therefore, long-chain n-alky 2H during Pleistocene cold-stages should also be depleted compared to present. Also note that 'colder' and 'drier' could have opposing effects. All other factors equal (e.g., biochemical fractionation), a drier atmosphere during Pleistocene cold-stages could result in a larger leaf water enrichment and 2H n-alkyls (if RH is lower, despite lower air temps), which would lessen the overall offset between modern and Pleistocene n-alkyl 2H.
Thanks, this is a good suggestion. We have edited this sentence to now read "Despite the plant fractionation associated with kinetic and plant physiology (Sachse et al., 2012), we hypothesize that δ^2H signatures of leaf wax n-alkanoic acids and n-alkanes are more depleted in OC from permafrost deposits formed during the colder Pleistocene (generally correlating with ^2H-depleted precipitation), compared to more enriched values in OC from active layer or surface permafrost formed during the warmer Holocene. " (lines 178-183)

L199-201, if species information is available for the grasses and birch, please indicate.
Both samples were grass samples, but one of them was collected in the tundra, and one of them further south in a birch forest. We have clarified this (line 227-229). Unfortunately, we do not have species information available.

L308, the sphagnum index could also include C23 (see Bush and McInerney, 2013, GCA). For modern sphagnum samples I've collected in NW Canada (>65N), C23 is usually abundant (unpublished data). This suggestion isn't critical, but might be a more accurate metric for sphagnum vs. woody plants.
We are aware that either C23 or C25 can be abundant, but meant to illustrate the general differences (very small) between the two sample types. But, for comparison we have now replaced the C25/(C25+C29) ratio with C23/(C23+C29) in Figure 2 and added the C23 ratio to Table 4. Also, we have added a sentence to the text (line 357-360). Here, the average values for topsoil vs. ICD samples are further apart, yet still not statistically significant.

L345, please delete 'it seems'. If there is uncertainty, this can be described in a more quantitative way.
We have deleted it (line 407).

L519-521, unclear if you are talking about potential overprinting of the fossil 2H in situ (e.g., with water in the frozen ICD), or following transport and deposition on the shelf. Please clarify. Also, give a citation that supports the statement that overprinting is enhanced in low pH environments.
We have clarified this by stating that the environmental water can be coming from various sources (e.g. in situ or during transport after thaw). Also, we have removed the statement on low pH environments as we could not support this with a proper reference, and it is less relevant to our study (lines 603-607).

Anonymous reviewer #1
If a positive decision is made I have no other detailed comments than the references need to be looked over. Some of them even lack publication year.
We have carefully read through the references list and edited/corrected where needed.

Anonymous reviewer #2
General comments:
I think the abstract is too long, and that it goes into too much specific detail. I think that it could and should be made more succinct.
We agree. This point was also brought up by Reviewer Trevor Porter. We have shortened the abstract.

The phrase "molecular-bulk upscaling challenge" is used without enough introduction/ definition. I understand what you mean by it, but I think that it would be better to explain what this is exactly in a bit more detail.
We have added a bit more specific description the first time we use this definition, in the introduction: " This $\delta^{13}C$-$\Delta^{14}C$ dual-carbon isotope approach carries the strong advantage that it operates on the bulk OC level, thereby circumventing the "molecular-bulk upscaling challenge". This challenge relates to issues associated with upscaling from the molecular isotope level to the bulk level. These issues relate to the relative concentration (n-alkanes and n-alkanoic acids represent only a fraction of the total OC) but also to processes such as selective degradation, differences in physical association, or dispersion differences. " (lines 132-138).

A general comment about the structure of the discussion is the separation of the 13C-14C data from the bulk geochemistry. Why are these measurements not included in this grouping? If you measure 13C or 14C on a bulk sample, isn't that "bulk geochemistry"?
You might be able to circumvent this issue just by renaming the bulk section to "bulk elemental geochemistry" or something like that.
We do not really follow the reviewer here. In the first section of the discussion we talk about %C, C/N values, as well as d13C on bulk samples. As such, we named this section " ... bulk geochemistry ... as it includes both elemental and isotopic measurements on bulk samples. The same is the case for the first section of the Results.

One thing that I think is also missing from the discussion is some mention of the possibility that the terrestrial sampling density may have missed some of the possible heterogeneity in permafrost chemistry. I realize that it's not easy to sample in this part of the world, but is there any reason to think that the results might look different if you had soil samples from 50 more sites? Why or why not? This would apply to the 2H data, as well as the other data.

This is a valid point. We have added the following text to the manuscript, at the end of section 4.2: "Finally, we realize that the amount of soil and ICD samples analyzed in this study is limited, and want to point out that the results may change when more data are analyzed in the near future." (line 492-494)
Regarding the second point/suggestion of this review comment, we think it would be too speculative to give more detail regarding the possible differences in results if more data were to be obtained.

Specific comments:
Line 78 – change "into" to "in"
Changed.

Line 99 – Personally, I'm not a fan of non-standard acronyms like this (ICD in this case). They require an elevated level of buy in from the reader, which I think takes away from the accessibility of the manuscript. That's just my opinion, there's plenty of precedent for this kind of thing of course.
We realize there are different opinions with respect to acronym usage, but prefer to continue using it as this shorter version is commonly used and it also improves readability.

Line 137 – At the introduction of the 2H discussion, it might help to frame the study better if you begin by saying that you propose the new tool, as well as evaluate the performance using a suite of other geochemical data including the aforementioned 13C-radiocarbon method.
This is a good suggestion. We have now in the revised ms better introduced our tools in the introduction: " We will evaluate the performance of this complementary tool using additional geochemical data as well as the bulk $\delta^{13}C$-$\Delta^{14}C$ mixing approach." (lines 148-150).

Line 142-143 - These are nice papers, but they aren't really the best references to support the assertion that "the isotopic value of local precipitation is a function of local climate"
Yes, we agree. We have added: Craig H. 1961. Isotopic variations in meteoric waters. Science 133: 1702–1703. (line 154)

Line 149 - If you mean to give the maximum range you could point out that precip in east africa can be upwards of +50 per mil, while the SLAP2 (Standard Light Antarctic Precipitation 2) standard is -427.5 per mil.
We have edited this to now present the maximum range, using the values/locations that this reviewer provides (lines 160-163).

Line 162-168 – The end of the introduction falls a little flat in my opinion. At the moment you say what you do in your study, followed by a general statement about why it's important to study these types of questions. What's missing to me is a statement that directly comments on how what you do with this study will help with these important questions. As it is currently written it doesn't setup the next section so effectively.
We see the point, and have therefore now added one more sentence that specifically mentions the use of our proposed tool, at the very end of the introduction: "Our proposed tool may be used to trace these temporal and spatial differences in OC release from permafrost thaw, as well as the extent of burial of OC in sedimentary reservoirs."(lines 191-193).

Line 195 - I think it is better to replace your internal lab sample codes with something more straightforward when reporting the results (things like "CH DY-3A" are meaningless to the reader and hard to remember). Include them in a data file or something if you want to be able to cross-reference with Vonk et al., 2013, but for presentation purposes I would simplify.
We have renamed the samples with TS-1, TS-2, ICD-1, ICD-2 etc, and include the original sampling ID in Table 1.

Line 232 – Remember to define acronyms at first use.
This acronym was defined in the previous paragraph.

Line 243 – Check super/subscripting for H3+.
We have edited this into H_3^+ (line 276).

Line 244 – Give units for H3+.
We have added units (‰ per V) and have also added a reference for the use of H_3^+ (Sessions et al., 2011) (line 277).

Line 248 - The "methylation effect" language is odd to me, since it makes it sound like what was quantified was the difference in 2H values between the derivatized and non derivatized standard, rather than the 2H value of the hydrogen in the added methyl group. Since the magnitude of the "methylation effect" will be different depending on what the 2H value of the covalently bonded hydrogen in the methylated fatty acid is in addition to the chain length, you want to do the correction by mass balance. Probably that is what you did, but the language doesn't make it sound that way.
We agree, we have changed this into "methylation correction" (lines 281-282).

Line 251 – This call to table 5 is out of order since you haven't called tables 2-4 yet.
This sentence is a general remark on how we report the d2H values, so we decided to not call table 5 at this particular place.

Line 259 – Not sure what you mean exactly by "with mean and standard deviations obtained from the literature values".
This was meant to refer to the end-member values for the d13C-D14C source apportionment, but is perhaps confusing. We have now specified this to "with mean and standard deviations obtained from our analysis (d2H on TS and ICD samples) and from literature (13C and 14C on end-members)" (lines 298-299).

Line 296 – I might add a few words to the start of the sentence that begins on this line to make it clear that you are discussing distributions within individual samples, and that you are still talking about topsoil samples only.
We have done this by adding " for Topsoil-PF samples" to this sentence (line 343).

Line 318 – This call to table 3 is out of order.
Indeed, this should be table 2. In the previous sentence, however, we have changed a call to table 2 into table 3. As all tables have already been called before, we did not change the order of the tables.

Line 354 – spell check.
Thanks, we have corrected this.

Line 417 – change "proxies" to "proxy"
We have changed this.

Line 452 – This is the first mention of results from the shelf-slope samples. In the methods you point out a reference for more information on the sampling procedures, but what about the laboratory analyses and results? This should be included in the earlier sections.
Yes, good point. We have added a brief paragraph at the end of section 2.2 (lines 286-290).

Line 454 - I like how you use the individual n-alkanes rather than arbitrarily averaging them together.
Thanks.

Lines 481 – 500 – Somewhere in this section, or somewhere else in the manuscript if it fits better, it would be good to discuss how variability within an end member might impact the results. This is important for both the 2H and the 13C-radiocarbon approaches, but it seems like it would be especially important for the radiocarbon. In addition to the acknowledged aging along the transect won't there be different ages within a topsoil permafrost? How might this impact the results if melting/erosion occurs at different depths/ages within a site?
We agree that variability within the end-members plays an important role and should be taken into account. For the 13C and 14C approaches, the amount of end-member data available is fairly good (and growing) with 30-40 data points for 13C and >300 data points for 14C. This is described in the second paragraph of section 4.3. Our Markov Chain Monte Carlo mass-balance model actually accounts for the end-member variability (described and referenced in section 3.2). When thawing and erosion occurs at different depths or ages, at various locations throughout a watershed or coastline, the signal will be averaged when measured in coastal sediments. We have now acknowledged this important point (lines 505-507) (reviewer Trevor Porter also posted a related comment). Regarding the amount of d2H end-member data available: we are aware that our sample set only exists of n=9 data points for each source. Variability in the mean end-member values may therefore change (or, perhaps, decrease) when more data become available. We have now also briefly mentioned this point at the end of section 4.2 (lines 492-494).

Line 568 – As with the end of the introduction, I think that the end of the conclusion could go a little further to bring this study back together with the big picture goals. Remind us how "increasing our understanding of the fate of thawing permafrost in the coastal environment" will help us and why we should care about it.
We have added a bit more "big picture" text to place the results of our study into context with the general goals outlined in the introduction. The final paragraph of the conclusions now reads:
"This study shows that δ^2H of leaf wax molecules has the potential to be used in quantitative source-apportionment studies of thawing permafrost in coastal or marine settings. It can serve as an alternative or complementary approach to the commonly applied bulk $\delta^{13}C$-$\Delta^{14}C$ method. We recommend continuing data collection and optimization of end-member definition and calibration. Refining the molecular d^2H proxy presented here will be beneficial in pinpointing the location and extent of OC release from thawing permafrost in the coastal or fluvial environment. With enhanced Arctic warming and associated intensification of permafrost thaw, constraining the amount and fate of permafrost OC release will help to assess the magnitude of the permafrost carbon feedback to climate warming." (lines 658-663).

Figure 2 – I would add the color legend to this figure that you already use on the other figures. I would also list n values in the caption or on the figure.
Yes, this has been changed.

Figure 5 - I would list the modern/ICD labels as headers rather than within the data.
OK, we have done this.

Distinguishing between old and modern permafrost sources in the Northeast Siberian land-shelf system with compound-specific δ^2H analysis

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Keywords: deuterium isotopes, yedoma, ice complex deposit, n-alkanoic acids, n-alkanes, organic matter, stable carbon isotopes, radiocarbon, Siberian Arctic, sediments, permafrost thaw

Abstract

Pleistocene ice complex permafrost deposits contain roughly a quarter of the organic carbon (OC) stored in permafrost terrain. When permafrost thaws, its OC is remobilized into the (aquatic) environment where it is available for degradation, transport or burial. Aquatic or coastal environments contain sedimentary reservoirs that can serve as archives of past climatic change. As permafrost thaw is increasing throughout the Arctic, these reservoirs are important locations to assess the fate of remobilized permafrost OC.

We here present compound-specific deuterium (δ^2H) analysis on leaf waxes as a tool to distinguish between OC released from thawing Pleistocene permafrost (Ice Complex Deposits; ICD) and from thawing Holocene permafrost (from near-surface soils). Bulk geochemistry (%OC, $\delta^{13}C$, %total nitrogen; TN) was analyzed as well as the concentrations and δ^2H signatures of long-chain *n*-alkanes (C_{21} to C_{33}) and mid/long-chain *n*-alkanoic acids (C_{16} to C_{30}) extracted from both ICD-PF samples (n=9) and modern vegetation/O-horizon (Topsoil-PF) samples (n=9) from across the northeast Siberian Arctic.

Results show that these Topsoil-PF samples have higher %OC, higher OC/TN values, and more depleted $\delta^{13}C$-OC values than ICD-PF samples, suggesting that these former samples trace a fresher soil and/or vegetation source. Whereas the two investigated sources differ on the bulk geochemical level, they are, however, virtually indistinguishable when using leaf wax concentrations and ratios.

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However, on the molecular-isotope level, leaf wax biomarker δ^2H values are statistically different between Topsoil-PF and ICD-PF. For example, the mean δ^2H value of C_{29} *n*-alkane was -246±13‰ (mean±stdev) for Topsoil-PF and -280±12‰ for ICD-PF. With a dynamic isotopic range (difference between two sources) of 34 to 50‰, the isotopic fingerprints of individual, abundant, biomarker molecules from leaf waxes can thus serve as end-members to distinguish between these two sources. We tested this molecular δ^2H tracer along with another source-distinguishing approach, dual-carbon ($\delta^{13}C$-$\Delta^{14}C$) isotope composition of bulk OC, for a surface sediment transect in the Laptev Sea. Results show that general offshore patterns along the shelf-slope transect are similar, but the source apportionment between the approaches vary, which may highlight the advantages of either. This study indicates that the application of δ^2H leaf wax values has potential to serve as a complementary quantitative measure of the source and differential fate of OC thawed out from different permafrost compartments.

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Deleted: , whereas the C_{31} *n*-alkane was -247±23‰ for Topsoil-PF and -297±15‰ for ICD-PF. The C_{28} *n*-alkanoic acid δ^2H value was -220±15‰ for Topsoil-PF and -267±16‰ for ICD-PF.

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1 Introduction

Climate warming is causing permafrost soils to thaw, exposing its organic matter (OM) to decomposition (e.g., Schuur et al., 2015; Zimov et al., 1993; Semiletov et al., 2012). Thaw will increase the hydrological connectivity of landscapes and will cause release of OM into the aquatic environment (Walvoord et al., 2012; Vonk et al., 2015; Anderson et al., 2011). Here, the OM can continue to decompose, generating greenhouse gases (e.g., Semiletov et al., 1996a,b; Anderson et al., 2009; Shakhova et al., 2015), or be destined for burial in inland and coastal sediments. These sedimentary archives serve as long- and short-term reservoirs that attenuate greenhouse gas emissions from thawing permafrost (Vonk and Gustafsson, 2013; Semiletov et al., 2011).

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The release of OM from thawing permafrost into aquatic sediments varies over time and space. A recent study showed that at the end of the last glacial, the surface active layer of terrestrial permafrost released about 4.5 Tg organic carbon (OC) per year from just the Lena watershed onto the nearby shelf, whereas current annual OC release is estimated to be only about a tenth of this (Tesi et al., 2016). In addition to active layer material, OM from deeper and older permafrost sources can also thaw and be released into the environment (Shakhova et al., 2007, 2014). This process currently dominates the delivery of terrestrial material onto the East Siberian Arctic shelf (Vonk et al., 2012; Semiletov et al., 1999) and is expected to increase due to accelerating coastal erosion rates (Günther et al., 2013).

Different permafrost OC stocks exhibit variable vulnerabilities to thaw remobilization (Schuur et al., 2015). In addition to a subsea permafrost OC stock, soils and sediments of the terrestrial northern permafrost zone store about 1300±200 Pg OC, with separate upscaling approaches applied for soil stocks (0-3m depth), deltaic sediments (full depth) and Yedoma sediments (full depth) (Hugelius et al., 2014). Yedoma sediments, a.k.a. Ice Complex Deposits (ICD) are polygenetic, ice-rich Pleistocene-aged deposits that are present in the unglaciated parts of Siberia and Alaska (Schirrmeister et al., 2011). These deposits contain roughly a quarter of the OC stored in permafrost terrain, but estimates vary from ca. 200-400 Pg C (Strauss et al., 2013; Schuur et al., 2015). The presence of massive ice wedges in ICD causes landscapes to collapse upon thaw, exposing deeper stocks of OC. This type of relatively abrupt thaw is increasing in many parts of the arctic landscape (Schuur et al., 2015). At the same time, deepening of the active layer causes gradual thaw that occurs across entire landscapes (Shiklomanov et al., 2013).

With a tool to detect and monitor different types of permafrost OM in coastal environments, one could assess (historical and spatial) variability in permafrost source input, degradation and thaw, as well as the relative degradation of different permafrost types. For example, the relative release of OC from ICD versus topsoil permafrost has earlier been distinguished and quantified through the use of dual-carbon isotopes ($\delta^{13}C$ and $\Delta^{14}C$) on bulk OC in the shelf environment of the Laptev and East Siberian Sea. It was shown that topsoil permafrost OC dominates in suspended particulate matter (Karlsson et al., 2011; 2016; Vonk et al., 2012) and ICD

permafrost OC dominates in the surface sediments (Vonk et al., 2012; Semiletov et al., 2011; 2012). Vonk et al. (2014) further showed that topsoil OC is actively degraded during horizontal transport whereas ICD permafrost OC rapidly settles. Winterfeld et al. (2015) showed, using dual-carbon isotopes on riverine material, that suspended particulate OC in the Lena Delta mostly consists of Holocene material instead of material from ICD permafrost.

This $\delta^{13}C$-$\Delta^{14}C$ dual-carbon isotope approach carries the strong advantage that it operates on the bulk OC level, thereby circumventing the "molecular-bulk upscaling challenge". This challenge relates to issues associated with upscaling from the molecular isotope level to the bulk level. These issues relate to the relative concentration (*n*-alkanes and *n*-alkanoic acids represent only a fraction of the total OC) but also to processes such as selective degradation, differences in physical association, or dispersion differences. However, the $\delta^{13}C$-$\Delta^{14}C$ approach also has drawbacks, such as a weak distinction between the $\delta^{13}C$ end-member values of Topsoil-PF versus ICD-PF. Also, the marine $\delta^{13}C$ end member values in coastal Arctic shelf waters are uncertain and may be more depleted than at mid-latitudes due to uptake of relatively depleted dissolved CO_2 values caused by cold polar water (Meyers, 1997; Tesi et al. *this special issue*) or degradation of terrestrial matter (Anderson et al., 2009; 2011; Semiletov et al., 2013; 2016), generating a potential overlap between marine and topsoil $\delta^{13}C$ end-members.

Here we propose a complementary tool to trace permafrost OC release into the coastal environment based on molecular $\delta^{2}H$ analysis on leaf waxes. We will evaluate the performance of this tool using additional geochemical data as well as a bulk $\delta^{13}C$-$\Delta^{14}C$ mixing approach. Isotopes in water molecules ($\delta^{2}H$ or $\delta^{18}O$) in glacial ice cores as well as in massive ground ice in the northern hemisphere have been used for reconstructing palaeotemperatures (e.g., Kotler and Burn, 2000; Johnson et al., 2001; Opel et al., 2011; Meyer et al., 2015; Wetterich et al., 2016) as the isotopic value of local precipitation is a function of local climate (Craig, 1961; Sachse et al., 2004; Smith and Freeman, 2006). Higher plants use water as their primary source of hydrogen during photosynthesis (Sternberg, 1988). The $\delta^{2}H$ isotope values of leaf wax *n*-alkanoic acids or *n*-alkanes are therefore reflecting the $\delta^{2}H$ isotopic value of local precipitation (e.g., Sachse et al., 2004; Sessions et al., 1999), after correction for the net fractionation during biosynthesis, and evapotranspiration (Leaney et al., 1985). Global precipitation values can vary immensely (Dansgaard, 1964) with values up to +50‰ in Eastern Africa but approaching -200‰ near the North Pole (www.iaea.org) or even below -400‰ in Antarctica (i.e. SLAP2 standard, Standard Light Antarctic Precipitation, is -427.5‰). Additionally, the fractionation between source water and plant wax molecules varies both in time and space, and can be up to -170‰ (Smith and Freeman, 2006; Sachse et al., 2004; Polissar and Freeman, 2010) but appears relatively small at higher latitudes (between -59 and -96‰; Shanahan et al., 2013; Wilkie et al., 2013; Porter et al., 2016). Differences in $\delta^{2}H$ signatures of leaf wax molecules from terrestrial regions with different (past) climates could therefore potentially be applied to derive the relative proportion of different types of thawing

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permafrost in nearby coastal settings. Despite the plant fractionation associated with kinetics and plant physiology (Sachse et al., 2012), we hypothesize that δ^2H signatures of leaf wax *n*-alkanoic acids and *n*-alkanes are more depleted in OC from permafrost deposits formed during the colder Pleistocene (generally correlating with ^2H-depleted precipitation), compared to more enriched values in OC from active layer or surface permafrost formed during the warmer Holocene.

This study investigates a source-specific δ^2H signature for both ICD permafrost and recent, surface soil permafrost in Northeast Siberia. Furthermore, we explore the possibilities of using these isotopic end-member values in regional source-apportionment calculations that aim to quantify the relative contribution of different sources of permafrost OC. As permafrost thaw progresses, particularly in ice-rich permafrost such as ICD, it is increasingly important to trace the fate of remobilized and decomposing OC in the Arctic environment. Our proposed tool may be used to trace these temporal and spatial differences in OC release from permafrost thaw, as well as the extent of burial of OC in sedimentary reservoirs.

2 Methods

2.1 Sampling

A total of 18 samples were collected throughout the Siberian Arctic. Recent surface soils (n=7) and vegetation (n=2) samples were analyzed and (from here on) referred to as the "topsoil" permafrost (Topsoil-PF) sample set, whereas ICD-PF samples were obtained from ICD soil profiles (n=7) and suspended particulates from ICD formations (n=2) (Fig. 1 and Table 1). Eight offshore sediments along a shelf-slope-continental rise transect in the Laptev Sea were collected in 2014, further marine sampling details can be found in Bröder et al. (2016b).

The Topsoil-PF samples represent O and A soil genetic horizons in sites with active soil formation. The sites where chosen to represent typical soil and vegetation types in the investigated permafrost landscapes, including both taiga and tundra sites. Samples were collected by depth or soil horizon increments from open soil pits using fixed volume sampling procedures.

The ICD-PF samples were collected from vertical exposures that were excavated to expose intact permafrost. Fixed-volume samples were collected by coring horizontally into the frozen sediments to extract ICD-PF samples from consecutive depths.

For more details about sampling sites, including location, vegetation and soil types see table 1 (terminology following the U.S.D.A. Soil Taxonomy; Soil Survey Staff, 2014). Sampling was done in late summer near the time of maximum annual active layer depth, in July 2010 (ICD-8 and ICD-9; Vonk et al. (2013)) and August 2011 (Palmtag et al., 2015) for the Kolyma River region, in August 2012 for the lower Lena River and Indigirka River (Siewert et al., 2015; Weiss et al., 2015) and in August 2013

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for the upper Lena River (Siewert et al., 2016). For more detailed descriptions of sample collection we refer to these references. The vegetation samples TS-8G (grass) and TS-9G (grass) were obtained from the tundra near Medvezhka River and a birch forest near Y4 stream, respectively, in July 2012.

Samples JCD-8 and JCD-9 were obtained in July 2010 at the Duvannyi Yar ICD exposure along the Kolyma River (Vonk et al., 2013). The particulate sediment samples were taken from thaw streams that were freshly formed from thawing ICD (transport time from thaw to sampling estimated to be less than 1h).

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2.2 Analytical methods

Freeze-dried samples were extracted using an ASE 200 accelerated solvent extractor (Dionex Corporation, USA) using DCM/MeOH (9:1 v/v) at 80°C ($5x10^6$ Pa) (Wiesenberg et al., 2004). After the extraction, solvent-rinsed activated copper and anhydrous sodium sulfate were added to the extracts to remove sulfur and excess water, respectively. After 24 h, extracts were filtered on pre-combusted glass wool and concentrated with the rotary evaporator. Extracts were transferred into glass tubes, evaporated to complete dryness and re-dissolved in 500 µl of DCM. Lipid fractionation was performed via column chromatography using amino-propyl Bond Elut (500 mg/3 ml) to retain the acid fraction and Al_2O_3 to separate the hydrocarbon and polar fractions (Vonk et al., 2010).

Prior to the analyses, saturated *n*-alkanes (hydrocarbon fraction) were further purified using 10% $AgNO_3$ coated silica gel to retain the unsaturated fraction. The acid fraction was methylated using a mixture of HCl, MilliQ water and methanol at 80°C overnight to obtain the fatty acid methyl ester (FAME) fraction. Methylated acids were extracted with hexane and further purified using 10% $AgNO_3$ coated silica gel. The hydrocarbon and FAME fractions were quantified via gas chromatography mass spectrometry (GC–MS) in full scan mode (50-650 m/z) using the response factors of commercially available standards (Sigma-Aldrich). The GC was equipped with a 30 m×250 µm DB5-ms (0.25 µm thick film) capillary GC column. Initial GC oven temperature was set at 60°C followed by a 10°C min^{-1} ramp until a final temperature of 310°C (hold time 10 min).

The hydrogen-isotopic composition of hydrocarbon and FAME fractions was measured with continuous-flow GC - isotope ratio - MS. Purified extracts were concentrated and injected (1-2 µl) into a Thermo Trace Ultra GC equipped with a 30m×250 µm HP5 (0.25 µm thick film) capillary GC column. Oven conditions were similar to the setting used for the quantification. The conversion of organic biomarkers to elemental hydrogen was accomplished by high-temperature conversion (HTC) at 1420°C (Thermo GC Isolink). After the HTC, H_2 was introduced into the isotope ratio MS (Thermo Scientific™ Delta V™IRMS) for compound-specific determination of δ^2H values via a Thermo Conflo IV. Following a linearity test, we only used peaks with amplitude (mass 2) between 1500 and 8000 mV for the evaluation. The δ^2H values were calibrated against saturated HMW *n*-alkanes using the reference

substance mix A4 (Biogeochemical Laboratories, Indiana University). The H_3^+ factor (Sessions et al., 2001) was determined every day and stayed constant (<3 ‰/V) throughout our analyses period. Each purified extract was injected three times. FAMEs were further corrected to account for the methylation agent by comparing the hydrogen abundance of lauric acid (C_{12}-FA; i.e. 12 carbon atoms) as acid and corresponding methyl ester. The average methylation correction for lauric acid was 23.97±3.9‰ (n=4). This correction was, normalized to chain length (i.e. increasing chain lengths result in lower corrections), applied to all the FAMEs. δ^2H values of *n*-alkanes and FAMEs are reported as mean, standard deviation and weighted average.

Details of the analytical methods for extraction, work-up, and purification of the eight offshore sediment samples for biomarker analysis that are included in our source-apportionment comparison (section 4.3) can be found in Bröder et al. (2016b). The δ^2H analysis on the shelf sediments was performed in parallel with the ICD-PF and Topsoil-PF samples, according to the method described above.

2.3 Source apportionment

The compound-specific δ^2H signatures in this study were used to differentiate between the two major sources (end-members), Topsoil-PF and ICD-PF, using an isotopic mass-balance model. We used a Markov chain Monte Carlo (MCMC) approach to account for the end-member variability (Andersson et al., 2015; Bosch et al., 2015). The end-members were represented by normal distributions, with mean and standard deviations obtained from our analysis (δ^2H on TS and ICD samples) and from literature ($\delta^{13}C$ and $\Delta^{14}C$ on end-members)". For each Laptev Sea station, the isotope signatures from three different terrestrial molecular markers (long-chain *n*-alkanes C_{27}, C_{29} and C_{31}) were used jointly to improve source apportionment precision. The δ^2H signatures for the two end-members were based on our Topsoil-PF and ICD-PF samples.

The compound-specific δ^2H-based source apportionment was compared to $\Delta^{14}C/\delta^{13}C$-based analysis of bulk OC using analogous MCMC techniques (e.g., Vonk et al., 2012). The $\Delta^{14}C/\delta^{13}C$-approach allows estimation of the relative contribution of a third source, marine, which does not affect the presently investigated (terrestrial) compounds. Accounting for the marine component to OC allows direct comparison of the Holocene and Pleistocene contributions. All MCMC calculations were made using Matlab scripts (ver. 2014b) using 200,000 iterations, a burn-in phase (initial search period) of 10,000 and a data thinning of 10.

The spatial extent of ICD in the Lena River Basin was calculated by overlaying the extent of the drainage basin (from WRIBASIN: Watersheds of the World published by the World Resources Institute, www.wri.org/publication/watersheds-world) with the extent of the Yedoma Region (digitized from Romanovsky, 1993) in an equal area map projection. It was assumed that 30% of the Yedoma Region consists of intact ICD (following Strauss et al., 2013).

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3 Results

3.1 Bulk geochemistry

The investigated Topsoil-PF and ICD-PF samples are, on a bulk geochemical level, very different. Mean organic carbon contents (as %OC) and total nitrogen content (as %TN) are 25±12 and 1.1±0.67 for Topsoil-PF samples, and 1.6±0.31 and 0.17±0.058 for ICD-PF samples, respectively (Table 1). This gives TOC/TN ratios of 25±8.0 for Topsoil-PF samples and 10±2.6 for ICD-PF samples. Stable carbon isotopic values of Topsoil-PF and ICD-PF samples are -27.8±1.3‰ and -25.7±0.75‰, respectively (Table 1).

3.2 Molecular geochemical composition

Long-chain *n*-alkanes and *n*-alkanoic acids are abundant in epicuticular waxes and therefore indicative for a source of higher plants (Eglinton and Hamilton, 1967). Concentrations of individual long-chain *n*-alkanes in Topsoil-PF samples ranged from 1 to 340 µg/gOC (C_{21}-C_{33}; Table 2) with an average chain length of 28±1.6. The sum of high-molecular weight (HMW) *n*-alkanes (>C_{21}) for Topsoil-PF samples was 418^{612}_{280} µg/gOC (median with interquartile range) and the most abundant *n*-alkanes added up to 214^{494}_{148} µg/gOC (sum of C_{25}-C_{27}-C_{29}-C_{31}) (Table 4, Fig. 2a). For ICD-PF samples, the individual concentrations of long-chain *n*-alkanes were between 4 and 160 µg/gOC, and the average chain length 27±0.7 (Table 2). The sum of high-molecular weight *n*-alkanes, and most abundant *n*-alkanes were 698^{806}_{630} µg/gOC and 347^{405}_{323} µg/gOC, respectively (Table 4, Fig. 2a). The carbon preference index (CPI), a molecular ratio indicative for degradation status with values >5 typical for fresher terrestrial material and values approaching 1 typical for more degraded samples (Hedges and Prahl, 1993), showed values for Topsoil-PF samples of 7.3±3.6 (average±standard deviation) and ICD-PF samples of 3.6±0.8 (CPI C_{23}-C_{31}; Table 4, Fig. 2c). The $C_{25}/(C_{25}+C_{29})$ ratio, indicative for the input of peat moss (*Sphagnum sp.*) material (Vonk and Gustafsson, 2009; *Sphagnum* values 0.72, higher plants 0.07; Nott et al., 2000) was 0.33±0.22 (average±standard deviation) and 0.34±0.05 for Topsoil-PF and ICD-PF samples, respectively (Table 4). Another commonly used *Sphagnum* proxy (Bush and McInerney, 2013), $C_{23}/(C_{23}+C_{29})$, resulted in a sharper contrast between ICD-PF and Topsoil-PF samples (0.39±0.13 and 0.25±0.23, respectively; Fig. 2e and Table 4).

Long-chain *n*-alkanoic acids (C_{22} and above) were abundant in concentrations between 0.122 and 2670 µg/gOC for individual homologues in topsoils, with the sum of HMW *n*-alkanoic acids (>C_{22}) being 6397^{7454}_{3167} µg/gOC (median and IQR) and the most abundant *n*-alkanoic acids (sum of C_{22}-C_{24}-C_{26}-C_{28}) adding up to 4700^{6092}_{2670} µg/gOC (Table 3, 4 and Fig. 2b). ICD-PF samples contained individual long-chain *n*-alkanoic acids in 2.17 and 18700 µg/gOC (Table 2), a sum of HMW *n*-alkanoic acids of 8290^{11430}_{6290} µg/gOC, and the sum of most abundant, even *n*-alkanoic acids of 6630^{8790}_{5285}µg/gOC (Table 4). Topsoil-PF and ICD-PF samples had average chain lengths of 24.1±1.1 and 24.3±0.59, and CPI (C_{22}-C_{28}) values of 5.9±2.7 (average±standard deviation) and 5.0±1.6, respectively (Table 4). Shorter-chain *n*-alkanoic acids C_{16} and

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C_{18} are produced in basically all types of life in soils or aquatic environments, and are not specific for higher plants. Topsoil-PF contained C_{16} and C_{18} homologues in concentrations between 220 and 4600 µg/gOC, and ICD-PF samples between 200 and 10400 µg/gOC (Table 3).

Degradation of organic matter involves the loss of functional groups, e.g. the loss of carboxylic acids (Meyers and Ishiwatari, 1993). A high ratio of HMW *n*-alkanoic acids over HMW *n*-alkanes in a sample therefore implies a relatively fresh, less degraded, status (i.e. relatively more functional groups present). For Topsoil-PF samples, the HMW *n*-alkanoic acid/HMW *n*-alkane ratio varied between 5.6 and 25 with an average value of 13±7.6, whereas ICD-PF samples varied between 7.6 and 140 with an average value of 29±43 (Table 4, Fig. 2f).

3.3 Molecular isotopic composition

We measured δ^2H values in long-chain *n*-alkanes and *n*-alkanoic acids between -119 and -313‰ (Fig. 3, Table 5). Mean values for HMW *n*-alkanes (C_{25}-C_{27}-C_{29}-C_{31}) were between -201 and -247‰ for Topsoil-PF samples and between -221 and -297‰ for ICD-PF samples, with consistently lower δ^2H for longer chain lengths. For HMW *n*-alkanoic acids (C_{22}-C_{24}-C_{26}-C_{28}) mean δ^2H values were between -203 and -236‰ for Topsoil-PF samples and between -261 and -278‰ for ICD-PF samples (Table 5). The decrease in δ^2H values with increasing chain length is less distinct for *n*-alkanoic acids but one can observe a decrease of around 25-30‰ from C_{22} to C_{26} (Fig. 3). For ICD-PF samples, the isotopic depletion for the average of the three most abundant *n*-alkanes is comparable to the average for *n*-alkanoic acids, whereas in Topsoil-PF samples, the isotopic depletion for the three most abundant *n*-alkanes is a bit larger than for *n*-alkanoic acids (Fig. 4).

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4 Discussion

4.1 Using bulk geochemistry and molecular proxies

Bulk geochemical and isotopic analysis, as well as analysis of molecular proxies remained inconclusive in distinguishing between the two investigated sources in this study. Topsoil-PF samples have a higher organic content, higher TOC/TN values (representing fresh, higher plant material; Meyers, 1994) and more depleted $\delta^{13}C$ values (indicative for terrestrial C3 plants; Meyers, 1997) than ICD-PF samples, suggesting that these samples indeed trace a fresh soil and/or vegetation source (Table 1). The $\delta^{13}C$ values of a larger ICD-PF and Topsoil-PF dataset have earlier been summarized (Vonk et al., 2012 and references therein; Schirrmeister et al., 2011) giving values of -26.3±0.67‰ (n=374) and -28.2±2.0‰ (n=30), respectively. Our values (Table 1) are in a similar range. Despite the differences between these two sources in their bulk geochemistry, it is hard to use these parameters for source distinction as their variability is fairly high, and their behavior in the environment is not conservative, but e.g. affected by degradation processes. On a molecular geochemical level the two investigated sources are virtually indistinguishable as there is a considerable variation in molecular concentrations and proxy values (Fig.

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2). Only one of the tested parameters, the CPI C_{23}-C_{31} of *n*-alkanes, showed a statistically significantly different value for the two investigated sources.

4.2 Evaluation of molecular δ^2H values as a source end-member

To alleviate the difficulty to distinguish between Topsoil-PF and ICD-PF with just bulk and molecular geochemical characteristics, we explore the δ^2H values of leaf wax molecules (i.e. long chain *n*-alkanoic acids and *n*-alkanes) to differentiate between their relative source contributions. The overall mean δ^2H of the four most abundant *n*-alkanoic acids is -231±29‰ and -271±13‰ for Topsoil-PF and ICD-PF samples, respectively. These values compare well with available literature (Fig. 5). Pautler et al. (2014) measured δ^2H values on C_{29} *n*-alkanes in modern soils of the Yukon, Canada of -252±9.1‰ (n=4) and aged soil δ^2H values of -269±8.6‰ (n=13; 24-25 ^{14}C-ka ago) and -273±16.4‰ (n=9; for MIS 4, ~70 ^{14}C-ka ago). Yang et al. (2011) also reported C_{29} *n*-alkane δ^2H values for modern vegetation from Alaska and Arctic Canada with an average value of -252±43‰ (n=8). Zech et al. (2011) reported values of C_{29} *n*-alkanes collected from a permafrost exposure along the Tumara River in northeast Siberia, with an average value of -266±7.5‰ (n=23) for glacial paleosoils and -247±9.4‰ (n=17) for interglacial paleosoils. Our values for C_{29} *n*-alkanes for Topsoil-PF (-246±13‰; n=9) and ICD-PF (-280±12‰; n=9) are in a similar range (Fig. 5). For C_{28} *n*-alkanoic acids, Wilkie et al. (2013) measured -252±8.7‰ (n=6) for modern vegetation in northeast Siberia, whereas Porter et al. (2016) measured -269±2.7‰ (n=7) for ca. 31 cal ka BP old soils in the Yukon. Compared to these studies, our values for C_{28} *n*-alkanoic acids are somewhat more enriched for Topsoil-PF with -220±15‰ (n=7) but roughly in the same range for ICD-PF with -267±16‰ (n=9).

The mean isotopic difference between the most abundant *n*-alkanoic acids of the two investigated sources is around 40‰ (δ^2H values of -231±29‰ and -271±13‰ for Topsoil-PF and ICD-PF samples, respectively). Despite the relatively large standard deviations, the isotopic differences are statistically significant for each of the *n*-alkanoic acids individually (C_{22}, C_{24}, C_{26}, C_{28}; Fig. 3). The isotopic differences between the two sources for the mean value of the four most abundant *n*-alkanes is 35‰, with a mean value of -229±33‰ and -264±34‰ for Topsoil-PF and ICD-PF samples, respectively. Here, the individual *n*-alkane isotopic signatures are statistically significantly different for C_{27}, C_{29}, C_{31} (Fig. 3) in Topsoil-PF and ICD-PF samples. The selection and application of individual chain length δ^2H values as end-members, in contrast to mean chain length values, might be more appropriate for several reasons; (i) to reduce variability (δ^2H ranges for C_{29} and C_{31} *n*-alkanes and C_{22} and C_{24} *n*-alkanoic acids are relatively low; Fig. 3), (ii) to target the most abundant species (C_{29} and C_{31} *n*-alkanes are generally more abundant in soils and ICD-PF compared to shorter chain lengths; Table 2), and (iii) to make use of the largest dynamic range between source end-member values (C_{31} *n*-alkane δ^2H values of Topsoil-PF and ICD-PF differ by 50‰). Based on these arguments, the C_{28} *n*-alkanoic acid and the C_{29} or C_{31} *n*-alkanes are most appropriate to use for source-apportionment. The available previous studies (Fig. 5) have also selected these chain lengths (C_{28} *n*-alkanoic acid and C_{29} *n*-alkanes) for proxy development.

The use of molecular δ^2H values as tracers of terrestrial material in a marine or coastal setting has the advantage that it avoids uncertainty issues related to definition of the marine end-member. On the other hand, the inherent bulk-upscaling challenge of any molecular proxy, is a disadvantage of the δ^2H approach as it introduces unknowns related to the molecular-bulk upscaling effort (e.g. taking into account sorting and recalcitrance; discussed in depth in 4.3). We also want to emphasize that δ^2H leaf wax values in the two studied end-member sets (Topsoil-PF vs. ICD-PF) largely depend on the climate (warm vs. cold) and continentality (near the coast vs. further inland) during plant formation, and associated differences in fractionation mechanisms. Consequently, when δ^2H values in samples are used for source-apportionment, this may represent the fraction leaf wax produced in cold vs. warm conditions (as well as degree of continentality), and not necessarily the fraction Topsoil-PF vs. ICD-PF.

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Finally, we realize that the amount of soil and ICD samples analyzed in this study is limited, and want to point out that the results may change when more data are analyzed in the near future. Additionally, studies have shown that the δ^2H signature of ice within ICD permafrost deposits can range from roughly -150‰ to -260‰ depending on the type of ice (wedge ice vs. pore or texture ice) as well as the period of formation (different Pleistocene cold stages) (Opel et al., 2017 and references therein). The source of water (i.e. type of ice) and age of the deposit will therefore influence the *n*-alkane or *n*-alkanoic acid δ^2H signal. However, regardless of the natural variability associated with the processes mentioned above, both ICD and texture-ice isotopic compositions appear to reflect long-term climate changes (Opel et al., 2017; Schwamborn et al., 2006; Dereviagin et al., 2013; Porter et al., 2016) which, likely, were also captured in the *n*-alkane or *n*-alkanoic acid δ^2H signal. Unfortunately, we do not have ^{14}C-ages available for all ICD samples, so cross-referencing to published stratigraphies in the region is not possible. Coastal sediments, however, will represent a mixture of material released from different depths, outcrops, and stratigraphies within the catchment or coast. For source-apportionment applications, we reason that a growing body of leaf wax δ^2H end-member data from the ICD region can overcome the variability issues highlighted above.

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4.3 Comparison with ^{13}C-^{14}C source-apportionment: a case-study

Bulk OC dual-carbon isotope data provide a quantitative apportionment tool to assess the relative contributions of Topsoil-PF vs. ICD-PF. Here, we present a case-study of a shelf-slope transect in the Laptev Sea (Fig. 1) where both these source-apportionment tools for the first time can be applied, compared and evaluated. The shelf-slope transect of eight surface sediment samples stretches over 600 km from the nearshore zone (72.7°N, <10m water depth) to the continental rise (78.9°N, >3000m depth) (Table 6). More molecular and bulk geochemical characteristics of these samples can be found in Bröder et al. (2016b).

The $\delta^{13}C$-$\Delta^{14}C$ source-apportionment uses three end-members (marine, Topsoil-PF, and ICD-PF). End-member values are based on previously published values (Tesi et al., 2016); with a $\delta^{13}C$ value of -27.0±1.2‰ (n=38; Rodionow et al., 2006; Tesi et al., 2014; Gundelwein et al., 2007; Bird et al., 2002) for Topsoil-PF, and -26.3±0.67‰ (n=374; Vonk et al., 2012; Schirrmeister et al., 2011) for ICD-PF. The Topsoil-PF $\Delta^{14}C$ endmember was defined as -232±147‰ (n=29; Winterfeld et al., 2015; Jasinski et al., 1998; Kaiser et al., 2007; Höfle et al., 2013; Palmtag et al., 2015). For ICD-PF we used a $\Delta^{14}C$ value of -940±84‰ (n=300; Vonk et al., 2012 and references therein). The marine end-member value was -21.0±2.6‰ (n=10; Panova et al., 2015) and -50.4±12‰ (n=10; Panova et al., 2015) for $\delta^{13}C$ and $\Delta^{14}C$, respectively. Calculations were made using a Markov chain Monte Carlo approach (see 2.3).

For $\delta^{2}H$ source-apportionment there is no need to include a marine end-member as marine organisms do not produce long-chain *n*-alkanes or *n*-alkanoic acids. We were unfortunately only able to analyze *n*-alkanes in the shelf-slope transect samples, and no *n*-alkanoic acids, due to limitations in sample volume. We used the $\delta^{2}H$ values of the C_{27}, C_{29} and C_{31} *n*-alkanes, individually. In other words, these three chain lengths are taken as independent markers, providing an overdetermined system (i.e. two sources defined with three different markers). This is more representative than using the average (concentration-weighted) $\delta^{2}H$ value for these *n*-alkanes as the end-member values for each chain length are different. For Topsoil-PF we used -215±39‰, -246±13‰, and -247±23‰ for C_{27}, C_{29} and C_{31} *n*-alkanes, and for ICD-PF we applied -259±18‰, -297±15‰, and -282±13‰ for C_{27}, C_{29} and C_{31} *n*-alkanes, respectively (see also Table 5). Afterwards, we averaged the three end-member contributions derived from the three calculations for each station, thereby taking the variability introduced by the end-members into account.

The source apportionment of OC from Topsoil-PF and ICD-PF to surface sediments along the Laptev Sea transect differ between the bulk $\delta^{13}C$-$\Delta^{14}C$ and leaf wax $\delta^{2}H$ approaches (Table 6). The former approach suggests Topsoil-PF contributions between 21-70%, generally decreasing offshore, and, consequently, ICD-PF contributions of 30-79%, generally increasing offshore. The latter (leaf wax $\delta^{2}H$) approach results in a more extreme division of sources with Topsoil-PF contributions of 83-91% and ICD-PF contributions of 9-17%, with similar patterns nearshore and offshore (Table 6). A contribution of 9-17% may seem more in line with the estimated extent of ICD in the Lena River basin: 12% of the basin falls within the Yedoma Region (as defined by Romanovsky, 1993) and about 3% consists of intact ICD (see section 2.3). However, the cross-shelf sites are also strongly influenced by coastal and/or subsea erosion (Karlsson et al. 2011; Vonk et al., 2012; Semiletov et al., 2012; 2016) so the catchment characteristics are only one part of the story. It is challenging to interpret the differences between the two proxies but we elaborate below on potential reasons.

Assumptions in the bulk $\delta^{13}C$-$\Delta^{14}C$ approach may affect these results. First, the outcome of the bulk $\delta^{13}C$-$\Delta^{14}C$ approach is sensitive to the definition of the marine

end-member. Changes in the currently used $\delta^{13}C$ and $\Delta^{14}C$ value of the marine end-member of the East Siberian Arctic Shelf (n=10; Panova et al., 2015) would likely alter the relative Topsoil-PF and ICD-PF contributions. The currently used standard deviation for the $\delta^{13}C$ marine end-member is 2.6‰, which is much higher than the values for the terrestrial end-members. Second, lateral transport time enroute the shelf-slope transect (>600 kilometers) causing potentially significant aging of sediments and its organic carbon is not accounted for in the source-apportionment. Lateral transport time results in older surface OC ages on the shelf, compared to those at the initial coastal deposition. Without correcting for this factor, the source-apportionment will generate lower contributions of the (younger) Topsoil-PF component. In an attempt to estimate this effect, we recalculated (similar to Bröder et al. 2016a) the relative source contributions of Topsoil-PF, ICD-PF (and marine) with the bulk $\delta^{13}C$-$\Delta^{14}C$ approach with the assumption that the Topsoil-PF ^{14}C age would be subject to a cross-shelf lateral transport time of 5000 yrs. We assumed a linear aging along the transect based on distance from the coast, with a maximum value of 5000 yrs aging at station SW-01. This resulted in Topsoil-PF contributions that were up to 20% higher (for the deepest stations) compared to the source-apportionment where lateral transport time was unaccounted for (Table 6; Fig. 6).

Assumptions in the leaf wax $\delta^{2}H$ source-apportionment approach could potentially also impact the outcomes, and hence differences with the bulk $\delta^{13}C$-$\Delta^{14}C$ results. First, there is an inherent assumption related to the molecular to bulk level upscaling challenge. We assume that the physical association of *n*-alkanes in different source end-members (Topsoil-PF vs. ICD-PF) as well as their fractionation in the coastal system is similar. However, previous research has shown that *n*-alkanes behave rather differently upon their release into coastal waters; *n*-alkanes originating from surface soil or vegetation debris are not bound to minerals and remain in suspension during transport while being actively degraded, whereas *n*-alkanes originating in deeper mineral soils settle quickly and are protected from extensive degradation (Vonk et al., 2010). It is possible that most of the *n*-alkanes in the Laptev Sea sediment transect originate in (deeper) mineral soils. An effect of physical association, as well as the potential effect of hydrodynamic sorting patterns (Tesi et al., 2016) on the leaf wax $\delta^{2}H$ values of both sources could impact the source-apportionment. Another factor that can introduce a bias in our leaf wax $\delta^{2}H$ approach is a proton exchange of the C-bound H-atoms in *n*-alkanes with environmental water, either from in situ sources (soil water) or during transport (river or ocean water, or sediment pore water). As there is no evidence for such exchange in young (<1 million years), cold sediments (Sessions et al., 2004) we suspect this process may be minimal in our samples (and end-members).

When accounting for an estimated lateral transport time, the difference in estimates of source contribution by the two different approaches (bulk $\delta^{13}C$-$\Delta^{14}C$ and leaf wax $\delta^{2}H$) increases offshore, from about a 25% difference near the coast to a 40% difference at stations SW-01 and SW-03. This increasing offset between the results of the two end-member mixing methods may be caused by several factors such as

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variability in the marine end-member (e.g. due to changes in seasonal ice cover), a selective degradation (of the topsoil OC) enroute that introduces a source bias or isotopic fractionation, or remaining factors related to the lateral transport time (incorrect assumption of 5000 years, non-linear aging along transect). These differences highlight that both source-apportionment tools still could be fine-tuned further by (i) increasing the sample size of sources to reduce end-member uncertainties, (ii) continuous adjustments in end-member values and Markov chain Monte Carlo calculations based on latest knowledge, and (iii) assuring regional testing and verification of the method when applied to new environments.

5 Conclusions

Leaf wax δ^2H values in samples from aquatic recipient environments can be used to source-apportion the incoming terrestrial OC into two end-members; a Pleistocene ICD permafrost source and a younger, Holocene, topsoil source. Mean isotopic values of the C_{29} *n*-alkane, C_{31} *n*-alkane, and C_{28} *n*-alkanoic acid showed a dynamic, statistically significant range of 34, 50 and 46‰ between Topsoil-PF and ICD-PF samples, respectively, with ICD-PF samples being consistently more depleted indicative of formation during the colder and drier Pleistocene.

A case-study where we tested two isotopic proxies (leaf wax δ^2H and bulk $\delta^{13}C$-$\Delta^{14}C$) to calculate the relative terrestrial source contribution of Topsoil-PF and ICD-PF along a Laptev Sea surface sediment transect, showed that the two proxies yield variable results but overall generate similar trends offshore. We reason that variability is caused by factors such as lateral transport time, remaining uncertainties in end-member definition, or environmental factors such as physical association.

Both methods (leaf wax δ^2H and bulk $\delta^{13}C$-$\Delta^{14}C$) bring along their inherent disadvantages and advantages. The molecular approach has the distinct advantage that it circumvents the uncertainties that are associated with marine end-member definition in the case of bulk OC mixing model analysis. However, application of molecular δ^2H in source-apportionment studies brings along challenges related to the molecular-bulk upscaling step. Bulk $\delta^{13}C$-$\Delta^{14}C$ source-apportionment, on the other hand, has the advantage to operate on a bulk and perhaps more representative level, but is hampered by remaining uncertainties associated with the marine end-member.

This study shows that δ^2H of leaf wax molecules has the potential to be used in quantitative source-apportionment studies of thawing permafrost in coastal or marine settings. It can serve as an alternative or complementary approach to the commonly applied bulk $\delta^{13}C$-$\Delta^{14}C$ method. We recommend continuing data collection and optimization of end-member definition and calibration. Refining the molecular δ^2H proxy presented here will be beneficial in pinpointing the location and extent of OC release from thawing permafrost in the coastal or fluvial environment. With enhanced Arctic warming and associated intensification of permafrost thaw, constraining the amount and fate of permafrost OC release will help to assess the magnitude of the permafrost carbon feedback to climate warming.

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Data availability

All data are available in Tables 1 through 6, as well as Supplementary Table S1.

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Author contributions

Land-based samples were collected by GH and JEV, ship-based samples were collected by IS, OD, ÖG, TT, LB, and JEV. Laboratory analysis was performed by LB, TT, and HH. Markov chain Monte Carlo simulations were run by AA. The manuscript was written by JEV with input of all co-authors.

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Figure 1

Map of coastal northeast Siberia showing the extent of ice complex permafrost (ICD; red) overlaid with the location of ice complex (n=9; black diamonds) and topsoil samples (n=9; green squares). The shelf-slope Laptev Sea transect is shown with yellow stars.

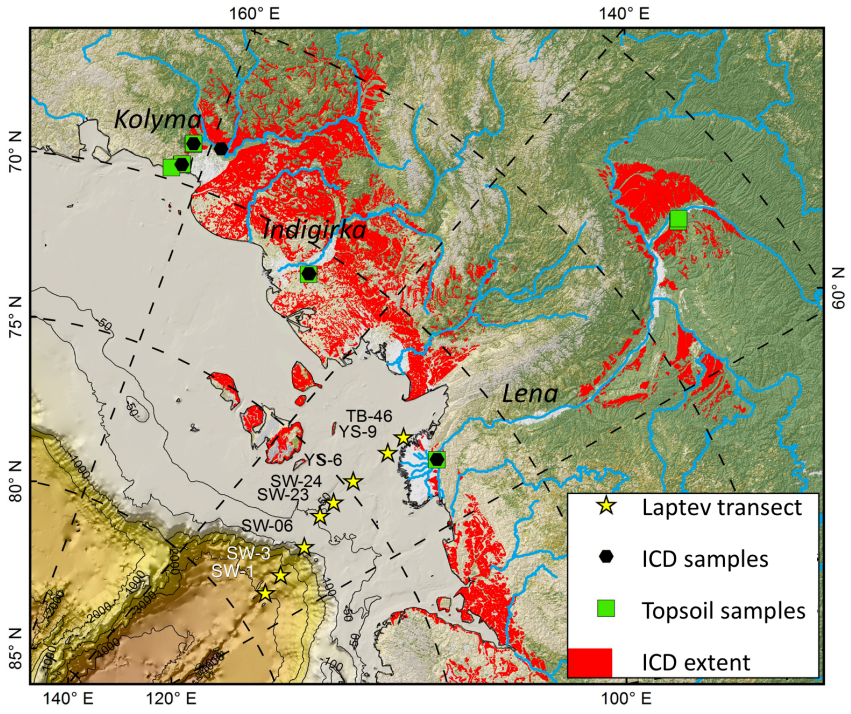

Figure 2

Molecular concentrations and ratios of topsoil Holocene permafrost (green; n=9) and deeper Pleistocene permafrost (blue; n=9) samples, with (a) the sum of odd *n*-alkanes C_{25}-C_{31}, (b) the sum of even *n*-alkanoic acids C_{22}-C_{28}, (C) the Carbon Preference Index (CPI) for *n*-alkanes C_{23}-C_{31}, (d), the CPI for *n*-alkanoic acids C_{22}-C_{28}, (e) the ratio of C_{23} over C_{23}+C_{29} *n*-alkanes, and (f) the sum of high-molecular weight (HMW) *n*-alkanoic acids over HMW *n*-alkanes. The CPI is calculated as $CPI_{i\text{-}n} = ½ \Sigma (X_i+X_{i+2}+...+X_n)/ \Sigma (X_{i-1}+X_{i+1}+...+X_{n-1}) + ½ \Sigma (X_i+X_{i+2}+...+X_n)/ \Sigma (X_{i+1}+X_{i+3}+...+X_{n+1})$, where X is concentration. Stars indicate that the two compared values are statistically significant (95% confidence). Note that panel a and b are reported as median with IQR (interquartile range) and the other panels are reported as average±standard deviation.

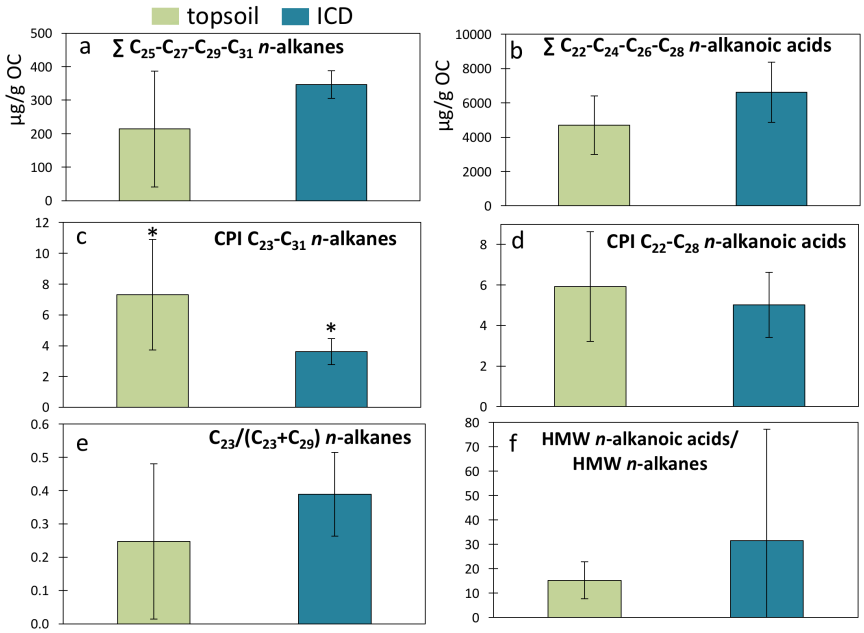

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Figure 3

Molecular isotopic signature against chain length of long chain *n*-alkanoic acids (top) and *n*-alkanes (bottom) for Holocene topsoil samples (green) and Pleistocene ice complex samples (ICD; blue). Stars indicate that the two compared values are statistically significant (95% confidence). Standard deviations are represented as vertical bars, and are smaller than the sample circles when not visible.

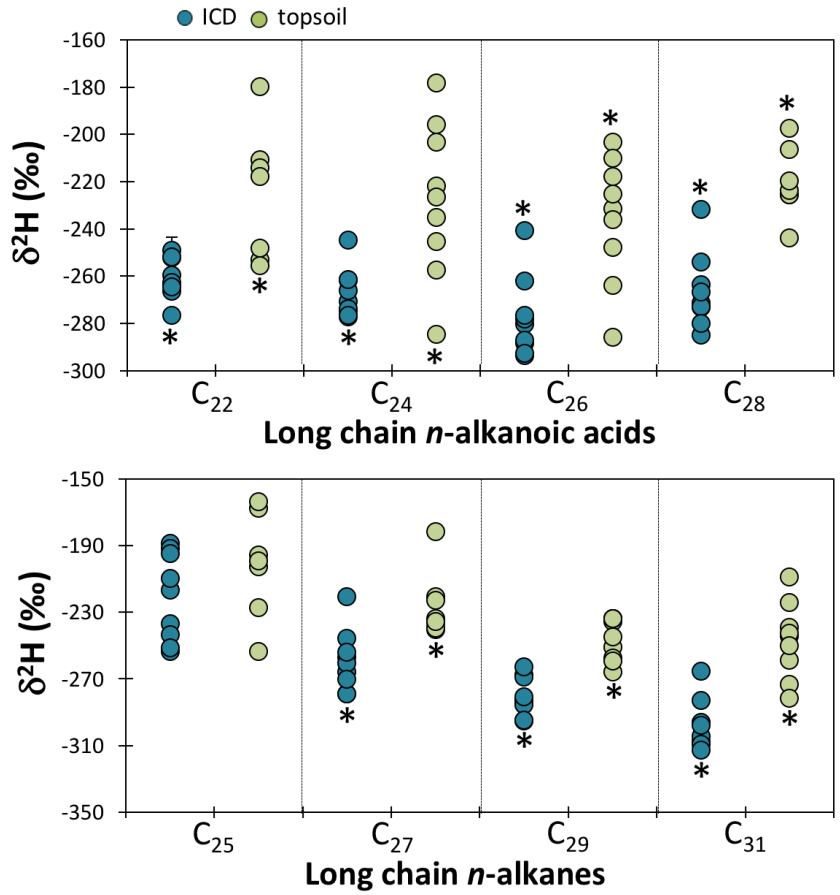

Figure 4

Concentration-weighted mean δ^2H values of C_{27}-C_{29}-C_{31} *n*-alkanes plotted against concentration-weighted mean δ^2H values of C_{24}-C_{26}-C_{28} *n*-alkanoic acids to illustrate the fractionation differences between these two leaf wax markers. Dashed line indicates an identical fractionation.

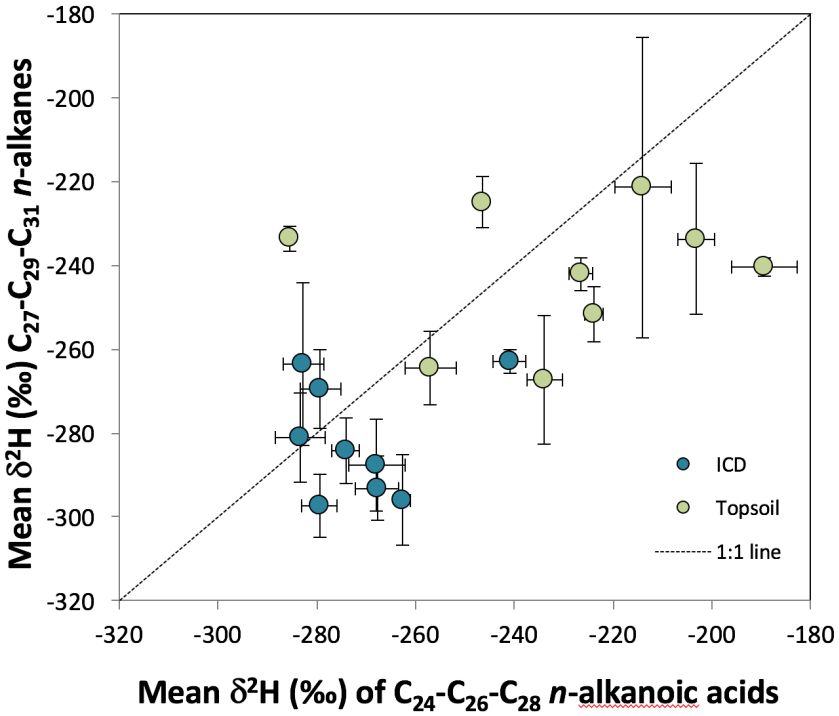

Figure 5

Comparison of δ^2H values of C_{28} *n*-alkanoic acid (left) and C_{29} *n*-alkane (right) in modern (Topsoil-PF; green circles) and ICD-PF for this study (blue circles) and available literature, with crosses from Zech et al. (2011; glacial and interglacial paleosoils from permafrost bluff exposure at Tumara River northeast Siberia), black triangles from Yang et al. (2011; C3 plants and trees from Canada and Alaska), light grey triangles from Wilkie et al. (2013; C3 plants from the El'gygytgyn lake basin, Siberia), white triangles from Pautler et al. (2014; modern and paleosoils from the Yukon territory, Canada) and dark grey triangles from Porter et al. (2016; muck deposits from the Yukon territory, Canada).

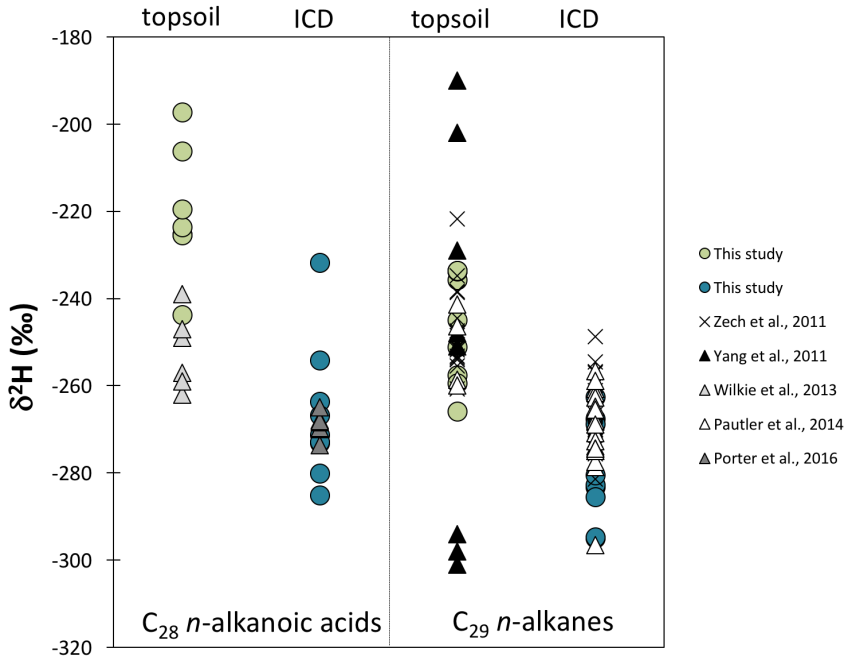

Figure 6

Contribution of OC from Topsoil-PF (green) and ICD-PF (blue) sources to surface sediments along a shelf-slope transect in the Laptev Sea (see also Bröder et al., 2016b for further transect information), calculated with a $\delta^{13}C$- $\Delta^{14}C$ (triangles) and leaf wax $\delta^{2}H$ mixing model (circles). Stations are plotted against log water depth (m; see also Table 6) following the transect order from the coastal, nearshore, zone in the South (furthest left; TB-46, 6 m depth) towards the continental rise in the North (furthest right; SW-01, 3146 m depth). Topsoil $\Delta^{14}C$ end-member values are corrected for cross-shelf transport time (see section 4.2).

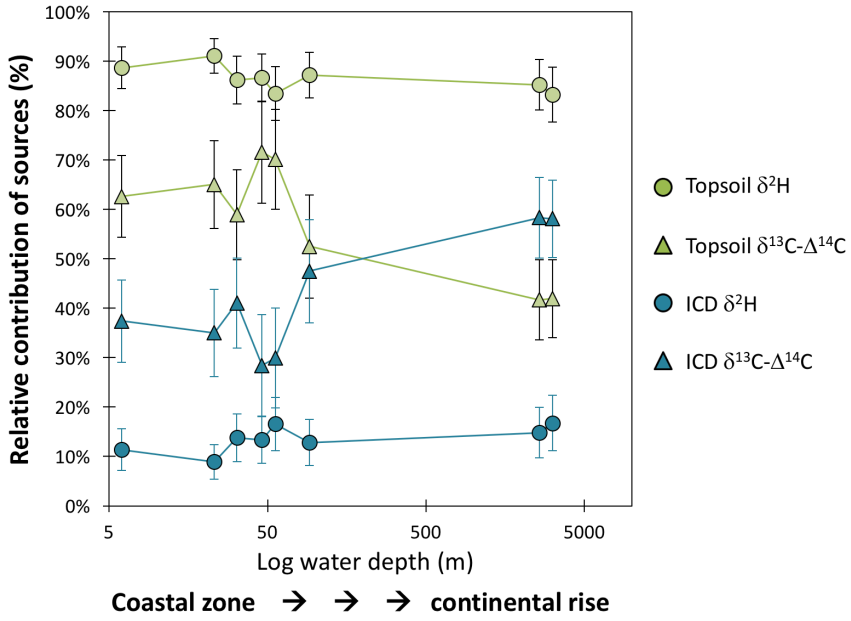

Table 1

Site characteristics and geochemical properties of eight topsoil and eight ice complex deposit samples. A table with more detailed sample descriptions can be found in Supplementary Table 1. n.a. indicates "not available".

Sample code	Sample ID	Current vegetation	Watershed	Description	Lat	Lon	TOC	$\delta^{13}C$	^{14}C-yrs		
					°N	°E	%	‰		%	
Topsoil (modern vegetation and O-horizon samples)											
TS-1	KU EXP 1-1, 0-16 cm	Tundra	Lena	Surface O-horizon; 0-16 cm	72.34	126.29	11	-27.0	n.a.	0.40	27.5
TS-2	CH YED2, 0-4 cm	Tundra	Kolyma	Surface O-A horizon; 0-4 cm	69.46	161.79	17	-28.4	n.a.	0.64	26.5
TS-3	SP T3-3B,	Alas grassland	Lena	Alas soil (Mollisol), mix of O and A horizon	62.32	129.50	15	-27.9	n.a.	1.40	10.7
TS-4	SP T2-7,	Larch taiga	Lena	Taiga soil (turbel), mix of O and A horizon	62.25	129.62	13	-28.4	n.a.	0.45	28.0
TS-5	KY T2-3,	Tussock tundra	Indigirka	Tundra soil (turbel), O-horizon	70.83	147.48	29	-28.5	n.a.	1.56	18.7
TS-6	CH T2-1,	Tussock tundra	Kolyma	Tundra soil (turbel), mix of O, Ojj and Ajj horizons	69.44	161.77	21	-26.4	n.a.	0.57	36.7
TS-7	CH YED3, 0-10 cm	Larch taiga	Kolyma	Surface O-hor; 0-10 cm	68.77	161.41	39	-29.6	n.a.	1.29	30.7
TS-8G[a]	CH Medv grass[a]	Grass tundra	Kolyma	Vegetation	69.64	162.54	41	-25.2	n.a.		
TS-9G[a]	CH Y4 grass[a]	Larch taiga	Kolyma	Vegetation	68.74	161.41	40	-28.5	n.a.		
	Mean values						**25**	**-27.8**		**1.1**	**24.8**
	Ice complex deposits										
ICD-1	KU EXP 1-3, 212-216 cm	Tundra	Lena	Very deep undisturbed yedoma ca. 10 m below surface	72.34	126.29	1.3	-27.5	n.a.	0.08	15.7
ICD-2	CH YED1, 300-305 cm	Tussock tundra	Kolyma	Deep undisturbed yedoma ca. 3 m below surface	69.47	161.77	1.4	-26.3	n.a.	0.14	10.2
ICD-3	CH YED2, 300-305 cm	Tussock tundra	Kolyma	Deep undisturbed yedoma ca. 3 m below surface	69.46	161.79	2.3	-25.8	n.a.	0.27	8.6
ICD-4	CH YED3, 520-525 cm	Larch taiga	Kolyma	Deep undisturbed yedoma ca. 5 m below surface	68.77	161.41	1.4	-25.5	n.a.	0.15	9.7
ICD-5	KY EXP1, 0-5 cm	Tussock tundra	Indigirka	Undisturbed yedoma ca. 2 m below surface	70.83	147.44	1.5	-25.5	27920 ±210	0.18	8.5

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ICD-6	KY EXP2, 110-115 cm	Tussock tundra	Indigirka	Deep undisturbed yedoma ca. 4.5 m below surface	70.83	147.44	1.6	-25.6	17270 ±80	0.19	8.6
ICD-7	KY EXP3, 185-190 cm	Tussock tundra	Indigirka	Undisturbed yedoma ca. 2 m below surface	70.83	147.49	1.5	-25.2	n.a.	0.17	8.5
ICD-8	CH DY-3A	Larch taiga	Kolyma	Particulate matter from thaw streams	68.63	159.15	1.5[b]	-25.2[b]	19370 ±70	-	-
ICD-9	CH DY-4A	Larch taiga	Kolyma	Particulate matter from thaw streams	68.63	159.15	1.4[b]	-25.1[b]	28040 ±140	-	-
	Mean values						**1.6**	**-25.7**		**0.2**	**10.0**

a vegetation/grass samples, labelled with "G"
b data from Vonk et al., 2013

Table 2
Long-chain *n*-alkane concentrations (in µg/gOC) of topsoil Holocene samples (modern vegetation/O-horizon) and Pleistocene ice complex samples.

	C21	C22	C23	C24	C25	C26	C27	C28	C29	C30	C31	C32	C33
	µg/g OC												
Topsoil (modern vegetation and O-horizon samples)													
TS-1	44	88	96	45	41	10	45	4.4	27	2.5	36	1.5	7.2
TS-2	24	15	21	12	40	10	160	10	150	6.5	150	3.5	17
TS-3	2.5	2.4	5.9	2.6	13	4.7	42	16	74	4.7	85	2.7	24
TS-4	19	3.3	7.1	2.7	27	4.5	47	6.7	98	9.1	150	5.7	38
TS-5	35	8.4	26	9.9	38	13	91	18	180	14	230	8.1	43
TS-6	14	5.1	16	5.7	19	4.0	26	3.7	48	5.0	120	4.0	32
TS-7	46	12	18	8.8	22	16	61	27	220	23	340	12	48
TS-8G	4.1	1.7	18	10	61	16	47	13	30	5.3	10	1.1	1.1
TS-9G	4.7	2.6	18	15	45	21	50	16	31	6.8	9.8	1.5	2.6
Ice complex deposits													
ICD-1	57	79	100	49	82	23	170	16	137	8.5	140	4.4	25
ICD-2	55	89	100	70	70	27	75	20	130	12	120	5.3	28
ICD-3	40	64	74	31	54	15	79	22	110	10	160	4.8	32
ICD-4	60	93	98	47	55	20	84	22	140	12	150	6.0	39
ICD-5	46	79	86	56	49	20	55	13	75	7.0	100	4.7	38
ICD-6	41	73	87	68	62	29	65	20	98	11	120	4.9	27
ICD-7	50	83	83	43	41	16	65	17	100	8.3	120	4.5	42
ICD-8	4.2	7.3	23	30	55	42	82	38	100	18	110	5.0	21
ICD-9	6.2	6.2	16	11	29	15	51	20	79	9.3	85	4.1	23

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Table 3
Long-chain *n*-alkanoic acids concentrations (in µg/gOC) of topsoil Holocene samples (modern vegetation/O-horizon) and Pleistocene ice complex samples.

	C16	C18	C20	C21	C22	C23	C24	C25	C26	C27	C28	C29	C30
	µg/gOC												
Topsoil (modern vegetation and O-horizon samples)													
TS-1	511	220	176	80.5	539	311	1100	4.95	684	90.5	350	32.8	58.1
TS-2	1740	664	673	235	1380	496	1390	543	1740	409	1580	113	305
TS-3	664	296	480	116	1020	504	1710	415	1550	250	1060	132	456
TS-4	1140	408	665	235	1400	431	1410	425	1250	242	651	143	455
TS-5	513	343	530	133	1140	359	1410	1.58	896	119	494	67.8	224
TS-6	1080	537	418	236	1420	790	2670	2.82	1570	127	657	46.6	174
TS-7	1420	352	538	281	1850	722	2010	651	1790	642	1580	730	1971
TS-8G	3640	855	691	44.1	609	63.5	156	26.0	224	0.122	99.3	9.91	28.1
TS-9G	4600	887	966	53.6	815	66.7	261	28.6	232	11.5	124	8.10	30.2
Ice complex deposits													
ICD-1	1750	1600	4560	1460	9460	2300	8930	2020	5830	1030	3660	293	635
ICD-2	10400	4030	5800	2410	17100	7270	18600	6610	16600	5860	14800	6810	18700
ICD-3	665	554	892	263	2070	1060	3070	646	2340	272	1310	133	532
ICD-4	1400	769	1030	252	2040	910	3120	644	2440	266	1160	124	432
ICD-5	426	304	447	126	1220	511	1970	70.4	1390	133	712	60.7	233
ICD-6	722	539	583	153	1370	606	2270	457	1970	181	1030	86.4	333
ICD-7	446	313	543	158	1330	562	2350	401	1370	154	743	63.1	230
ICD-8	920	402	895	108	1070	294	1180	184	799	70.3	331	34.4	100
ICD-9	327	200	559	74	803	229	1010	2.17	718	64.9	334	28.7	104

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Table 4

Sum of most abundant long-chain *n*-alkanoic acids and *n*-alkanes (concentrations in µg/gOC), and characteristic ratios of *n*-alkanoic acids and *n*-alkanes of topsoil Holocene (modern vegetation/O-horizon) and Pleistocene ice complex samples.

	***n*-alkanoic acids**				***n*-alkanes**				
	ΣHMW[a] (>C_{22}) *µg/gOC*	**ΣC_{22}-C_{28} (even)** *µg/gOC*	**CPI[b]**	**HMW acids/ HMW alkanes[a]**	**ΣHMW[a] (>C_{21})** *µg/gOC*	**ΣC_{25}-C_{31} (odd)** *µg/gOC*	**CPI [c]**	**C_{23}/ (C_{23}+C_{23})**	**C_{25}/ (C_{25}+C_{29})**
Topsoil (modern vegetation and O-horizon samples)									
TS-1	3167	2670	5.8	7.1	447	148	2.7	0.78	0.60
TS-2	7958	6090	3.8	13	612	494	11	0.12	0.21
TS-3	7095	5340	4.1	25	280	214	7.2	0.07	0.15
TS-4	6397	4700	3.7	15	418	323	12	0.07	0.24
TS-5	4715	3940	6.8	6.6	717	543	9.1	0.12	0.17
TS-6	7454	6310	6.0	25	300	211	9.9	0.25	0.28
TS-7	11950	7230	2.9	14	857	647	7.8	0.08	0.09
TS-8G	1216	1090	9.5	5.6	217	148	3.7	0.37	0.67
TS-9G	1577	1430	11	7.1	223	135	2.5	0.36	0.59
Mean±stdev	*5726±3431*	*4310±2190*	*5.9*	*13*	*452±230*	*318±195*	*7.3*	*0.25*	*0.33*
Median and IQR	6397^{7454}_{3167}	4700^{2670}_{6092}	*2.7*	*7.6*	418^{621}_{280}	214^{494}_{148}	*3.6*	*0.23*	*0.22*
Ice complex deposits									
ICD-1	34854	27883	4.1	39	893	530	4.9	0.43	0.38
ICD-2	112356	67078	2.8	140	806	398	3.0	0.44	0.35
ICD-3	11430	8791	4.1	16	698	405	4.6	0.40	0.33
ICD-4	11145	8768	4.4	14	825	428	3.8	0.42	0.29
ICD-5	6293	5285	6.5	10	630	280	2.9	0.54	0.40
ICD-6	8293	6629	4.9	12	708	347	2.7	0.47	0.39
ICD-7	7196	5787	4.7	11	671	323	3.5	0.45	0.29
ICD-8	4063	3380	5.5	7.6	533	344	2.7	0.19	0.35
ICD-9	3295	2867	8.3	9.3	355	244	4.3	0.17	0.27
Mean±stdev	*22103±35150*	*15160±20800*	*5.0*	*29*	*680±163*	*367±85*	*3.6*	*0.39*	*0.34*
Median and IQR	8290^{11430}_{6290}	6630^{8790}_{5285}	*1.6*	*43*	698^{806}_{630}	347^{405}_{323}	*0.8*	*0.13*	*0.05*

a HMW; high-molecular weight

b CPI; carbon preference index for chain lengths C_{22}-C_{28}, for calculation see caption of Fig. 2.

c CPI; carbon preference index for chain lengths C_{23}-C_{31}, for calculation see caption of Fig. 2.

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Table 5

δ^2H signatures (in ‰) of *n*-alkanoic acids and *n*-alkanes of topsoil Holocene (modern vegetation/O-horizon) and Pleistocene ice complex samples.

	***n*-alkanoic acids**							***n*-alkanes**			
	C16	**C18**	**C20**	**C22**	**C24**	**C26**	**C28**	**C25**	**C27**	**C29**	**C31**
Topsoil (modern vegetation and O-horizon samples)											
TS-1	-162	-180		-119	-178	-203	-197	-168	-240	-236	-244
TS-2	-188	-192		-211	-222	-232	-225	-196	-237	-251	-239
TS-3				-126	-203	-218	-225		-125	-234	-259
TS-4	-171	-213		-180	-196	-210	-206		-182	-245	-243
TS-5		-235	-185	-253	-257	-264	-244	-164	-240	-266	-273
TS-6	-189	-222		-214	-235	-236	-224	-203	-221	-258	-282
TS-7	-184		-190	-218	-227	-225	-220	-199	-234	-259	-250
TS-8G	-258	-246	-253	-256	-285	-286		-253	-236	-234	-224
TS-9G	-237	-244	-251	-248	-245	-248		-227	-223	-234	-209
Mean	*-199*	*-219*	*-220*	*-203*	*-228*	*-236*	*-220*	*-201*	*-215*	*-246*	*-247*
St.dev	35	25	37	52	33	27	15	*32*	*39*	*13*	*23*
Ice complex deposits											
ICD-1	-194	-227	-243	-252	-245	-241	-232	-237	-257	-268	-265
ICD-2			-231	-264	-271	-280	-271	-217	-266	-283	-297
ICD-3				-249	-262	-278	-264	-254	-279	-283	-307
ICD-4			-209	-252	-266	-277	-254	-243	-261	-285	-305
ICD-5			-169	-260	-275	-288	-273	-189	-245	-269	-283
ICD-6	-211	-216	-252	-266	-274	-294	-285	-192	-254	-281	-296
ICD-7			-191	-263	-277	-287	-273	-210	-279	-295	-309
ICD-8	-244	-256	-277	-277	-277	-293	-280	-195	-221	-263	-298
ICD-9	-228	-229	-261	-265	-262	-262	-267	-251	-270	-295	-313
Mean	*-219*	*-232*	*-229*	*-261*	*-268*	*-278*	*-267*	*-221*	*-259*	*-280*	*-297*
St.dev	21	17	37	8.6	10	17	16	*26*	*18*	*12*	*15*

Deleted: KU EXP 1-1, 0-16 cm

Deleted: CH YED2, 0-4 cm

Deleted: SP T3-3B

Deleted: SP T2-7

Deleted: KY T2-3

Deleted: CH T2-1

Deleted: CH YED3, 0-10 cm

Deleted: CH Medv grass

Deleted: CH Y4 grass

Deleted: KU EXP 1-3, 212-216 cm

Deleted: CH YED1, 300-305 cm

Deleted: CH YED2, 300-305 cm

Deleted: CH YED3, 520-525 cm

Deleted: KY EXP1, 0-5 cm

Deleted: KY EXP2, 110-115 cm

Deleted: KY EXP3, 185-200 cm

Deleted: CH DY-3A

Deleted: CH DY-4A

Table 6

Location, sampling depth and isotopic values of samples along a surface sediment transect in the Laptev Sea (data from Bröder et al., 2016b), with percentage topsoil (TS) and ice complex deposit (ICD) OC contributions to the samples based on source-apportionment calculations with δ^2H leaf wax end-members versus $\delta^{13}C$-$\Delta^{14}C$ end-members (end-member values are described in the text).

				Sample values						**Source contributions**			
ID[a]	**Lat**	**Long**	**Depth**	**C_{27}**	**C_{29}**	**C_{31}**	**$C_{27\text{-}29\text{-}31}$**[b]	**$\delta^{13}C$**	**$\Delta^{14}C$**	**TS**	**ICD**	**TS**[c]	**ICD**[c]
	N	°E	m	‰	‰	‰	‰	‰	‰	using δ^2H		using $\delta^{13}C$-$\Delta^{14}C$	
TB-46	72.700	130.180	6	-236.2	-237.4	-230.4	-235.0	-26.5	-436	89%	11%	63% (63%)	37% (37%)
YS-9	73.366	129.997	23	-233.7	-231.0	-227.8	-231.1	-26.1	-415	91%	8.9%	63% (65%)	37% (35%)
YS-6	74.724	130.016	32	-234.2	-241.0	-235.4	-236.8	-25.6	-465	86%	14%	51% (59%)	49% (41%)
SW-24	75.599	129.558	46	-229.3	-236.5	-243.5	-236.4	-24.8	-284	87%	13%	70% (72%)	30% (28%)
SW-23	76.171	129.333	56	-219.9	-243.3	-243.3	-236.0	-25.0	-333	83%	17%	65% (70%)	35% (30%)
SW-06	77.142	127.378	92	-219.5	-237.0	-241.4	-233.2	-23.2	-364	87%	13%	39% (53%)	61% (47%)
SW-03	78.238	126.150	2601	-221.1	-238.0	-247.7	-235.9	-22.6	-426	85%	15%	23% (42%)	77% (58%)
SW-01	78.942	125.243	3146	-223.8	-241.8	-246.0	-238.0	-22.3	-418	83%	17%	21% (42%)	79% (58%)

a Location, depth and bulk carbon isotope data from Bröder et al. (2016b)

b weighted average based on individual concentrations

c numbers in brackets are source contributions using the $\delta^{13}C$-$\Delta^{14}C$ approach but with additional corrections for cross-shelf lateral transport time of topsoil OC (similar as in Bröder et al., 2016a); we applied linear aging along the transect based on the distance from the coast, with a maximum aging of 5000 years for station SW-01.

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Mean (median)	*5726 (6397)*	*4310 (4700)*	*5.9*	*13*	*452 (418)*
St.dev (IQR)	*3431 (4290)*	*2190 (4320)*	*2.7*	*7.6*	*230 (332)*

Page 32: [2] Deleted	Jorien	6/7/17 8:36:00 PM				
Mean	*22103 (8290)*	*15160 (6630)*	*5.0*	*29*	*680 (698)*	*367 (347)*
St.dev	*35150(5140)*	*20880 (3510)*	*1.6*	*43*	*163 (176)*	*85 (81)*